



## Novel Bioactive Compounds in Methanolic Extract of *Andrographis paniculata* through GC-MS

Pragya Shrivastava<sup>1</sup>, Kush Kumar Nayak<sup>2</sup>, Zenu Jha<sup>3</sup>, Varaprasad Kolla<sup>1\*</sup>

<sup>1</sup>Amity Institute of Biotechnology, Amity University Chhattisgarh, Raipur – 493225, India

<sup>2</sup>School of Studies in Biotechnology, Shaheed Mahendra Karma Vishwavidyalaya, Baster, Jagdalpur Chhattisgarh-494001 India

<sup>3</sup>Department of Plant Molecular Biology and Biotechnology, Indira Gandhi Krishi Vishwavidyalaya, Raipur, Chhattisgarh - 493225, India

(Received: 16 March 2025

Revised: 20 April 2025

Accepted: 15 June 2025)

### KEYWORDS

*Andrographis paniculata*, gas chromatography–mass spectrometry (GC MS), methanolic extract, phytoconstituents, novel compound.

### ABSTRACT:

#### Introduction

Medicinal plants have garnered significant attention in biotechnology research due to their pivotal role in synthesizing bioactive compounds essential for various pharmaceutical applications. Historically, these plants have been primary sources of numerous biologically active substances, including therapeutic agents, natural dyes, fragrances, and flavour enhancers, especially in India. Herbal remedies often derive from raw plant extracts rich in diverse phytochemicals, predominantly secondary metabolites, which can vary considerably among different plant species. *Andrographis paniculata* is a well established medicinal herb known for its anti inflammatory, antiviral, and hepatoprotective properties. Despite extensive study of its major diterpenoids, andrographolide, many minor phytochemicals remain unexplored.

#### Objective

To uncover and profile previously unreported phytoconstituents in *A. paniculata* leaf extract using gas chromatography–mass spectrometry (GC MS).

#### Methods

Botanical identity of *A. paniculata* was validated via morphological and taxonomic criteria. Air dried leaves were pulverized and subjected to methanol extraction by Soxhlet and the concentrated extract underwent. The GC-MS evaluation was performed following standard procedures with a Mass Hunter GC/MS Acquisition B.07.05.2479 system (Agilent Technologies, Inc.).

#### Result

GC MS profiling identified 14 phytoconstituents, of which one of compounds Dodecanedioic has not been previously reported in *A. paniculata* based on library match quality and fragmentation patterns.

#### Conclusion

This study expands the phytochemical repertoire of *A. paniculata* by reporting one newly compound and fourteen, several compounds of which as fatty acid methyl esters and phenolic ethers contribute to its pharmacological actions. These findings support further bioactivity assays (antioxidant, cytotoxicity, anti inflammatory) to evaluate therapeutic potential and may inform isolation and structure activity relationship studies.



## 1. Introduction

*Andrographis paniculata* commonly known as "Kalmegh" or the "King of Bitters," is a widely used medicinal herb in traditional healthcare systems as Ayurveda, Siddha, and Traditional Chinese Medicine<sup>8</sup>. Indigenous to India and Southeast Asia, *A. paniculata* has long been employed for the treatment of a broad range of ailments including fever, liver disorders, respiratory infections, and inflammation<sup>1</sup>. The bitter principles in its leaves and aerial parts are associated with its therapeutic potential, making it a staple in herbal preparations and polyherbal formulations<sup>6</sup>. Numerous ethnobotanical reports describe its role in managing gastrointestinal disturbances, skin diseases, and even as a preventive remedy for chronic illnesses. Its longstanding traditional use is supported by a variety of experimental and clinical studies which confirm its bioactivity<sup>9</sup>. However, the specific chemical constituents responsible for these effects have not yet been fully identified or characterized in all extract forms, particularly those obtained through methanolic extraction<sup>5</sup>. The pharmacological value of *A. paniculata* is largely attributed to its rich content of diterpenoids, flavonoids, and polyphenols. Among these, andrographolide a labdane diterpenoid, is the most studied compound known for its anti-inflammatory, anti-viral, antioxidant, anti-cancer, immunomodulatory, and hepatoprotective activities<sup>10</sup>. Other bioactive constituents such as neoandrographolide, andrograpanin, and various flavones also contribute to its medicinal profile<sup>3</sup>. These compounds have shown significant potential in modulating biological pathways related to oxidative stress, immune response, and microbial infection. Despite the wealth of knowledge on these compounds, research is heavily focused on specific extracts or individual molecules, while the broader phytochemical landscape remains underexplored, especially when methanol is used as a solvent for extraction. Gas Chromatography–Mass Spectrometry (GC-MS) is a powerful analytical technique widely used for the separation, identification, and quantification of volatile and semi-volatile organic compounds<sup>4</sup>. While some studies have utilized GC-MS to analyze extracts of *A. paniculata*, they primarily focus on aqueous or ethanolic extractions. The methanolic extract, which often yields a unique and richer spectrum of secondary metabolites, has not been comprehensively studied using GC-MS. As a result, there exists a considerable

knowledge gap in the identification of lesser-known or newly emerging phytoconstituents present in methanol-soluble fractions<sup>2</sup>. This underrepresentation has limited our understanding of the full therapeutic potential and chemical diversity of *A. paniculata*. It is essential to extend phytochemical investigations beyond known compounds and standard extraction techniques to uncover the hidden reservoir of bioactive molecules that may contribute to its pharmacological properties<sup>11</sup>.

**Study Objective and Rationale.** The primary objective of this study is to perform an in-depth GC-MS analysis of the methanolic extract of *Andrographis paniculata* to identify and characterize its phytoconstituents. The rationale behind this research stems from the need to bridge the existing knowledge gap in the chemical profiling of *A. paniculata* and to expand our understanding of its full phytochemical spectrum.

By applying GC-MS to methanol-extracted samples, this study aims to uncover potentially novel or underreported compounds that may have significant biological relevance<sup>7</sup>. Such profiling can not only validate traditional uses with a scientific basis but also provide insights for future pharmacological studies and drug development efforts. Ultimately, the findings will help to map out a more complete phytochemical profile of this important medicinal plant and pave the way for the discovery of new bioactive agents<sup>12</sup>.

## 2. Materials & Methods

### 2.1. Plant sample collection and Extraction of crude extract

Fresh, healthy leaves of *Andrographis paniculata* were collected from the medicinal plant garden of Amity University, Chhattisgarh. The plant material was thoroughly washed, shade-dried, and ground into fine powder. 20 grams of powdered material were subjected to extraction using 200 mL of methanol through Soxhlet apparatus for 8 hours. The extract was filtered, and the solvent was evaporated using a rotary evaporator under reduced pressure. The resulting crude extract was stored at 4 °C until further analysis by GC-MS.

### 2.2. GC-MS Analysis

Gas Chromatography-Mass Spectrometry (GC-MS) analysis was performed using a Mass Hunter GC/MS Acquisition B.07.05.2479 system (Agilent Technologies, Inc.) system or its equivalent. The analytical column used



was an Rxi-5Sil MS capillary column (30 m length  $\times$  0.25 mm internal diameter  $\times$  0.25  $\mu$ m film thickness), coated with 5% diphenyl/95% dimethylpolysiloxane. Helium was employed as the carrier gas at a constant flow rate of 1.0 mL/min. A 1–2  $\mu$ L aliquot of the methanolic extract was injected into the system in split mode at an injector temperature of 260 °C. The oven temperature was initially held at 80 °C for 4 minutes, followed by a linear ramp of 5 °C/min up to 280 °C, where it was held constant for 6 minutes. The ion source operated in electron impact (EI) mode at 70 eV. The mass detector scanned a mass range of 50 to 500 amu.

### 2.3. Data Analysis and Compound Identification

Chromatograms were recorded and peaks were analysed based on retention times as per second and corresponding mass spectra. Identification of compounds was carried out by comparing the acquired spectra with reference spectra available in NIST and Wiley libraries. Quantitative data were derived by calculating the relative percentage of each compound based on its peak area in the total ion chromatogram (TIC). Where applicable, retention indices were computed to aid in compound confirmation.

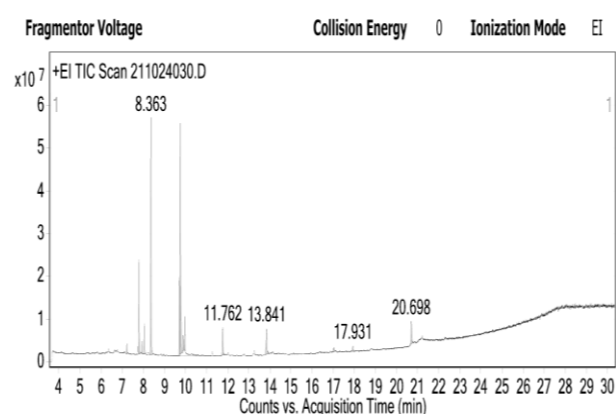
## 3. Results and Discussion

### Compound Identification and GC-MS Chromatogram

The methanolic extract of the plant sample was subjected to compound profiling to identify the phytochemical constituents. The mentioned in table 1 of GC-MS analysis of the methanolic extract revealed the presence of multiple fourteen bioactive compounds, identified based on their retention times and mass spectral data. The chromatogram displayed in figure 1 shows five highest distinct peaks, indicating a complex mixture of phytochemicals within the extract. Among these five identified compounds, the major constituents were observed at retention times corresponding to significant peak areas. The most prominent compounds included Methyl 8-methyl-nonanoate; Dodecanedioic acid; 9,12-Octadecadienoyl chloride, (Z, Z); [1-(3,3-Dimethyloxiran-2 ylmethyl)-3,7-dimethylocta 2,6-dienyl] trimethylsilane, and 9,12-Octadecadiynoic acid, trimethylsilyl ester are reported as known for their

pharmacological properties as anti-inflammatory, antioxidant, and antimicrobial activities. Additionally, among of these rest of mentioned table 2 of minor bioactive constituents were identified, which may contribute synergistically to the overall bioactivity of the extract. Notably, a one of the Dodecanedioic acids identified compounds appeared to be either novel and not reported in previous literature from this plant species, indicating the uniqueness of the phytochemical profile under methanol extraction. This Dodecanedioic acid is an alpha, omega-dicarboxylic acid that is dodecane in which the methyl group have been oxidised to the corresponding carboxylic acid. It has a role as an EC 1.1.1.1 (alcohol dehydrogenase) inhibitor and a human metabolite. It is an alpha, omega-dicarboxylic acid and a dicarboxylic fatty acid. It is a conjugate acid of dodecanedioate (2-) it derives from a hydride of a dodecane. Dodecanedioic acid has been reported in *Drosophila melanogaster*, *homo sapiens*, and other organism but not yet in *A. paniculata* confirmed by.

Overall, the mention mass spectrum in figure 2 methanol extract demonstrated a rich chemical profile, suggesting its suitability for further biological screening and potential pharmaceutical applications. These findings suggest that methanol is an effective solvent for extracting a broad spectrum of bioactive compounds, making it suitable for further pharmacological and functional studies.



**Figure 1. GC–MS analysis of high prominent peak of compounds:**



**Table 1.** Fourteen Compounds identified by GC–MS are listed by retention time (RT), peak height, area, % area (relative to total), symmetry factor, and peak width. The table shows RT (min), height (counts), area (counts), normalized area (%), symmetry (front/rear area ratio), and width (area/height). Data are arranged in ascending RT to aid clear visual comparison of chromatographic performance

| Compound Label | Retention Time | Height      | Height % | Area       | Area % | Area Sum % | Symmetry | Width |
|----------------|----------------|-------------|----------|------------|--------|------------|----------|-------|
| Cpd 1          | 7.226          | 2348175.66  | 4.22     | 3163584.33 | 2.56   | 0.86       | 0.97     | 0.056 |
| Cpd 2          | 7.796          | 22207661.81 | 39.9     | 30039689   | 24.28  | 8.14       | 2        | 0.077 |
| Cpd 3          | 7.943          | 2984674.55  | 5.36     | 3106579.6  | 2.51   | 0.84       | 1.02     | 0.038 |
| Cpd 4          | 8.056          | 7152704.48  | 12.85    | 8799543.59 | 7.11   | 2.39       | 0.74     | 0.06  |
| Cpd 5          | 8.363          | 55665247.99 | 100      | 103595467  | 83.74  | 28.08      | 1.55     | 0.14  |
| Cpd 6          | 9.72           | 18511776.5  | 33.26    | 23585606.5 | 19.06  | 6.39       | 0.59     | 0.045 |
| Cpd 7          | 9.756          | 54187175.13 | 97.34    | 123712853  | 100    | 33.54      | 1.84     | 0.119 |
| Cpd 8          | 9.893          | 3856618.81  | 6.93     | 7770700.83 | 6.28   | 2.11       | 1.64     | 0.071 |
| Cpd 9          | 9.967          | 7926398.33  | 14.24    | 11915874.3 | 9.63   | 3.23       | 1.14     | 0.055 |
| Cpd 10         | 11.762         | 6539455.82  | 11.75    | 14086817.6 | 11.39  | 3.82       | 1.15     | 0.143 |
| Cpd 11         | 13.841         | 6059374.53  | 10.89    | 17972176.3 | 14.53  | 4.87       | 0.97     | 0.178 |
| Cpd 12         | 17.027         | 1087134.47  | 1.95     | 2481565.16 | 2.01   | 0.67       | 0.91     | 0.082 |
| Cpd 13         | 17.931         | 1300137.77  | 2.34     | 3087314.99 | 2.5    | 0.84       | 0.66     | 0.101 |
| Cpd 14         | 20.698         | 5330163.72  | 9.58     | 15564378.8 | 12.58  | 4.22       | 1.65     | 0.129 |

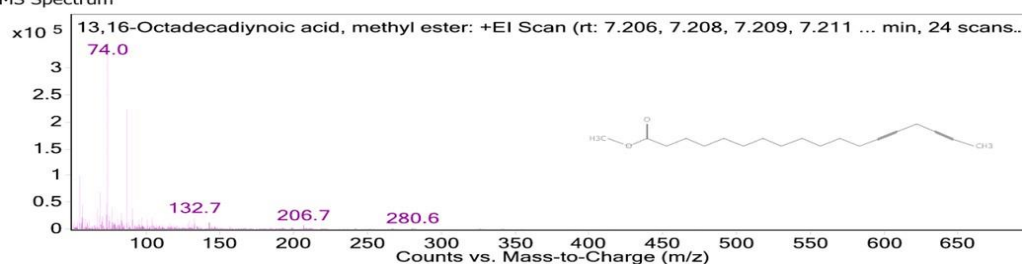
**Table 2.** List of identified Compound names, molecular formula, and retention time identified in GC-MS analysis.

| Compound Label | Retention Time | Compound Name                            | Molecular Formula                              |
|----------------|----------------|--|--|
| Cpd 1          | 7.226          | 13,16-Octadecadienoic acid, methyl ester | C <sub>19</sub> H <sub>30</sub> O <sub>2</sub> |
| Cpd 2          | 7.796          | 3,7,11,15-Tetramethyl-2 hexadecen-1-ol   | C <sub>20</sub> H <sub>40</sub> O              |
| Cpd 3          | 7.943          | 17-Octadecynoic acid                     | C <sub>18</sub> H <sub>32</sub> O <sub>2</sub> |
| Cpd 4          | 8.056          | 3,7,11,15-Tetramethyl-2 hexadecen-1-ol   | C <sub>20</sub> H <sub>40</sub> O              |
| Cpd 5          | 8.363          | Methyl 8-methyl-nonanoate                | C <sub>11</sub> H <sub>22</sub> O <sub>2</sub> |
| Cpd 6          | 9.72           | Methyl 11,12 octadecadienoate            | C <sub>19</sub> H <sub>34</sub> O <sub>2</sub> |
| Cpd 7          | 9.756          | 9-Octadecenoic acid (Z)-, methyl ester   | C <sub>19</sub> H <sub>36</sub> O <sub>2</sub> |
| Cpd 8          | 9.893          | Phytol                                   | C <sub>20</sub> H <sub>40</sub> O              |

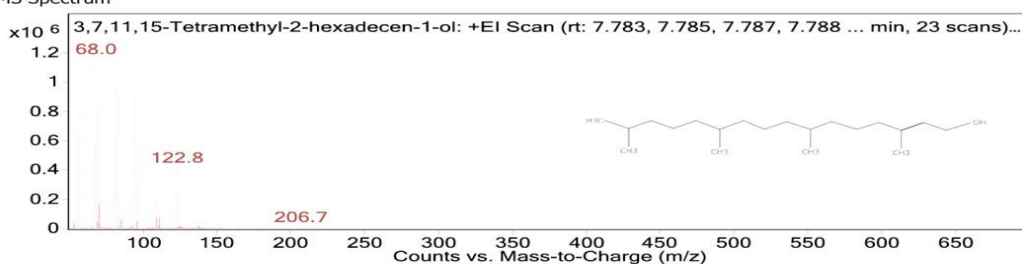


|        |        |   |  |
|--------|--------|---|--|
| Cpd 9  | 9.967  | Methyl 8-methyl-nonanoate   | C <sub>11</sub> H <sub>22</sub> O <sub>2</sub>                 |
| Cpd 10 | 11.762 | Dodecanedioic acid  | C <sub>12</sub> H <sub>22</sub> O <sub>4</sub>                 |
| Cpd 11 | 13.841 | 9,12-Octadecadienoyl chloride, (Z,Z)  | C <sub>18</sub> H <sub>31</sub> ClO                            |
| Cpd 12 | 17.027 | 9,12,15-Octadecatrienoic acid, 2-[(trimethylsilyl)oxy]-1-[[[(trimethylsilyl)oxy]methyl]ethyl ester, (Z,Z,Z) | C <sub>27</sub> H <sub>52</sub> O <sub>4</sub> Si <sub>2</sub> |
| Cpd 13 | 17.931 | [1-(3,3-Dimethyloxiran-2-ylmethyl)-3,7-dimethylocta-2,6-dienyl]trimethylsilane                              | C <sub>18</sub> H <sub>34</sub> OSi                            |
| Cpd 14 | 20.698 | 9,12-Octadecadiynoic acid, trimethylsilyl ester   | C <sub>21</sub> H <sub>36</sub> O <sub>2</sub> Si              |

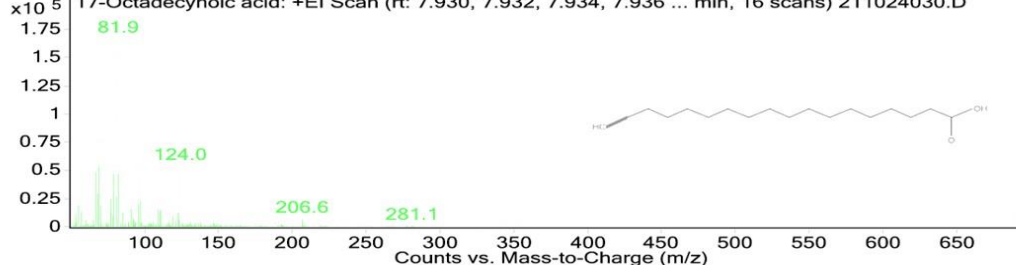
MS Spectrum



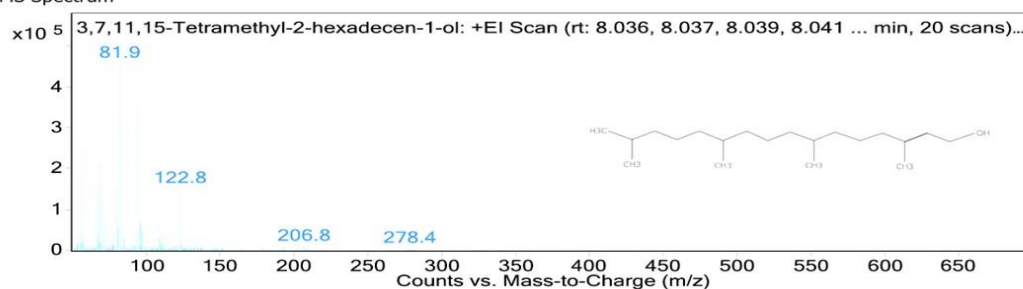
MS Spectrum



MS Spectrum

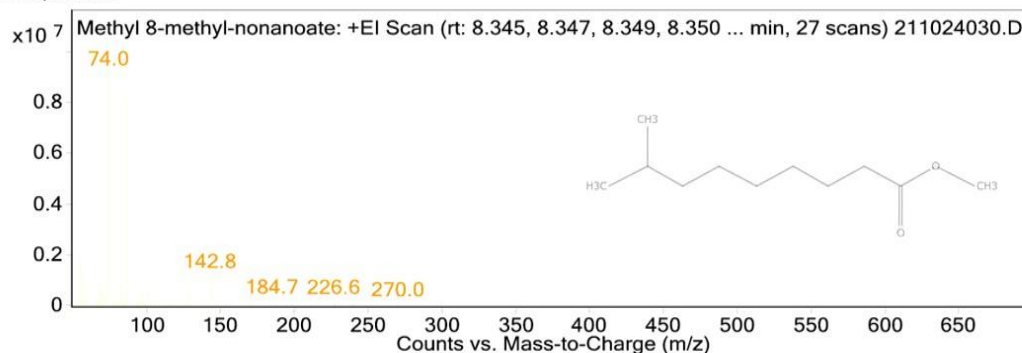


MS Spectrum

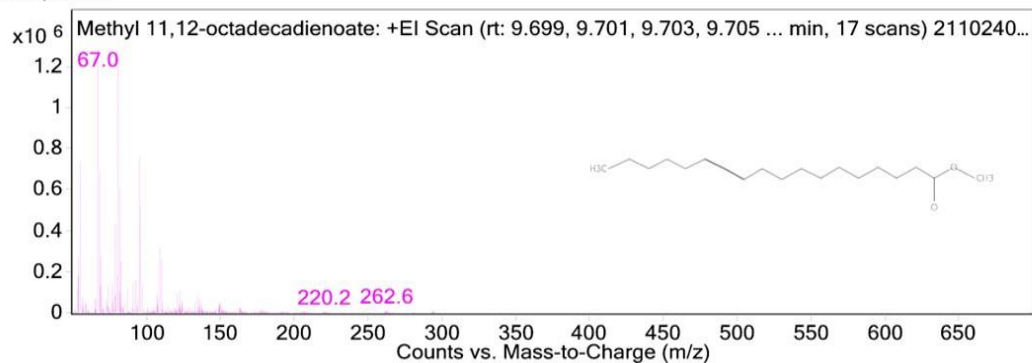




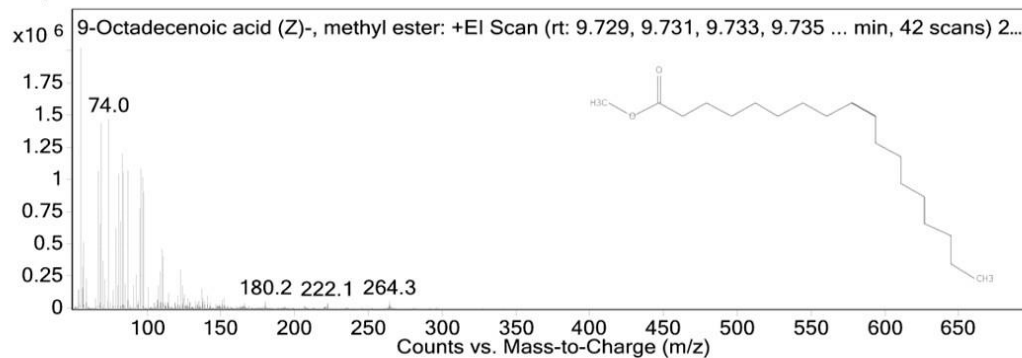
MS Spectrum



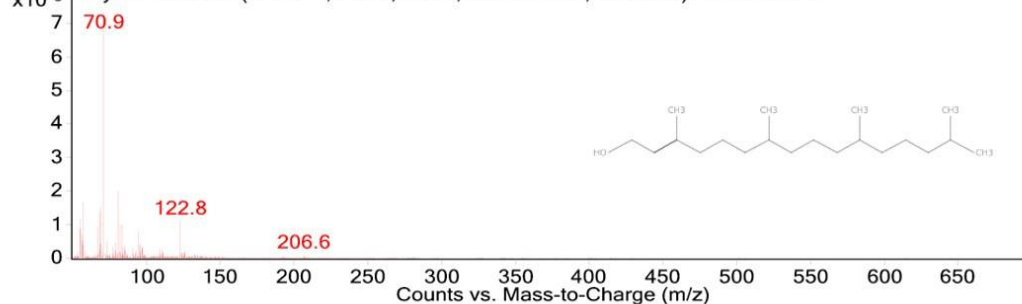
MS Spectrum



MS Spectrum

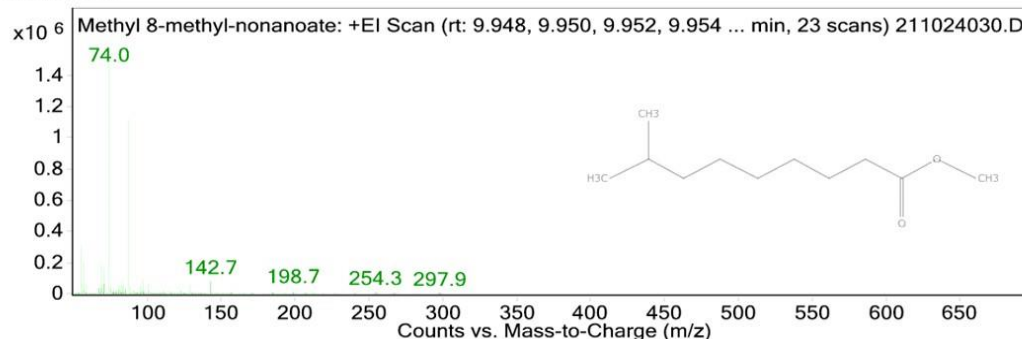


Phytol: +EI Scan (rt: 9.871, 9.873, 9.874, 9.876 ... min, 32 scans) 211024030.D

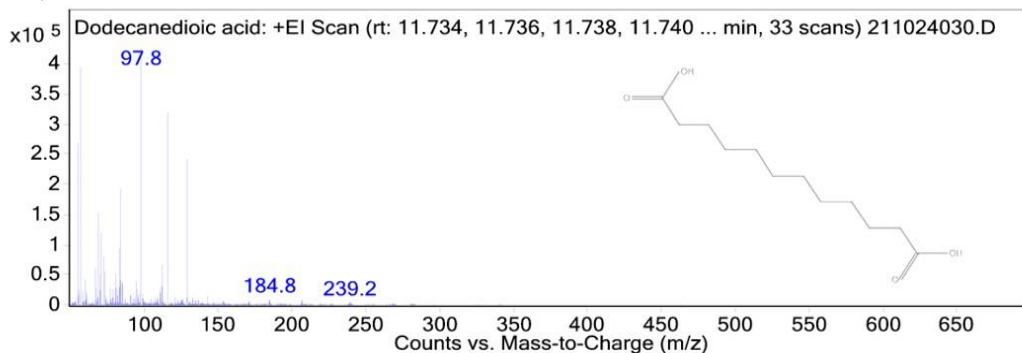




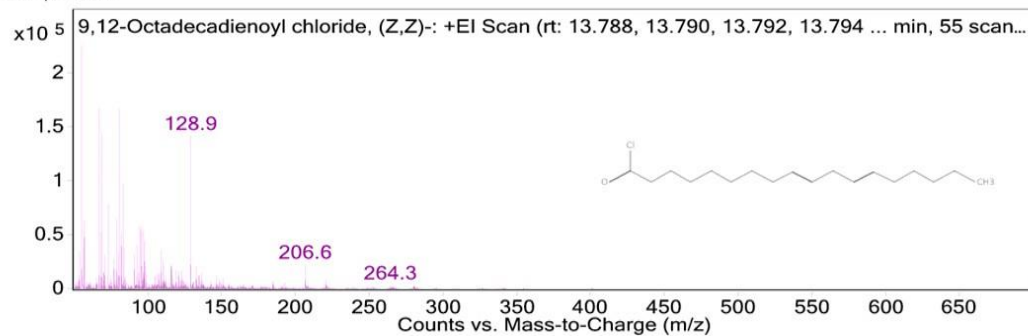
MS Spectrum



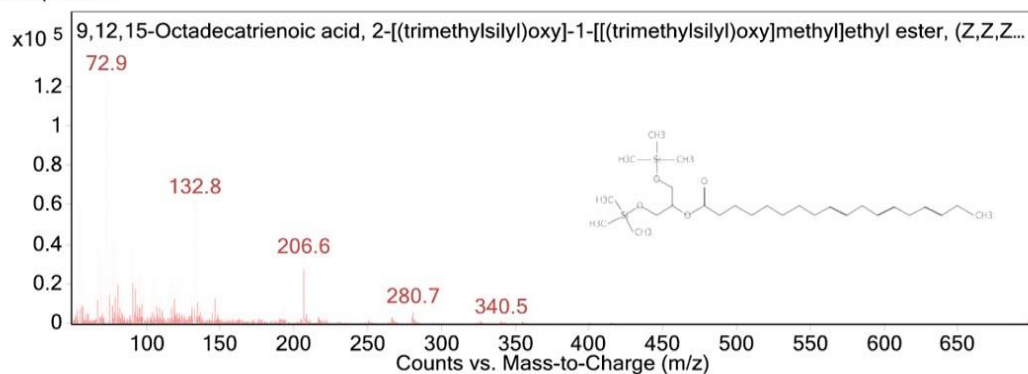
MS Spectrum

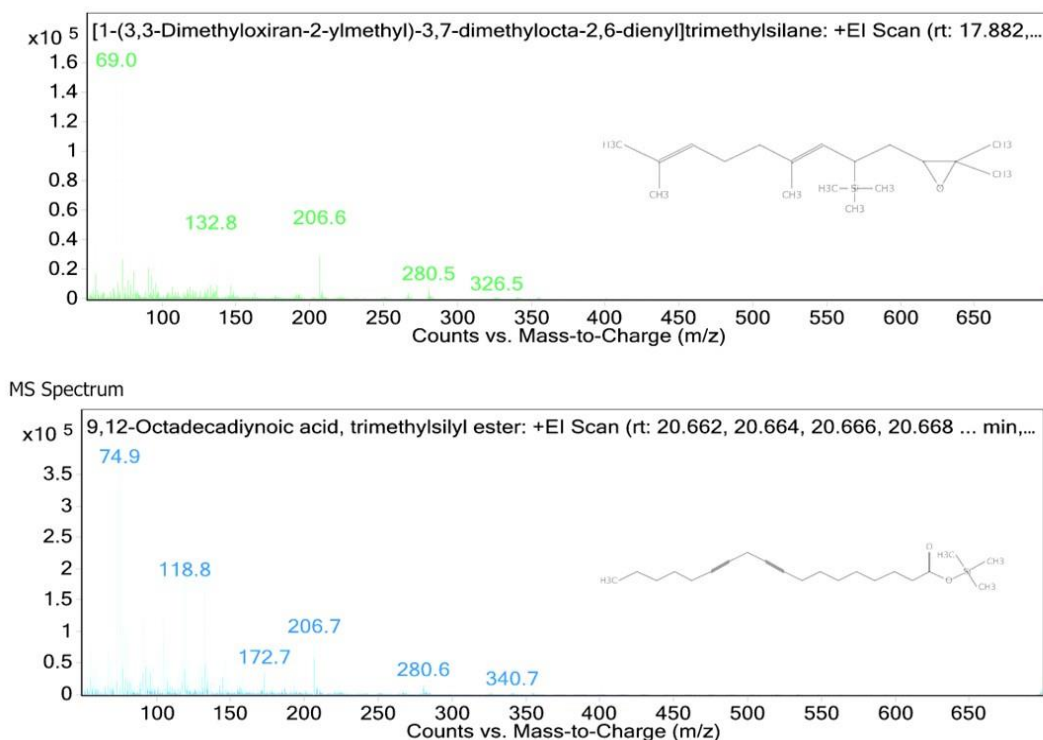


MS Spectrum



MS Spectrum





**Figure 2:** Mass spectra of fourteen compounds identified in the methanolic extract of *A. paniculata*.”

#### 4. Conclusion

The present study successfully employed GC-MS profiling to uncover the phytochemical landscape of the methanolic extract of *A. paniculata*. Among these fourteen identified constituents, one of them previously unreported bioactive compound was detected, alongside thirteen known bioactive compounds with various pharmacological potentials. The discovery of the novel compound adds a new dimension to the therapeutic value of *A. paniculata*, highlighting its promise as a reservoir of unexplored medicinal agents. These findings not only contribute to the existing phytochemical database of the plant but also provide a foundation for future pharmacological investigations and potential drug development based on its bioactive constituents.

#### References

1. Cragg, G. M., & Newman, D. J. (2005). Plants as a source of anti-cancer agents. *Journal of ethnopharmacology*, 100(1-2), 72-79.
2. Das, P., Tiwari, P., & Ramesh, D. B. (2019). Research in Pharmaceutical Sciences in India: A Scientometric assessment of research output during 1998–2017. *Research Journal of Pharmacy and Technology*, 12(4), 1551-1558.
3. Dwivedi, M. K., Sonter, S., Mishra, S., Singh, P., & Singh, P. K. (2021). Secondary metabolite profiling and characterization of diterpenes and flavones from the methanolic extract of *Andrographis paniculata* using HPLC-LC-MS/MS. *Future Journal of Pharmaceutical Sciences*, 7, 1-28.
4. El-Saadony, M. T., Saad, A. M., Mohammed, D. D. M., Korma, S. A., Alshahrani, M. Y., Ezzat Ahmed, A., ... & Ibrahim, (2025) S. A. Medicinal plants: Bioactive compounds, biological activities, combating multidrug-resistant microorganisms, and human health benefits-A comprehensive review. *Frontiers in Immunology*, 16, 1491777.
5. Fardiyah, Q., Ersam, T., Slamet, A., & Kurniawan, F. (2020). New potential and characterization of *Andrographis paniculata* L. Ness plant extracts as photoprotective agent. *Arabian Journal of Chemistry*, 13(12), 8888-8897.
6. Favour, O. E., Angela, O. C., Ezinne, G. T., Juliet, A. C., Simeon, B., Godson, C. C., ... & Nnaemeka, O. H. (2025). Current Insights on the Effects of



- Medicinal Plants in the Management of Obesity and Infectious Diseases: An Update from 2020. *Aspects of Molecular Medicine*, 100075.
7. Hayat, J., Akodad, M., Moumen, A., Baghour, M., Skalli, A., Ezrari, S., & Belmalha, S. (2020). Phytochemical screening, polyphenols, flavonoids and tannin content, antioxidant activities and FTIR characterization of *Marrubium vulgare* L. from 2 different localities of Northeast of Morocco. *Heliyon*, 6(11).
  8. Jayakumar, T., Hsieh, C. Y., Lee, J. J., & Sheu, J. R. (2013). Experimental and clinical pharmacology of *Andrographis paniculata* and its major bioactive phytoconstituent andrographolide. *Evidence-Based Complementary and Alternative Medicine*, 2013(1), 846740.
  9. Karimi, E., & Jaafar, H. Z. (2011). HPLC and GC-MS determination of bioactive compounds in microwave obtained extracts of three varieties of *Labisia pumila* Benth. *Molecules*, 16(8), 6791-6805.
  10. Mayavanshi, A. V., & Gajjar, S. S. (2008). Floating drug delivery systems to increase gastric retention of drugs: A Review. *Research Journal of Pharmacy and Technology*, 1(4), 345-348.
  11. Mehta, S., Sharma, A. K., & Singh, R. K. (2021). Pharmacological activities and molecular mechanisms of pure and crude extract of *andrographis paniculata*: An update. *Phytomedicine Plus*, 1(4), 100085.
  12. Mu, J., Brozinick, J. T., Valladares, O., Bucan, M., & Birnbaum, M. J. (2001). A role for AMP-activated protein kinase in contraction-and hypoxia-regulated glucose transport in skeletal muscle. *Molecular cell*, 7(5), 1085-1094.
  13. Schlenk, H., Gellerman, J. L., Tillotson, J. A., & Mangold, H. K. (1957). Paper chromatography of lipides. *Journal of the American Oil Chemists' Society*, 34(8), 377-386.
  14. Singla, S., Sharma, S., Basu, S., Shetti, N. P., & Aminabhavi, T. M. (2021). Photocatalytic water splitting hydrogen production via environmental benign carbon-based nanomaterials. *International Journal of Hydrogen Energy*, 46(68), 33696-33717.
  15. Zagoskina, N. V., Zubova, M. Y., Nechaeva, T. L., Kazantseva, V. V., Goncharuk, E. A., Katanskaya, V. M., ... & Aksenova, M. A. (2023). Polyphenols in plants: Structure, biosynthesis, abiotic stress regulation, and practical applications. *International Journal of Molecular Sciences*, 24(18), 13874.
  16. Mariani, M. L., & Penissi, A. B. (2018). Development and Validation of a Novel HPLC Method for the Analysis and Quantification of Dehydroleucodine in Plant Extracts. *Current Chromatography*, 5(2), 104-111.