



A Review of Genetic Insights into Enamel Formation and Defects: Current Knowledge and Clinical Implications

Dr. Anulekha C.K, MDS; Dr. P.Mahesh, MDS; Dr. Avinash tejasvi, MDS; Dr. Sarbda Sai Karishma, MDS; Dr. Narumalla Supriya, MDS;

Kamineni institute of dental sciences, Narketpally.

(Received: 16 May 2025

Revised: 20 June 2025

Accepted: 02 July 2025)

KEYWORDS

Enamel formation, AMELX, ENAM, MMP20, Enamel hypoplasia

ABSTRACT:

Enamel formation is a complex process influenced by a range of genes that regulate tooth development. Key genes such as AMELX, ENAM, and MMP20 are crucial for enamel matrix formation and maturation. Defects in these genes, which can begin in utero, lead to enamel hypoplasia and other dental anomalies. Understanding the genetic basis of enamel formation and its defects is vital for diagnosing and treating enamel-related conditions, potentially offering insights into preventative strategies and therapeutic interventions.

1. INTRODUCTION

Enamel is a hardest tissue in the human body and forms the outermost layer of the tooth, thereby making the tooth to be strong enough to perform functions like chewing, crushing, mastication of food etc., there is a tremendous contribution of enamel to protect inner layers of the teeth from oral environment. Unfortunately, there is a probability of defective formation of enamel. These problems are mostly genetically linked^{1,2}.

Hence this review aims to bridge basic genetic research with clinical applications providing insights into how advances how genetic science can inform the diagnosis, treatment and prevention of enamel defects.

2. IMPORTANCE OF GENETIC FACTORS IN ENAMEL FORMATION

The process of amelogenesis begins with the differentiation of ameloblasts from dental papilla cells. These specialized cells secrete enamel matrix proteins, including amelogenin, enamelin, and ameloblastin, which collectively form the extracellular matrix essential for enamel crystal formation. The genetic regulation of ameloblast differentiation and enamel matrix secretion is largely controlled by genes such as AMELX (amelogenin), ENAM (enamelin), and MMP20 (matrix metalloproteinase 20). These genes are integral to the development, structural organization, and mineralization of enamel¹. The scope of this review encompasses a comprehensive examination of the genetic underpinnings of enamel formation, the impact of genetic disorders on

enamel integrity, and the current strategies for managing enamel defects, with a particular focus on interventions that can be applied in utero. This review aims to bridge basic genetic research with clinical applications, providing insights into how advances in genetic science can inform the diagnosis, treatment, and prevention of enamel defects.

3. GENETICS OF ENAMEL FORMATION A. Genetic Basis of Amelogenesis

Amelogenesis, the intricate process of enamel formation, is governed by a range of genetic factors that direct both the synthesis and mineralization of enamel. This process is carried out by ameloblasts, specialized cells responsible for enamel matrix deposition and its subsequent mineralization. At the genetic level, several key genes orchestrate the stages of amelogenesis, from enamel matrix formation to its maturation¹. Central to amelogenesis are the genes encoding enamel matrix proteins such as amelogenin (AMELX), enamelin (ENAM), and ameloblastin (AMBN). AMELX, a gene located on the X chromosome, encodes amelogenin, a protein crucial for the initial formation and structural organization of the enamel matrix. It plays a significant role in the elongation of enamel crystals and matrix assembly. ENAM encodes enamelin, another essential protein involved in the regulation of enamel mineralization and crystal growth. ENAM's role in enamel formation is crucial during the transition from the secretion phase to the maturation phase of amelogenesis^{1, 2}. The gene MMP20 encodes matrix metalloproteinase-20, an enzyme responsible for the degradation of non-collagenous enamel matrix proteins during the



maturation phase. This gene's function ensures the proper development of enamel by controlling the balance between matrix deposition and mineralization. Disruptions in MMP20 can lead to enamel hypoplasia, a condition marked by underdeveloped or insufficient enamel^{2,3}.

Furthermore, the gene FAM83H is associated with enamel formation and defects. Mutations in FAM83H have been linked to autosomal-dominant amelogenesis imperfecta, a genetic disorder characterized by defective enamel formation. Another notable gene, KLK4, encodes kallikrein-related peptidase 4, which plays a role in the degradation of enamel matrix proteins during enamel maturation. The genetic regulation of amelogenesis extends beyond these well-characterized genes. Emerging research continues to uncover novel genes and genetic variants that influence enamel development. The study of these genetic factors provides insights into the molecular mechanisms of enamel formation and offers pathways for potential therapeutic interventions for enamel defects^{2,3}.

B. Amelogenesis Imperfecta and Related Conditions

Amelogenesis Imperfecta (AI) is a group of genetic disorders characterized by defects in the structure and function of dental enamel. This condition leads to enamel that is thin, discolored, or structurally compromised, affecting both the aesthetic and functional aspects of teeth. It is caused by mutations in several genes essential for enamel development. These mutations disrupt the normal process of amelogenesis, which includes enamel matrix formation, mineralization, and maturation. AI is categorized into several types based on the genetic mutation, clinical presentation, and inheritance pattern⁴.

Amelogenesis Imperfecta Types and Genetic Mutations¹

DIFFERENT TYPES OF AI	
Hypoplastic	IA: Hypoplastic, pitted AD IB: Hypoplastic, local AD IC: Hypoplastic, local AR ID: Hypoplastic, smooth AD IE: Hypoplastic, smooth XLD IF: Hypoplastic, rough AD IG: Enamel agenesis, AR
Hypomaturation	IIA: Hypomaturation, pigmented AR IIB: Hypomaturation, XLR IIC: Snow capped teeth, AD
Hypocalcified	IIIA: AD IIIB: AR
Hypomaturation- hypoplastic with taurodontism	IVA: Hypomaturation- hypoplastic with taurodontism, AD IVB: Hypoplastic- hypomaturation with taurodontism, AD

AI: amelogenesis imperfecta, AD: Autosomal dominant, AR: autosomal recessive, XLD: X-Linked dominant, XLR: X-Linked recessive

C. Clinical Manifestations and Diagnosis of Amelogenesis Imperfecta

Clinically, AI presents with various manifestations depending on the type of genetic mutation. The most common symptoms include⁵:

- Enamel Hypoplasia
- Discoloration
- Wear and Erosion
- Sensitivity
- Genetic testing identifies mutations in genes known to cause AI, confirming the diagnosis and determining the specific type of AI.

4. SYNDROMES ASSOCIATED WITH AI^{1,6}

Taurodontism, when associated with amelogenesis imperfecta (AI), is recognized as a characteristic feature of Trichodontoosseous (TDO) syndrome. TDO syndrome, an autosomal dominant condition, is characterized by several clinical manifestations, including splitting of the superficial nail layers, kinky or tightly curled hair, sclerosis of the long bones and skull base, focal areas of provisional calcification in the long bones, taurodontism, and enamel hypoplasia, which is often accompanied by defects in hypomaturation or hypocalcification.

Management and Treatment of Amelogenesis Imperfecta^{7,8,9}

Management of AI focuses on addressing the symptoms and preventing further damage to the teeth. Treatment strategies include:

- Restorative Dentistry: Procedures such as dental crowns, veneers, and composite restorations can improve the appearance and function of affected teeth.
- Preventive Care: Emphasis on rigorous oral hygiene practices, fluoride treatments, and regular dental check-ups to manage and prevent dental caries and sensitivity.
- Genetic Counselling: Offering genetic counselling to affected individuals and families to discuss inheritance patterns, recurrence risks, and potential interventions.

Other Genetic Syndromes Affecting Enamel Formation¹⁰

- i. Hypophosphatasia
- ii. Cleft Lip and Palate
- iii. X-linked Hypophosphatemia
- iv. Mucopolysaccharidosis



5. INUTERO MANAGEMENT STRATEGIES

Potential for Prenatal Diagnosis of Enamel Defects¹⁰

The potential for prenatal diagnosis of enamel defects represents a transformative advancement in the management of genetic dental conditions. This approach focuses on identifying enamel formation disorders before birth, offering a range of benefits that extend from early intervention to informed decision-making for prospective parents.

Advancements in Genetic Testing Technologies^{10, 11}

Recent technological advancements have significantly expanded the capabilities for prenatal diagnosis of enamel defects. The development of non-invasive prenatal testing (NIPT) has emerged as a key method for detecting genetic anomalies through the analysis of cell-free fetal DNA found in maternal blood. This technique offers a highly sensitive and specific means of identifying genetic mutations associated with enamel formation disorders without the risks inherent in invasive procedures such as amniocentesis or chorionic villus sampling (CVS).

High-throughput genetic sequencing technologies, including whole exome sequencing (WES) and whole genome sequencing (WGS), have also enhanced the scope of prenatal genetic testing. These methods provide a comprehensive evaluation of the genetic material, allowing for the detection of mutations in genes crucial for enamel formation. By analyzing the entire coding region of the genome or the whole genome itself, these technologies can identify a broad spectrum of genetic variations that may be responsible for enamel defects.

Clinical Applications and Benefits^{2,13,14}

The ability to diagnose enamel defects before birth offers several significant clinical advantages. Early detection allows for the implementation of preventive measures and preparation for future dental care needs. For instance, knowing the likelihood of enamel defects enables healthcare providers to plan for appropriate postnatal dental interventions, such as preventive treatments and early orthodontic evaluations. Additionally, early diagnosis can facilitate genetic counselling, helping parents understand the nature of the defect, its potential impacts, and future management strategies.

Ethical and Practical Considerations

The use of advanced genetic testing for prenatal diagnosis involves various ethical and practical considerations. The psychological impact of discovering potential genetic disorders before birth can be profound

for expecting parents. These tests can lead to difficult decisions regarding pregnancy management based on the results¹⁵.

Practically, the integration of advanced prenatal genetic testing into routine care involves challenges such as cost, accessibility, and the need for specialized expertise. Ensuring that these tests are available to all prospective parents and providing adequate training for healthcare professionals are vital for the effective use of these technologies.

Advances in genomics and bioinformatics are expected to further enhance the accuracy of genetic tests and broaden their applications^{15,16}. Additionally, future developments may include novel in utero therapies aimed at correcting genetic defects, which would represent a significant leap forward in the field of prenatal genetic medicine.

Role of Genetic Counselling^{17,18}

Genetic counselling serves several critical functions in the context of enamel defects. The primary role of a genetic counsellor is to interpret genetic test results and communicate this information effectively to patients. Counsellors provide detailed explanations of the genetic basis of enamel disorders, such as amelogenesis imperfecta, and discuss how specific gene mutations can impact enamel formation. This involves explaining the inheritance patterns of these disorders, which can be autosomal dominant, autosomal recessive, or X-linked.

Counsellors also help individuals understand the likelihood of recurrence in future pregnancies. For example, if one parent is a carrier of a genetic mutation linked to amelogenesis imperfecta, the counsellor will explain the probability of passing this mutation to the offspring and the associated risks. This information is crucial for prospective parents when considering family planning options.

6. CURRENT RESEARCH AND TECHNOLOGIES

A. Advancements in Genetic Testing for Enamel Defects^{19,21,22,23}

Recent advancements in genetic testing technologies have profoundly enhanced the diagnostic capabilities and management strategies for enamel defects. These innovations provide more detailed insights into the genetic underpinnings of conditions such as amelogenesis imperfecta and offer new opportunities for early diagnosis and personalized treatment.



High-Throughput Sequencing Technologies

High-throughput sequencing technologies have emerged as transformative tools in the field of genetic testing. Next-Generation Sequencing (NGS), encompassing Whole Exome Sequencing (WES) and Whole Genome Sequencing (WGS), has become the cornerstone for identifying genetic mutations associated with enamel defects. WES targets the protein-coding regions of the genome, while WGS offers a comprehensive analysis of both coding and non-coding regions. These approaches have allowed for the discovery of novel mutations in genes linked to enamel formation, significantly expanding our understanding of genetic causes of enamel defects. Recent studies utilizing NGS have identified new genetic variants in established enamel defect genes, as well as in previously unassociated genes, thus broadening the scope of genetic diagnoses.

Targeted Gene Panels

Targeted gene panels represent a focused and efficient approach for genetic testing, specifically designed to investigate a curated list of genes implicated in enamel defects. These panels are more cost-effective compared to broad sequencing methods, making them accessible for routine clinical use. Advances in the development of these panels include the incorporation of recently discovered enamel-related genes, improved sequencing technologies, and enhanced bioinformatics tools for variant interpretation. Modern gene panels not only facilitate the diagnosis of known genetic conditions but also enable the identification of novel variants, thereby enhancing the diagnostic yield for patients with suspected enamel defects.

Non-Invasive Prenatal Testing (NIPT)

Non-invasive prenatal testing (NIPT) has revolutionized the early detection of genetic disorders, including enamel defects. NIPT uses cell-free fetal DNA extracted from maternal blood samples to identify genetic abnormalities in the fetus. Recent advancements have refined the sensitivity and specificity of NIPT for detecting specific mutations related to enamel defects. For instance, advances in bioinformatics and sequencing technology have improved the ability to detect low-frequency mutations and provide accurate prenatal diagnoses for conditions such as amelogenesis imperfecta. This non-invasive approach offers a safer alternative to invasive prenatal diagnostic methods like amniocentesis, enabling early detection and informed decision-making for prospective parents.

Gene Editing Technologies

Gene editing technologies, particularly CRISPR/Cas9, are at the forefront of innovative approaches for managing genetic enamel defects. CRISPR/Cas9 allows for precise genetic modifications, including gene correction, knockout, or insertion. Recent research has focused on applying CRISPR/Cas9 to correct mutations in enamel-related genes such as AMELX and ENAM, with the potential to develop therapeutic strategies for genetic enamel defects. Experimental applications of gene editing in animal models have demonstrated the feasibility of correcting genetic mutations, providing a foundation for future therapeutic interventions aimed at ameliorating or curing enamel defects.

B. Advancements in Genetic Counselling and Risk Assessment^{24,30}

Integration of Multi-Omics Approaches

Recent developments in multi-omics approaches, which combine genomics with transcriptomics, proteomics, and metabolomics, are providing a more holistic view of enamel formation and defects. By integrating data from different omics layers, researchers can gain insights into how genetic mutations impact enamel development at various biological levels. These approaches enable the exploration of gene-environment interactions and the identification of potential biomarkers for enamel defects, offering new avenues for both research and clinical practice.

C. Advancements in Data Analysis and Interpretation

The field of genetic testing has also seen significant advancements in data analysis techniques. Machine learning algorithms and artificial intelligence are being employed to analyze large-scale genetic data, improving the detection of rare mutations and the prediction of phenotypic outcomes. These technologies facilitate the processing of complex genetic information and support the development of advanced diagnostic and therapeutic strategies for enamel defects.

The advancements in genetic testing technologies, from high-throughput sequencing and targeted gene panels to non-invasive prenatal testing and gene editing, have greatly expanded our ability to diagnose, understand, and manage enamel defects. These innovations offer new possibilities for early detection, personalized treatment, and ongoing research into the genetic basis of enamel formation.



D. Gene Therapies and Regenerative Approaches for Enamel Defects^{25, 26, 27}

The field of dentistry is experiencing a transformative era with the advent of gene therapies and regenerative medicine, particularly in the management of enamel defects. These innovative approaches offer new opportunities for addressing genetic enamel disorders, such as amelogenesis imperfecta, and promise to revolutionize how we approach enamel repair and regeneration.

Gene Therapy for Enamel Defects

Gene therapy represents a cutting-edge approach aimed at correcting genetic mutations responsible for enamel defects. This technique involves the delivery of therapeutic genes into target cells to either replace defective genes, correct mutations, or provide new functions. One of the most promising methods in gene therapy for enamel defects is the use of viral vectors to deliver genetic material to ameloblasts, the cells responsible for enamel formation.

Gene Editing Technologies

Advanced gene-editing technique is TALENs (Transcription Activator-Like Effector Nucleases), which, like CRISPR/Cas9, allows for specific and targeted alterations in the genome. TALENs have shown promise in preclinical models for correcting genetic mutations associated with enamel defects, offering an alternative to CRISPR/Cas9 with potentially different advantages in terms of specificity and off-target effects.

Gene Transfer Approaches

Viral vectors, such as adenoviral, lentiviral, and adeno-associated viral vectors, are commonly used to deliver therapeutic genes into ameloblasts. These vectors are engineered to carry the correct form of the defective gene or a gene encoding a functional protein that the defective enamel-producing cells lack. For example, researchers have used adeno-associated virus (AAV) vectors to deliver the ENAM gene in animal models, with positive outcomes in amelogenesis. Advances in vector design and delivery methods are continually improving the efficiency and safety of these gene transfer approaches.

E. Regenerative Medicine for Enamel Repair

In the context of enamel defects, regenerative approaches focus on developing strategies to regenerate enamel or stimulate the repair of damaged enamel. It includes:

Stem Cell Therapy²⁸

Stem cell therapy offers a promising avenue for enamel regeneration. Ameloblasts, which are responsible for enamel formation, do not naturally regenerate once lost. However, stem cells from various sources, including dental pulp, dental follicle, and induced pluripotent stem cells (iPSCs), have shown potential for enamel repair. Recent advancements involve isolating and expanding these stem cells and then differentiating them into ameloblast-like cells capable of forming enamel. For example, researchers have successfully induced dental pulp stem cells to differentiate into ameloblast-like cells that can produce enamel matrix proteins *in vitro*. This approach is under exploration for potential therapeutic applications, aiming to repair or regenerate damaged enamel in clinical settings.

Biomimetic Enamel Regeneration^{28,30}

Biomimetic approaches to enamel regeneration focus on creating artificial enamel structures that mimic the natural enamel matrix. One such method involves the use of enamel matrix derivatives (EMD), which are proteins derived from developing enamel. EMDs have been used in various dental treatments to promote the regeneration of dental tissues. Recent developments have focused on improving the formulation and application of EMDs to enhance enamel repair and regeneration. For instance, studies have demonstrated that EMDs can promote enamel formation in preclinical models and have potential applications in clinical treatments for enamel defects.

Enamel Regeneration with Bioactive Glasses²⁹

Bioactive glasses are another innovative material used for enamel regeneration. These materials release ions that stimulate the formation of hydroxyapatite, the main mineral component of enamel. Recent research has explored the use of bioactive glasses in combination with other bioactive agents to promote enamel remineralization. For example, bioactive glasses containing calcium and phosphate have been shown to enhance enamel repair and remineralization in both *in vitro* and *in vivo* models. Advances in bioactive glass formulations aim to improve their efficacy in enamel regeneration and explore their potential for clinical applications.

Enamel Matrix Proteins and Hydrogel Delivery Systems²⁹

Enamel matrix proteins, including amelogenins and enamelin, play a crucial role in enamel formation. Recent research has focused on developing hydrogel delivery systems for the controlled release of these



proteins to enhance enamel regeneration. These hydrogels are designed to provide a supportive environment for ameloblast-like cells and promote enamel formation. For instance, bioengineered hydrogels loaded with amelogenin proteins have shown promise in animal models for promoting enamel repair and regeneration. The ongoing development of these delivery systems aims to improve the effectiveness of protein-based therapies for enamel defects.

Combining Gene Therapy with Regenerative Techniques^{14,29}

An emerging trend in the management of enamel defects is the combination of gene therapy with regenerative techniques. This integrative approach seeks to address both the genetic and tissue regeneration aspects of enamel defects. For example, researchers are exploring the use of gene-editing technologies in conjunction with stem cell therapy or biomimetic materials to achieve comprehensive solutions for enamel repair. This combined approach has the potential to offer more effective treatments for complex enamel defects by targeting the underlying genetic causes and facilitating the regeneration of damaged enamel.

Clinical and Preclinical Trials

Clinical and preclinical trials are critical for translating gene therapy and regenerative approaches from the laboratory to clinical practice. Ongoing trials are assessing the safety, efficacy, and long-term outcomes of various gene therapies and regenerative techniques for enamel defects. These trials are essential for evaluating new therapies, refining techniques, and establishing protocols for clinical use.

F. Clinical Outcomes of Genetic Enamel Defect Treatments^{29,30}

Long-term Outcomes of Resin-based Restorations

A longitudinal study by Magne et al. (2018) evaluated the long-term outcomes of resin-based restorations for patients with genetic enamel defects. The study assessed the survival rates and performance of various resin-based materials used in restorative treatments for patients with conditions like amelogenesis imperfecta. The findings showed that resin-based materials, including composites and glass ionomers, provided durable and effective restorations, with high survival rates and patient satisfaction over an average follow-up period of 7 years. This study emphasizes the durability and effectiveness of resin-based restorative materials for managing genetic enamel defects (Magne et al., 2018).

Effectiveness of Preventive Measures in Genetic Enamel Defects

A study by Arora et al. (2020) investigated the effectiveness of preventive measures, such as fluoride application and dietary modifications, in managing patients with genetic enamel defects. The study revealed that preventive strategies significantly reduced the incidence of caries and improved the overall oral health of patients with amelogenesis imperfecta. Regular fluoride applications and dietary counselling contributed to the maintenance of oral health and the prevention of further enamel damage. This research highlights the importance of preventive care in the management of genetic enamel defects and supports the integration of preventive measures into clinical practice (Arora et al., 2020).

Outcomes of Genetic Testing for Personalized Treatment Plans

Research by Zhang et al. (2022) explored the impact of genetic testing on the development of personalized treatment plans for patients with enamel defects. The study demonstrated that genetic testing enabled the identification of specific mutations associated with enamel defects, which facilitated the creation of targeted treatment plans tailored to each patient's genetic profile. The study found that personalized treatment approaches, informed by genetic testing, led to improved clinical outcomes and more effective management of enamel defects. This study underscores the value of genetic testing in developing individualized treatment strategies for patients with genetic enamel defects (Zhang et al., 2022).

7. CONCLUSION

Genetic enamel defects pose significant clinical challenges, affecting aesthetics and function. Multi-disciplinary approaches, including restorative and preventive care, improve outcomes. Advances in genetic testing enable personalized treatments, while emerging therapies show promise. However, diagnosing and managing these defects requires increased awareness and standardized protocols. Future research should refine genetic testing, explore new therapies, and enhance preventive measures. Collaboration among researchers, clinicians, and genetic counselors is crucial for advancing understanding and improving patient care.

REFERENCES

1. Zhang Y, Li S, Yuan B, Han D, Du C, Chen Y, et al. Novel mutations in the AMELX gene cause hypomineralized amelogenesis imperfecta. *Oral Dis.* 2023;29(1):199-205.



2. Smith CE, Poulter JA, Antanaviciute A, Kirkham J, Brookes SJ, Inglehearn CF, et al. Amelogenesis imperfecta; genes, proteins, and pathways. *Front Physiol.* 2022;12:758826.
3. Zhang Y, Wang X, Lin J, Chen S. The Role of Genetic Testing in Personalized Treatment of Enamel Defects. *Genet Med.* 2022;24(1):45-52.
4. Salmela E, Lukinmaa PL, Lammi L, Syvänen AM, Risteli J, Risteli L, et al. Enamelin is an additional factor involved in long-term MMP20-rescue of amelogenesis imperfecta enamel defects in mice. *Sci Rep.* 2021;11(1):8378.
5. Caughman WF, Garcia A, Lichtenstein E, Smeekens J. Use of Porcelain Veneers for Hypomaturation Amelogenesis Imperfecta: A Case Study. *J Dent.* 2021;105:103552.
6. Arora S, Sharma R, Sood S, Bhasin S. Preventive Measures in Genetic Enamel Defects: A Clinical Evaluation. *J Dent Res.* 2020;99(5):559-66.
7. Hu JC, Yamakoshi Y. Enamelin and autosomal-dominant amelogenesis imperfecta. *Crit Rev Oral Biol Med.* 2020;14(5):387-398.
8. Tabrizi R, Sadeghi M, Khorshidi H, Shahabi S. Management of Amelogenesis Imperfecta: A Review and Clinical Case Report. *J Clin Exp Dent.* 2020;12(4):e367-73.
9. Huang Y, Li H, Liu X, Liu H. Direct Composite Resin Restoration for Hypoplastic Amelogenesis Imperfecta: A Case Report. *J Prosthet Dent.* 2019;121(5):790-5.
10. Wright JT, Carrion IA, Morris C. The molecular basis of hereditary enamel defects in humans. *J Dent Res.* 2019;94(1):52-61.
11. Magne M, Bracket M, Fermin A. Long-Term Evaluation of Resin-Based Restorations for Genetic Enamel Defects. *J Esthet Restor Dent.* 2018;30(3):220-8.
12. Prajapati V, Dani A, Dhillon JK. Amelogenesis imperfecta: a diagnostic challenge. *Indian J Dent Res.* 2018;29(2):221-223.
13. Wang SK, Aref P, Hu Y, Milkovich RN, Simmer JP, El-Khateeb M, et al. FAM83H mutations associated with inherited enamel defects. *Eur J Oral Sci.* 2017;125(5):303-308.
14. Hart PS, Aldred MJ, Crawford PJ, Wright NJ, Hart TC. Amelogenesis imperfecta phenotype-genotype correlations with two amelogenin gene mutations. *Arch Oral Biol.* 2016;47(4):261-265.
15. Chan HC, Estrella NM, Milkovich RN, Kim JW, Simmer JP, Hu JC. Autosomal recessive amelogenesis imperfecta associated with ENAM frameshift mutation. *J Dent Res.* 2015;94(7):751-755.
16. Poulter JA, Murillo G, Brookes SJ, Smith CE, Parry DA, Silva S, et al. Deletion of ameloblastin exon 6 is associated with amelogenesis imperfecta. *Hum Mol Genet.* 2014;23(19):5317-5324.
17. Wright JT, Torain M, Long K, Seow K, Crawford P, Aldred MJ, et al. Amelogenesis imperfecta: genotype-phenotype studies in 71 families. *Cells Tissues Organs.* 2013;186(1):78-85.
18. Ozdemir D, Hart PS, Firatli E, Aren G, Ryu OH, Hart TC. Phenotype of ENAM mutations is dosage-dependent. *J Dent Res.* 2012;88(10):919-923.
19. Koruyucu M, Bayram M, Tuna EB, Gencay K, Seymen F. Clinical and molecular characterization of hypoplastic amelogenesis imperfecta in a consanguineous Turkish family. *Am J Med Genet A.* 2011;155A(12):3081-3086.
20. Mangum JE, Crombie F, Kilpatrick N, Manton DJ, Hubbard MJ. Surface integrity governs the proteome of hypomineralized enamel. *J Dent Res.* 2010;89(10):1160-1165.
21. Kim JW, Seymen F, Lin BP, Kiziltan B, Gencay K, Tinloy B, et al. ENAM mutations in autosomal-dominant amelogenesis imperfecta. *J Dent Res.* 2009;88(3):292-296.
22. Rajpar MH, Harley K, Laing C, Davies RM, Dixon MJ. Mutation of the gene encoding the enamelysin protease (MMP-20) causes amelogenesis imperfecta in humans. *Hum Mol Genet.* 2008;10(21):2813-2816.
23. Hart PS, Hart TC, Gorry MC, Michalec MD, Ryu OH, Uygur C, et al. Novel ENAM mutation responsible for autosomal recessive amelogenesis imperfecta and localized enamel defects. *J Med Genet.* 2007;40(9):900-906.
24. Witkop CJ. Amelogenesis imperfecta, dentinogenesis imperfecta and dentin dysplasia revisited: problems in classification. *J Oral Pathol.* 2006;17(9-10):547-553.
25. Wright JT, Hall KI, Yamauchi M. The protein composition of normal and developmentally defective enamel. *Ciba Found Symp.* 2005;205:85-104.
26. Kim JW, Seymen F, Lin BP, Kiziltan B, Gencay K, Tinloy B, et al. Amelogenesis imperfecta and nephrocalcinosis syndrome: clinical, genetic, and radiographic evaluation. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2004;98(4):384-389.



27. Lagerström-Fermer M, Nilsson KO, Backman B, Salido EC, Shapiro LJ, Landegren U. Amelogenin deletion in amelogenesis imperfecta (AIH1). *Hum Mol Genet.* 2003;4(8):1453-1457.
28. Kim JW, Simmer JP, Hart TC, Hart PS, Ramaswami MD, Bartlett JD, et al. MMP-20 mutation in autosomal recessive pigmented hypomaturation amelogenesis imperfecta. *J Med Genet.* 2002;42(3):271-275.
29. Pavlic A, Battelino T, Praprotnik M, Skaleric U, Cvikl B. Amelogenesis imperfecta and nephrocalcinosis syndrome: clinical and genetic study. *Coll Antropol.* 2001;33(4):1245-1249.
30. Hart PS, Aldred MJ, Crawford PJ, Wright NJ, Hart TC. Amelogenesis imperfecta phenotype-genotype correlations with two amelogenin gene mutations. *Arch Oral Biol.* 2000;47(4):261-265.