

ANTIFUNGAL ACTIVITY OF MICROBIAL METABOLITES AGAINST *Colletotrichum gloesporioides*

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ABSTRACT

The objective of the present study was to evaluate *in vitro* the concentrated fraction of secondary metabolites produced by the *Bacillus bacteria cereus* against the phytopathogenic fungus *Colletotrichum gloesporioides*. Using the agar diffusion technique and the characterization of secondary metabolites using gas chromatography coupled to mass spectrometry (GC-MS) technique. **Results:** The microbial metabolites extracted from *Bacillus cereus* presented a 48% inhibition percentage and geraniol, geranate and sulcatone were identified, which presented *in vitro* antagonism against *C. gloesporioides* phytopathogen of agricultural interest at regional, national and international levels. **Conclusion.** The production of microbial secondary metabolites in the future may be an environmentally friendly biological alternative to replace agrochemicals.

Keywords: Bacteria, microbial extract, inhibition, phytopathogen.

1. INTRODUCTION

According to Whitelaw et al., (2007), the genus *Colletotrichum* causes diseases in virtually all agricultural crops in the world. The typical symptoms of infection by *Colletotrichum* are called 'anthracnose', which is characterized by necrotic sinking of the tissue where masses of conidia are produced within an acervulus (Freeman et al., 2000). Anthracnose occurs in developing and mature plant tissues, affecting fruits during their development in the field, as well as mature fruits during storage (Prusky et al., 2000).

However, avocado crops are very susceptible to the attack of pathogenic fungi that rot tissues responsible for important diseases such as anthracnose, characterized by dark, sunken, circular, ellipsoidal lesions, becoming a limiting factor in production. The disease has been controlled through the use of fungicides such as copper oxychloride and captafol, which are not easily eliminated by standard packaging procedures, leading to the rejection of 11.5 to 19.4% of the fruit destined for export (Trinidad-Ángeles et al., 2017), as well as the application of the fungicide prochloraz, which has been restricted for being a priority contaminant as a probable human carcinogen (Twizeyimana et al., 2013).

For this, the use of beneficial microbial inoculants has been promoted (Kumar et al., 2021). Previous studies have used organisms such as plant growth promoting bacteria (PGPR), which play an important role in increasing plant biomass, carbohydrate and protein content, photosynthetic pigments, modulating the expression of metabolites, outperforming exogenous hormones applied to improve characteristics associated with low yields caused by stress (Ismail et al., 2021), generating multiple benefits in plants after inoculation through various biological mechanisms, becoming an alternative to replace agrochemicals used to counteract disease management and consequently cause environmental and serious damage to human health (Toghueo, 2020). Within the PGPR, *Bacillus* have been studied *sp* recognized as safe organisms in food application and development of phytosanitary products (Obianom&Sivakumar, 2018) with great metabolic versatility that allows it to carry out biological control of pests and diseases by different mechanisms (Villarreal-Delgado et al., 2018). Studies reported by Shimshoni et al. (2020) indicate that *B. cereus*, *B. licheniformes* and *B. subtilis* produce antibiotics and other inhibitory substances against post-harvest pathogens of avocado. Likewise, microorganisms (*Glomus spp*, *Trichoderma spp*, *Bacillus spp*, *Pseudomonas spp*) in an avocado seedling

production system reporting favorable effects of microorganisms on seedling growth and nutrient assimilation (Sotomayor *et al.*, 2022). The objective of this study was to evaluate *in vitro* the fraction of secondary metabolites produced by *B. cereus* against the fungus *Colletotrichum gloeosporioides*.

2. MATERIALS AND METHODS

Antagonist strain. The strain used in this study is a rhizospheric bacterium associated with avocado and banana crops with plant growth promoting activity molecularly identified as *Bacillus cereus*, which was activated and purified on nutrient agar medium.

Pathogenic fungus. The fungus used for the antifungal tests was isolated from avocado crops (*Persea americana*) in Montes de María, Department of Sucre. This phytopathogen was isolated by the Agricultural Bioprospecting Research Group in the Microbiological Research Laboratory of the University of Sucre.

Inhibitory activity of bacterial extracts against *Colletotrichum gloeosporioides*. The ability of the bacteria to inhibit the growth of the phytopathogenic fungus *Colletotrichum gloeosporioides*, was carried out using confrontation tests and qualitative estimation. For this purpose, the fungus was sown on potato dextrose agar (PDA) and allowed to grow for three days at room temperature. A combination of PDA and R₂A media was prepared in a 1:1 ratio to inoculate *B. cereus*. After 2 days of incubation of the bacteria in the PDA-R₂A medium at 27 °C, the phytopathogenic fungus was inoculated by the direct sowing method. Each isolate was cut with a punch from the periphery of a 10-day-old colony, approximately 6 mm in diameter in growth area (Pérez *et al.*, 2011). The dishes were incubated at 30 °C for 7 days and absolute control was used without any type of treatment.

Liquid fermentation. *B. cereus* was taken, which was inoculated in Luria Bertani (LB) liquid medium and left shaking for 48 hours at 30°C. After this time, 1 ml of bacterial growth was taken and placed in 400 ml of 3s medium. The medium was left under constant shaking for 72 hours (Ortiz-Galeana *et al.*, 2018).

Extraction and characterization of metabolites produced by *B. cereus*. 100 ml of the fermented medium was taken and centrifuged at 7000 rpm for 45 minutes. 80 ml of ethyl acetate was added to each filtrate, then the organic fraction was collected and concentrated using a rotary evaporator. The concentrate was analyzed by gas chromatography coupled to mass spectrometry (GC-MS).

Antifungal activity of the extract. To carry out the antifungal activity, the agar diffusion technique was used, according to that proposed by Islam *et al.* (2012). The determination of the antifungal activity of the extract was carried out in PDA medium (20 mL) with the help of a sterile stainless-steel punch. A well was made and then 60 µL of extract from each medium and strain was added, followed by the placement of two agar blocks (0.5 cm in diameter) with the pathogenic fungus at the same distance. The dishes were incubated at 28° ± 3° C for 14 days and the inhibitory activity of each medium and strain was expressed as the percentage of inhibition of fungal growth, compared to the negative control (only ethyl acetate was inoculated in the well).

The percentage of growth inhibition was determined by the following equation:

$$\text{Inhibition (\%)} = \left[\frac{R_c - R_b}{R_c} \times 100 \right],$$

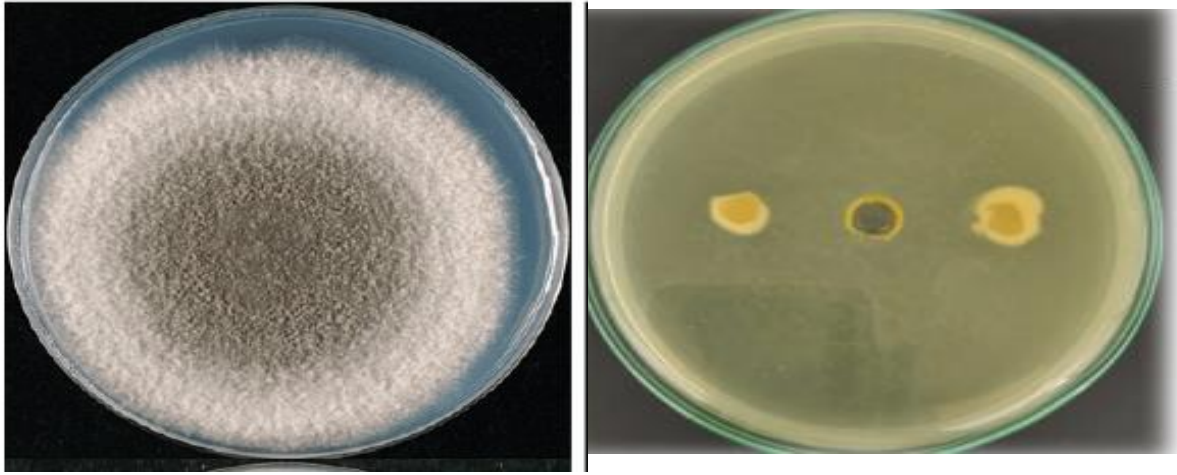
Where R_c is the radius of the control and R_b is the radius of the fungal colony interacting with the antagonistic bacteria (Rahman, 2007). Each treatment had 5 replicates.

Extraction and characterization of metabolites produced by *B. cereus*. 100 ml of the fermented medium was taken and centrifuged at 7000 rpm for 45 minutes. To each filtrate, 80 ml of ethyl acetate was added, then the organic fraction was collected and concentrated using a rotary evaporator. The concentrate was analyzed by gas chromatography coupled to mass spectrometry (GC-MS).

Statistical analysis. A completely randomized design was applied to determine the percentage inhibition of secondary metabolites. Also, Duncan's multiple range test was applied to establish the difference (p<0.05) in the percentage of inhibition against *B. cereus* with respect to the control. Five replicates per treatment were performed. Data were analyzed in the free version of InfoStat software.

3. RESULTS AND DISCUSSION

Figure 1 shows the inhibitory activity of *Bacillus cereus* cell suspension against *Colletotrichum gloeosporioides* with an inhibition percentage of 86,19% with respect to the control (figure 1), possibly due to the antagonistic action regulated by antibiosis, characteristic of the bacterial strain against pathogens.



Colletotrichum gloeosporioides (Control) **Bacteria extract from *Bacillus cereus***
Figure 1. In vitro, test of the inhibitory activity of bacterial extracts of *B. cereus* against *C. gloeosporioides*.

Figure 2 shows the inhibition efficiency of 15 treatments of secondary metabolite-type microbial extracts produced by *B. cereus*. The inhibition ranges of the secondary metabolite-type extracts ranged from 19.41 ± 86.19 %, showing irregular growth of the plant pathogen (*C. gloeosporioides*). It was also observed that the lower the concentration of the microbial extract (5.0 %), the lower the percentage of fungal inhibition (19.41 %), compared to the treatments with concentration (80 %), the higher the fungal inhibition (80.09, 86.14 and 86.19 %) (Figure 3).

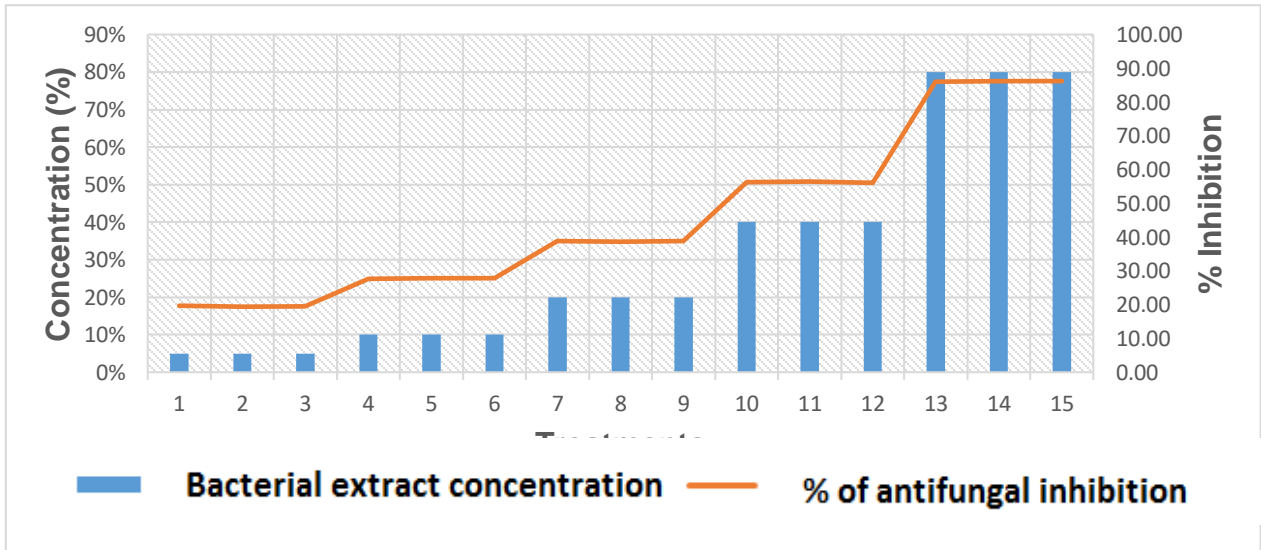


Figure 2. Percentage inhibition of secondary metabolite-like microbial extracts of *Bacillus cereus* against *Colletotrichum gloeosporioides*.

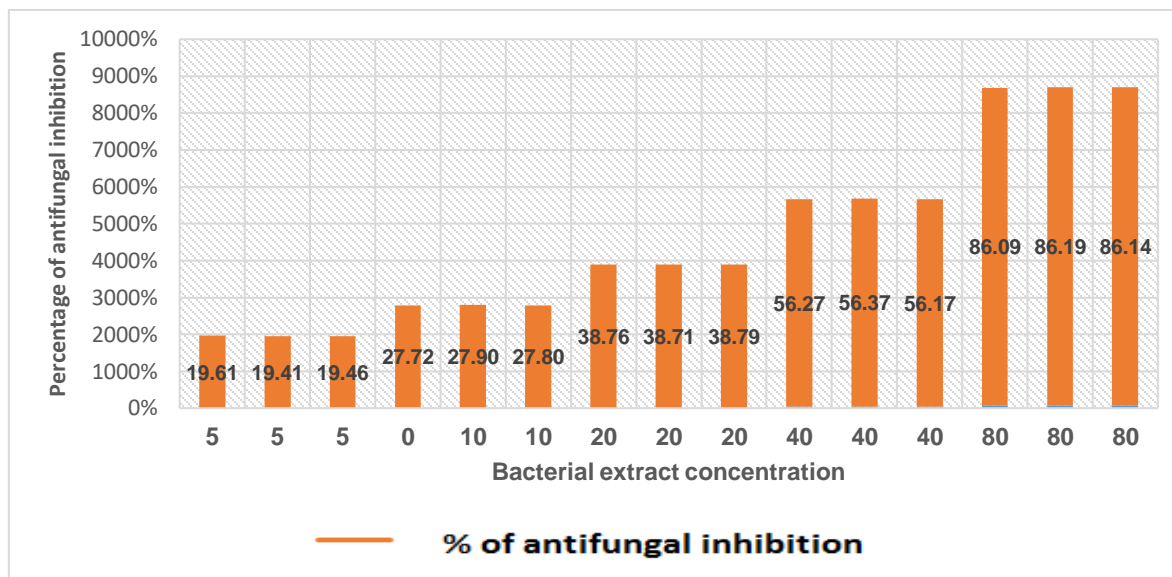


Figure 3. Percentage antifungal inhibition of different concentrations of secondary metabolite-type microbial extracts of *Bacillus cereus* against *Colletotrichum gloesporioides*.

The *Bacillus cereus* group contains closely related bacteria exhibiting a broad range of properties associated with their diverse lifestyles (Jensen et al., 2003). Some members have known biotechnological applications, while others may pose a potential health risk (Ehling-Schulz et al., 2019).

According to the results of the analysis of the bacterial extracts from *Bacillus cereus* by chromatography technique, 42 compounds were identified, of which geraniol showed an area percentage (30%), sulcatone (20%) and geranate (26%), these are the ones that are possibly producing the inhibition activity against pathogens such as *Colletotrichum gloesporioides*.

The biological control activity exhibited by *Bacillus* strains is possibly often due to the production of volatile organic compounds (VOCs), which can directly protect plants against plant pathogens and/or indirectly through the induction of plant resistance (Lim et al., 2017; Xing et al., 2018). These compounds were chemically classified into groups of esters, alkanes, alkenes, alkynes, alcohols, terpenoids, aldehydes, ketones and sulphur-containing compounds (Avalos, 2018). Previous studies have illustrated the antimicrobial activity of violacein produced by *Bacillus cereus* against various pathogens (Im et al., 2017; Cauz et al., 2019).

The genus *Bacillus* and its most representative species: *B. subtilis*, *B. brevis*, *B. cereus*, *B. pumilus*, *B. licheniformis* and *B. amyloliquefaciens* offer an alternative biological control thanks to their ability to be present in the soil as they are able to produce spores that give them resistance to different conditions such as desiccation, heat, organic solvents and UV irradiation (Melnick et al., 2008; Alvarez & Sánchez, 2016). The work carried out by Santander (2012) showed that the consortium of *B. subtilis* and *T. harzianum* increased the inhibitory activity of *C. gloesporioides* in mango fruits (*Mangifera indica* L) by evaluating the percentage of inhibition and sporulation. *B. cereus* is also able to produce protease and chitinase enzymes with activity on the chitin component of the fungal cell wall (Wang et al., 2009), which make them suitable for use as a biological control agent for plant pathogens (Layton et al., 2011).

Sansinenea & Ortiz, (2011), report that part of the studies on the production of microbial metabolites have emphasized the genus *Bacillus*, within the metabolites there are four families: bacteriocins, lantibiotics, lipopeptides and polyketides. Many of the species belonging to this genus have become biotechnological tools in order to control pathogens that affect the yield of crops of agricultural interest and also to reduce the application of chemical pesticides (Chen et al., 2020; Daraz et al., 2021). This idea is confirmed by the production and characterization of metabolites produced by *B. subtilis* against *Fusarium* sp., the results obtained allow the detection of a controlling effect against the fungus through the production of Iturin A with an inhibition percentage of 70% (Daraz et al., 2021).

According to Ariza & Sánchez, (2012), the bacterial phylotype identified as *Bacillus velezensis* has demonstrated the ability to inhibit *Alternaria solani* in vitro by producing metabolites such as iturins and acetophenone, which significantly affected the hyphae by means of perforations and swelling where the microbial metabolite acted. In addition, optimal metabolite production by *B. subtilis* is found at a pH between 6.0-8.0 and at a temperature between 25°C and 30°C. The results presented by Sidorova, et al., (2020); Jin et al., (2020); Shew et al., (2019); Kang & Hwang, (2018) according to the chromatographic analysis reports the synthesized metabolites as surfactin and Iturin which showed inhibition against *Fusarium* sp. This data can give us an indication for the manufacture of effective fungicides to protect crops from any pathogen.

According to studies by Bard et al., (1988); Stashenko et al., (2014); Linde et al., (2018), geraniol, geranate and sulcatone are monoterpenes that act on the cell membrane of pathogens generating an imbalance in the lipid membrane which causes an increase in the fluidity of the membrane causing a loss of potassium ions. On the other hand, Singh et al., (2016). Several studies have shown that geraniol is present in the extracted oil *Clinopodium pulchellum* (Kunth) (Lamiaceae) which showed inhibitory activity with *Candida albicans* (Tapia Manrique, 2018), as well as an inhibitory effect against the formation of pathogenic fungal biofilms and hyphal morphogenesis, destroying the cell wall by regulating the low ATPase activity on plasma membrane and reducing ergosterol levels.

4. CONCLUSIONS

In this study, secondary metabolites produced by *Bacillus cereus* inhibited *Colletotrichum gloesporioides*, which affect avocado crop yield, in the range of $19,41 \pm 86,19\%$ in vitro. The large-scale production of metabolites from the genus *Bacillus* can be considered a biological alternative for the management of different diseases in the field and the environmental sustainability of agricultural ecosystems.

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6. AUTHOR CONTRIBUTION. Alexander Perez Cordero: experiment execution, data analysis. Donicer Montes V and Yelitza Aguas M, conceptualization, writing - revision and editing. All authors have read and approved the manuscript.

7. CONFLICT OF INTEREST. All the authors of the manuscript declare that they have no conflict of interest

8. REFERENCES

- Álvarez, E. C., & Sánchez, L. C. (2016). Evaluación del crecimiento de cuatro especies del género *Bacillus* sp., primer paso para entender su efecto biocontrolador sobre *Fusarium* sp. *Nova*, *14*(26), 53-62.
- Avalos M, van Wezel GP, Raaijmakers JM, Garbeva P. Healthy scents: microbial volatiles as new frontier in antibiotic research? *Current opinion in microbiology*, 2018; 45:84-91. Doi: <https://doi.org/10.1016/j.mib.2018.02.011>.
- Ariza, Y., & Sánchez, L. (2012). Determinación de metabolitos secundarios a partir de *Bacillus subtilis* efecto biocontrolador sobre *Fusarium* sp. *Nova*, *10*(18), 149-155.
- Bard, M., Albrecht, M. R., Gupta, N., Guynn, C. J., & Stillwell, W. (1988). Geraniol interferes with membrane functions in strains of *Candida* and *Saccharomyces*. *Lipids*, *23*(6), 534-538.
- Cauz, A. C. G., Carretero, G. P. B., Saraiva, G. K. V., Park, P., Mortara, L., Cuccovia, I. M., Brocchi, M., Gueiros-Filho, F. J., 2019. Violacein targets the cytoplasmic membrane of bacteria. *ACS Infect Dis.* *5*, 539-549. <https://doi.org/10.1021/acsinfecdis.8b00245>.
- Chen, K., Tian, Z., He, H., Long, C. A., & Jiang, F. (2020). *Bacillus* species as potential biocontrol agents against citrus diseases. *Biological Control*, *151*, 104419.
- Daraz, U., Li, Y., Sun, Q., Zhang, M., & Ahmad, I. (2021). Inoculation of *Bacillus* spp. modulate the soil bacterial communities and available nutrients in the rhizosphere of vetiver plant irrigated with acid mine drainage. *Chemosphere*, *263*, 128345.
- Freeman, S.; Minz, D.; Jurkevitch, E.; Maymon, M.; Shabi, E. Molecular analyses of *Colletotrichum* species from almond and other fruits. *Phytopathology*, Saint Paul, v.90, n.6, p.608-614, 2000.
- Islam Md. R., Y. T. Jeong, Y. S. Lee, C. H. Song. (2012). *Isolation and Identification of Antifungal Compounds from Bacillus subtilis C9 Inhibiting the Growth of Plant Pathogenic Fungi*. *Mycobiology* *40*(1) : 59-66.

- Ismail, M. A., Amin, M. A., Eid, A. M., Hassan, S. E. D., Mahgoub, H. A., Lashin, I., ... & Fouda, A. (2021). Comparative Study between exogenously applied plant growth hormones versus metabolites of microbial endophytes as plant growth-promoting for *Phaseolus vulgaris* L. *Cells*, *10*(5), 1059.
- Jin, P., Wang, H., Tan, Z., Xuan, Z., Dahar, G. Y., Li, Q. X., ... & Liu, W. (2020). Antifungal mechanism of bacillomycin D from *Bacillus velezensis* HN-2 against *Colletotrichum gloeosporioides* Penz. *Pesticide Biochemistry and Physiology*, *163*, 102-107.
- Im, H., Choi, S. Y., Son, S., Mitchell, R. J., 2017. Combined application of bacterial predation and violacein to kill polymicrobial pathogenic communities. *Sci Rep.* *7*, 14415. <https://doi.org/10.1038/s41598-017-14567-7>.
- Kang, Y., & Hwang, I. (2018). Glutamate uptake is important for osmoregulation and survival in the rice pathogen *Burkholderia glumae*. *PLoS one*, *13*(1), e0190431.
- Kumar, R., Saurabh, K., Kumawat, N., Sundaram, P. K., Mishra, J. S., Singh, D. K., ... & Bhatt, B. P. (2021). Sustaining productivity through integrated use of microbes in agriculture. In *Role of Microbial Communities for Sustainability* (pp. 109-145). Springer, Singapore.
- Layton, C., Maldonado, E., Monroy, L., Ramírez, L. C. C., & Leal, L. C. S. (2011). *Bacillus* spp.; perspectiva de su efecto biocontrolador mediante antibiosis en cultivos afectados por fitopatógenos. *Nova*, *9*(16), 177-187.
- Linde, G. A., Colauto, N. B., Albertó, E., & Gazim, Z. C. (2016). Quimiotipos, extracción, composición y aplicaciones del aceite esencial de *Lippia alba*. *Revista Brasileira de Plantas Mediciniais*, *18*, 191-200.
- Lim SM, Yoon MY, Choi GJ, Choi YH, Jang KS, Shin TS, et al. Diffusible and volatile antifungal compounds produced by an antagonistic *Bacillus velezensis* G341 against various phytopathogenic fungi. *Plant Pathol. J.* 2017; *33*(5): 488-498. Doi: 10.5423/PPJ.OA.04.2017.0073.
- Melnick, R. L., Zidack, N. K., Bailey, B. A., Maximova, S. N., Guiltinan, M., & Backman, P. A. (2008). Bacterial endophytes: *Bacillus* spp. from annual crops as potential biological control agents of black pod rot of cacao. *Biological control*, *46*(1), 46-56.
- Obianom, C., & Sivakumar, D. (2018). Differential response to combined prochloraz and thyme oil drench treatment in avocados against the control of anthracnose and stem-end rot. *Phytoparasitica*, *46*(3), 273-281.
- Ortiz-Galeana, M. A., Hernández-Salmerón, J. E., Valenzuela-Aragón, B., los Santos-Villalobos, S., Rocha-Granados, M., & Santoyo, G. (2018). Diversidad de bacterias endófitas cultivables asociadas a plantas de arándano (*Vaccinium corymbosum* L.) cv. Biloxi con actividades promotoras del crecimiento vegetal. *Chilean journal of agricultural & animal sciences*, *34*(2), 140-151.
- Pérez, A., Rojas, J., Chamorro, L., & Pérez, K. (2011). Evaluación in vitro de actividad inhibitoria de extractos vegetales sobre cepas de hongos del género *Colletotrichum* sp. *Acta Agronómica*, *60*(2), 158-164.
- Prusky, D.; Freeman, S.; Dickman, M. ***Colletotrichum. Host specificity, pathology, and host-pathogen interaction.*** . St. Paul: American Phytopathological Society Press, 2000. 393 p.
- Rahman, M. A., Kadir, J., Mahmud, T. M. M., Rahman, R. A., & Begum, M. M. (2007). Screening of antagonistic bacteria for biocontrol activities on *Colletotrichum gloeosporioides* in papaya. *Asian Journal of Plant Sciences*.
- Sansinenea, E., & Ortiz, A. (2011). Secondary metabolites of soil *Bacillus* spp. *Biotechnology letters*, *33*(8), 1523-1538.
- Santander, A. J. (2012). Uso de *Trichoderma harzianum* Rifai y *Bacillus subtilis* (Ehrenberg) Cohn para el control de (*Colletotrichum gloeosporioides* penz.) causante de la antracnosis en mango (*Mangifera indica* L.). *Facultad de Agronomía Postgrado en Agronomía-Universidad Central de Venezuela*, 5-26.
- Shew, A. M., Durand-Morat, A., Nalley, L. L., Zhou, X. G., Rojas, C., & Thoma, G. (2019). Warming increases Bacterial Panicle Blight (*Burkholderia glumae*) occurrences and impacts on USA rice production. *PLoS one*, *14*(7), e0219199.
- Shimshoni, J. A., Bommuraj, V., Chen, Y., Sperling, R., Barel, S., Feygenberg, O., ... & Alkan, N. (2020). Postharvest fungicide for avocado fruits: antifungal efficacy and peel to pulp distribution kinetics. *Foods*, *9*(2), 124.
- Sidorova, T. M., Asaturova, A. M., Homyak, A. I., Zhevnova, N. A., Shternshis, M. V., & Tomashevich, N. S. (2020). Optimization of laboratory cultivation conditions for the synthesis of antifungal metabolites by *Bacillus subtilis* strains. *Saudi journal of biological sciences*, *27*(7), 1879-1885.
- Singh, S., Fátima, Z. y Hameed, S. (2016). Los conocimientos sobre el modo de acción del geraniolmonoterpenoide a base de hierbas anticáncida revelan la interrupción de múltiples mecanismos MDR y atributos de virulencia en *Candida albicans*. *Archivos de microbiología*, *198* (5), 459-472.

- Sotomayor, A., Mejía, P., Morocho, D., Gaona, P., Viteri, P., Medina, L., & Viera, W. (2022). Consorcios microbianos aplicados en un sistema de producción de plántulas de aguacate cultivar “Criollo”. *Manglar*, 19(1), 15-23.
- Stashenko, E. E., Martínez, J. R., Durán, D. C., Córdoba, Y., & Caballero, D. (2014). Estudio comparativo de la composición química y la actividad antioxidante de los aceites esenciales de algunas plantas del género *Lippia* (Verbenaceae) cultivadas en Colombia. *Revista de la Academia Colombiana de Ciencias Exactas, Físicas y Naturales*, 89-105.
- Tapia Manrique, E. R. (2018). Composición química, actividad antioxidante y antiCandidaalbicans del aceite esencial de *Clinopodium pulchellum* (Kunth) Govaerts “panizara”.
- Toghueo, R. M. K. (2020). Bioprospecting endophytic fungi from *Fusarium* genus as sources of bioactive metabolites. *Mycology*, 11(1), 1-21.
- Trinidad-Ángel, E., Ascencio-Valle, F. D. J., Ulloa, J. A., Ramírez-Ramírez, J. C., Ragazzo-Sánchez, J. A., Calderón-Santoyo, M., & Bautista Rosales, P. U. (2017). Identificación y caracterización de *Colletotrichum* spp. causante de antracnosis en aguacate Nayarit, México. *Revista mexicana de ciencias agrícolas*, 8(SPE19), 3953-3964.
- Twizeyimana, M., Förster, H., McDonald, V., Wang, D. H., Adaskaveg, J. E., & Eskalen, A. (2013). Identification and pathogenicity of fungal pathogens associated with stem-end rot of avocado in California. *Plant Disease*, 97(12), 1580-1584.
- Villarreal-Delgado, M. F., Villa-Rodríguez, E. D., Cira-Chávez, L. A., Estrada-Alvarado, M. I., Parra-Cota, F. I., & Santos-Villalobos, S. D. L. (2018). El género *Bacillus* como agente de control biológico y sus implicaciones en la bioseguridad agrícola. *Revista mexicana de fitopatología*, 36(1), 95-130.
- Wang, S. L., Chao, C. H., Liang, T. W., & Chen, C. C. (2009). Purification and characterization of protease and chitinase from *Bacillus cereus* TKU006 and conversion of marine wastes by these enzymes. *Marine biotechnology*, 11(3), 334-344.
- Whitelaw, W.M.A.; Curtis, S.J.; Huang, R.; Steel, C.C.; Blandchard, C.L.; Roffey, P.E. Phylogenetic relationships and pathogenicity of *Colletotrichum acutatum* isolates from grape in subtropical Australia. **Plant Pathology**, Chichester, v.56, n.3, p.448-463, 2007.
- Xing M, Zheng L, Deng Y, Xu D, Xi P, Li M, et al. Antifungal activity of natural volatile organic compounds against litchi downy blight pathogen *Peronophythora litchi*. *Molecules*, 2018; 23:358. Doi: <https://doi.org/10.3390/molecules23020358>.