

## Polyphenolic composition of grape stems

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### Abstract

This study is focused on the study of polyphenolic compounds in grape stems as by-product of winemaking industry. Two white varieties of Grüner Veltliner and Sauvignon and two red varieties of Blauer Portugieser and Cabernet Moravia were selected for the study. Antioxidant activity, concentration of total polyphenols and concentration of individual phenolic compounds were determined. The results show a higher concentration of polyphenols and higher values of antioxidant activity in red varieties. The Blauer Portugieser variety contained the highest concentrations of syringic acid 1.346 mg.L<sup>-1</sup>, caffeic acid 20 mg.L<sup>-1</sup>, ferulic acid 1.192 mg.L<sup>-1</sup>, coumaric acid 3.231 mg.L<sup>-1</sup>, trans-resveratrol 14.195 mg.L<sup>-1</sup>, catechin 79.314 mg.L<sup>-1</sup> and epicatechin 33.205 mg.L<sup>-1</sup>. Cabernet Moravia contained the highest concentration of protocatechuic acid 1.201 mg.L<sup>-1</sup>, the Sauvignon variety reached the highest concentration of gallic acid 4.015 mg.L<sup>-1</sup> and hydroxybenzoic acid 0.076 mg.L<sup>-1</sup>. The highest values of alpha-amino acids were determined in the Blauer Portugieser variety 165.3 mg L<sup>-1</sup> and the lowest in the Grüner Veltliner variety 33.3 mg L<sup>-1</sup>. The highest concentration of ammonia nitrogen was 214 mg L<sup>-1</sup> for the Blauer Portugieser variety and the lowest concentration of ammonia nitrogen was measured in Cabernet Moravia 35.7 mg L<sup>-1</sup>.

**Keywords:** antioxidant activity; grape stems; polyphenols; winemaking by-products

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### Introduction

Grape polyphenolics vary in chemical structure and activity and may be fundamentally categorized into two major classes: flavonoids and nonflavonoids. Flavonoids, the most abundant polyphenolics in grape, are distributed throughout the peel, seed, and stem, and include anthocyanins, proanthocyanidins (procyanidins and prodelphinidins), and flavan-3-ols (Garrido-Banuelos *et al.*, 2019). In contrast, hydroxycinnamic acids, the most abundant non-flavonoids in wine, include caftaric acid and coutaric acid (Lu and Foo, 1999). Most of these polyphenolic compounds occur as glycosylated derivatives in plants and foods and undergo enzymatic transformations in the gut before intestinal absorption (Bang *et al.*, 2015). In vinification, bioactive polyphenolic compounds are partially extracted while the majority remain as glycosides embedded in the grape peel, pulp, or seed (Chafer *et al.*, 2005). Additionally, the amount of polyphenols released into the final wine product greatly depends on the fermentation process, suggesting that an insufficient extraction technique

prevents the liberation of phytochemicals that are essentially confined in grape cell walls and pulp cell vacuoles (Yacco *et al.*, 2016).

For this reason, the waste from wine production is also an important source of phenolic compounds with non-negligible antioxidant capacity (Dineiro García *et al.*, 2009). Grape stems consist of polyphenols: flavonoids, stilbenoids and proanthocyanidins (Makris *et al.*, 2008; Karvela *et al.*, 2009), and thus are considered to be a significant source of antioxidants (Anastasiadi *et al.*, 2009).

Investigation into the application of antioxidants, such as polyphenols, has been increasing due to their health benefits (Di Donato *et al.*, 2017). In recent years, special attention has been given to the isolation of natural compounds from waste materials of the food and wine industry and their reuse or conversion into new products (Maier *et al.*, 2009). Bioactive compounds, such as polyphenols, can be obtained from grape stems by using different extraction methods. Optimal conditions for the extraction, including the type of solvent, process duration and temperature, determine the performance and quality of the obtained compounds (Pintać *et al.*, 2018).

In recent years, new technologies for more efficient and environmentally friendly extraction, such as the use of ultrasound, microwaves and pulsed-electric fields, have been studied. Most of them are expensive, difficult to scale up for larger amounts of extracted material or energy intensive (Okolie *et al.*, 2019). Eco-friendly technologies, such as hydrothermal treatment, have some advantages due to the absence of organic solvents and related corrosion problems, as well as being easy to operate and cost effective (Sepúlveda *et al.*, 2018). Moreover, some studies reported higher bioactive content after hot-water extractions compared to solvent extraction (Kabir *et al.*, 2015).

On a laboratory scale and for a smaller volume of studied material, classical extraction methods using organic solvents are still used. Many studies have used response surface methodologies (RSM) to optimise polyphenolic extraction conditions. However, the extraction conditions for RSM are generally limiting for scale-up and industrial applications, and the polyphenolic content is usually evaluated by generic methods such as total phenolic, flavonoid, tannin, total flavone content and antioxidant/antiradical assays (Di Donato *et al.*, 2017).

The major goal of the present study was to characterise individual polyphenolic compounds in grape stalks in white and blue grape varieties.

## Materials and Methods

### *Material*

The aim of the study was the evaluation of polyphenols as antioxidants in grape stems of the varieties *Vitis vinifera* L., namely the varieties Grüner Veltliner, Sauvignon blanc, Blauer Portugieser and Cabernet Moravia M-43. The material was obtained from the Rozlinky (Šakvice) vineyard in the Czech Republic. The age of all the vines from which the stems come is in the range of 10-12 years. The basic analytical parameters are in Table 1.

**Table 1.** Basic analytical parameters of grape must

Variety	Date of harvest	Sugar content [°NM]	pH	Total acidity [g.L <sup>-1</sup> ]
Gruner Veltliner	1.10.2019	21,50	3,28	7.80
Sauvignon	27.9.2019	22,80	3,15	8.10
Blauer Portugieser	5.10.2019	23,20	3,55	5.80
Cabernet Moravia	4.11.2019	23,80	3,68	4.50

#### *Extraction method*

The stems were crushed and homogenised for 20 seconds. Subsequently, 10 g of this homogenate was weighed and transferred quantitatively to a volumetric flask. Ninety ml of 75% methanol was used for the extraction. The extraction was carried out in the dark and cold on an IKA KS 260 Basic shaker for two hours. 50  $\mu\text{L}$  of 75% sulphur dioxide was added to the sample to prevent oxidation.

Individual spectrophotometric determinations were performed on a MIURA ONE automatic biochemical analyser (I.S.E. S.r.l.; Guidonia, RM, Italy). The samples were centrifuged and diluted 1:10 with water. To ensure objectivity of the results, all samples were measured three times, and results were reported as an average of these measurements. The individual methods were adapted to the analyser used, with incubation at 37 °C and incubation times adapted to the instrument's operating cycles.

For purposes of HPLC analysis, extracts were centrifuged and diluted 1:10 with 100 mM  $\text{HClO}_4$  and directly analysed. To ensure the objectivity of the results, all samples were measured three times, and results were reported as the average of these measurements.

#### *Antioxidant activity*

In each variant, the values of the antioxidant activity were observed. The procedure has been described previously (Sochor *et al.*, 2010a). During this procedure, a 150  $\mu\text{L}$  volume of the reagent (0.095 mmol 2,2-diphenyl-1-picrylhydrazyl, DPPH) was incubated with 15  $\mu\text{L}$  of the wine sample. The absorbance was measured at 505 nm for 10 minutes and the output ratio was calculated as a difference between the absorbance values measured at the 10<sup>th</sup> minute and the 2<sup>nd</sup> minute of the assay procedure. This determination was performed in triplicate in 2 mL samples. Antiradical activity was determined based on a calibration curve, using gallic acid (GA; 10-300  $\text{mg}\cdot\text{L}^{-1}$ ) as a standard. The results are expressed in equivalents of gallic acid (GA).

#### *Determination of reducing power (FRAP)*

The ferric reducing/antioxidant power (FRAP) method was modified to determine the reduction ability of the ferric ions. The pH of 198  $\mu\text{L}$  of base buffer containing 200 mmol sodium acetate was adjusted to 3.6 with acetic acid. Twelve  $\mu\text{L}$  of sample, 20  $\mu\text{L}$  of 20 mM  $\text{FeCl}_3$  solution and 20  $\mu\text{L}$  of 10 mmol TPTZ (2,4,6-tripyridyl-s-triazine) in 40 mM HCl were added. After 600 seconds, the absorbance at 620 nm was measured. The reducing power was calculated from the calibration curve using gallic acid (GA; 10-300  $\text{mg}\cdot\text{L}^{-1}$ ) as a standard (Pulido *et al.*, 2000).

#### *Determination of total flavanols*

Total flavanols concentration was determined using a method based on a reaction with p-dimethylaminocinnamaldehyde (DMACA). In contrast to the widely used reaction with vanillin, this method does not interfere with anthocyanins. In addition, it provides greater sensitivity and selectivity.

To 240  $\mu\text{L}$  reagent (0.1% DMACA and 300 mmol HCl in MeOH), 10  $\mu\text{L}$  sample was added, with a reaction time of 600 seconds. The absorbance at 620 nm was then measured. The concentration of total flavanols was determined based on a calibration curve using epicatechin as a standard (10-200  $\text{mg}\cdot\text{L}^{-1}$ ). The results are expressed as  $\text{mg}\cdot\text{L}^{-1}$  equivalents of catechin.

#### *Determination of alpha-amino nitrogen*

The primary amino groups are derivatised by ophthaldialdehyde and N-acetyl-L-cysteine (OPA/NAC) to form isoindoles in the basic medium. These derivatives were detected spectrophotometrically at 340 nm. The absorbance was proportional to the amount of primary amino nitrogen in the sample. Yeast non-assimilable amino nitrogen (e.g. acylated or blocked amines, proline and hydroxyproline) and ammonia nitrogen were not detected in this reaction. Therefore, yeast assimilable nitrogen compound (YANC) determination required independent assays of primary amino nitrogen and ammonia nitrogen. The analysis was performed using a Miura one® device (I.S.E. S.r.l. Via Luigi Einaudi, Italy), which is a spectrophotometer equipped with an

autosampler. Determination was performed in triplicate in 2 mL samples that were taken during the fermentation process and immediately frozen.

#### *Determination of ammonia nitrogen*

The enzyme glutamate dehydrogenase (GLDH) catalyses the condensation of ammonia and  $\alpha$ -ketoglutarate to L-glutamate with the concomitant oxidation of nicotinamide adenine dinucleotide (NADH)



The oxidation of NADH causes a decrease in absorbance at 340 nm, which is proportional to the amount of ammonia in the sample. The analysis was performed using the Miura one® device equipped with an autosampler as described above. Determination was performed in triplicate in 2 mL samples.

#### *Determination of individual antioxidant components*

Acetonitrile (ACN) was HPLC super gradient purified. Catechin, epicatechin, vanillic acid, protocatechuic acid, 4-hydroxybenzoic acid, gallic acid, syringic acid, p-coumaric acid, trans-resveratrol, tannic acid, coffee acid, ferulic acid, piceatannol, rutin, myricetin, quercetin, caemferol, isorhamnetin and perchloric acid were obtained from Sigma Chemical Co. (St. Louis, MO). Malvidin 3,5-diglucoside was purchased from Indofine Chemical Company, Inc. (Hillsborough, NJ). Other chemicals used were p.a. quality from a local supplier (Lachema, Penta). Cis-resveratrol and cis-piceid were prepared by photoisomerisation from their trans isomers.

#### Instrumentation:

Bim High Pressure System Shimadzu LC-10A, Controller system: SCL-10 Avp, 2 pumps: LC-10ADvp, Column thermostat with manual injection valve Rheodyne: CTO-10ACvp, DAD detector: SPD-M10Avp, Software: LCsolution

#### Separation conditions:

Column: Alltech Alltima HP C18 3  $\mu\text{m}$ ; 3 x 150 mm, separation temperature: 50 °C, sample injection volume: 20  $\mu\text{l}$ , mobile phase flow rate: 0.9 ml/min, mobile phase A: 15 mM HClO<sub>4</sub>, mobile phase B: 15 mM HClO<sub>4</sub>, 80% ACN

#### Gradient programme:

0.00 min 3% B, 3.00 min 6% B, 15.00 min 24% B, 18.00 min 30% B, 19.50 min 36% B, 21.00 min 48% B, 21.50 min 60% B, 22.00 min 60% B, 22.01 min 0% B, 23.99 min 0% B, 24.00 min 3% B

The total time between two samples was 27 minutes. Data in the range of 200–520 nm were recorded for 24 minutes.

Determination of individual components based on standard calibration curves: catechin; epicatechin (200 nm), vanilla acid; protocatechuic acid; 4-hydroxybenzoic acid (260 nm), gallic acid; syringic acid; cis-piceid; cis-resveratrol (280 nm), p-coumaric acid; trans-piceid; trans-resveratrol (310 nm), coffee acid; ferulic acid and its derivatives; piceatannol (325 nm), anthocyanins (520 nm).

The hydroxycinnamic acid derivatives were calibrated with the basic acids from which they were derived. Anthocyanins were calibrated to malvidin-3,5-diglucoside.

#### *Statistical analysis*

Statistical analyses and figures were generated using Excel 2016 software packages (manufactured by Microsoft Office, USA) and Statistica 10 statistical software (Copyright © StatSoft). Differences between means and contribution to the homogenous groups were determined using Fishers least significant differences test (LSD tests), level of significance was  $\alpha = 0.05$ .

## Results and Discussion

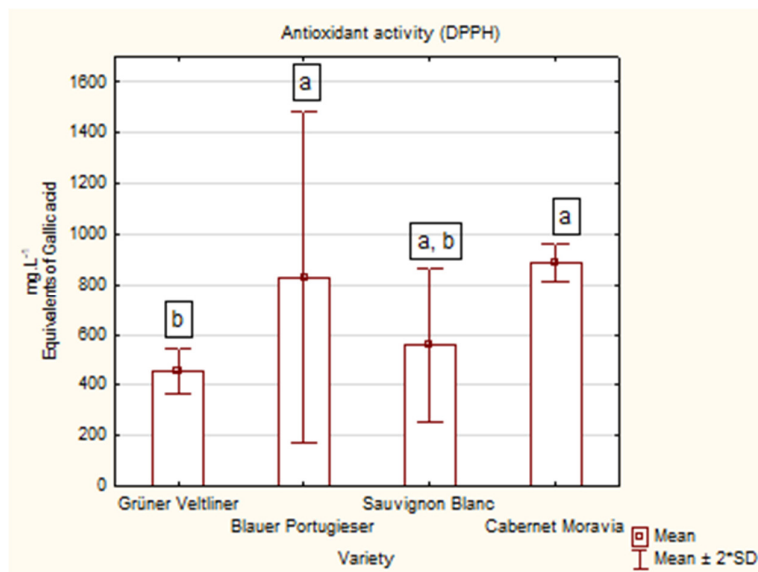
The first step in modern practices of winemaking is de-stemming resulting in a high fraction of waste material constituted by stems. Afterwards, grapes are pressed to obtain must and grape pomace (mainly skins and seeds) with another high fraction of waste. Grape stems constitute from 3 to 6% of the raw matter processed in a winery (Cabanis, 2000). All results are obtained after extraction with methanol and are expressed in  $\text{mg.L}^{-1}$  of methanol extract, so values may differ from those from other studies.

### *Determination of antioxidant activity*

It has been shown that grapevine extract is a promising alternative to sulphur dioxide, which is used in wineries for the so-called sulphurisation of wine to preserve it. The antioxidant and antimicrobial effects of sulphur dioxide are similar in nature to the effects of phenolic substances extracted from the wine cane. Use of wine cane extracts is not only economically but also ecologically advantageous. An important benefit of this finding could be the partial elimination of the use of sulphur dioxide, which is often associated with the emergence of certain diseases; therefore, quantitative limits for the use of sulphur dioxide have already been introduced in the past (Ruiz-Moreno *et al.*, 2015).

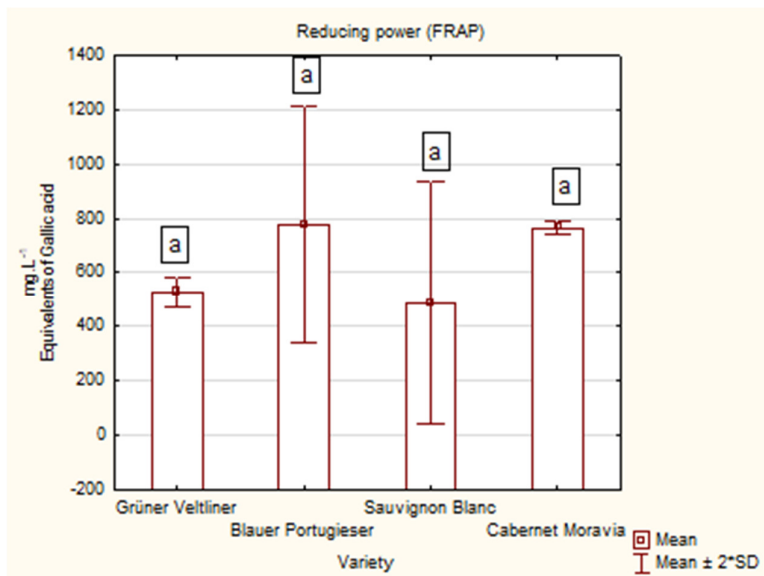
Figure 1 shows the values of antiradical activity by the DPPH method in the following varieties: Grüner Veltliner, Blue Portugieser, Sauvignon and Cabernet Moravia. The figure shows that the lowest values, on average  $458 \text{ mg.L}^{-1}$ , were measured for the Grüner Veltliner variety. Other values were quite comparable (in the range of  $705 \text{ mg.L}^{-1}$  to  $888 \text{ mg.L}^{-1}$ ).

Results of the determination of reducing power using gallic acid as a standard for the FRAP method are shown on the Figure 2 for the following varieties: Blauer Portugieser variety ( $775.6 \text{ mg.L}^{-1}$ ) and Cabernet Moravia ( $765.9 \text{ mg.L}^{-1}$ ), with lower values obtained for Grüner Veltliner ( $529 \text{ mg.L}^{-1}$ ) and Sauvignon ( $486.8 \text{ mg.L}^{-1}$ ).



**Figure 1.** Values of antioxidant activity (DPPH)

Results of this study show significant contribution of grape stems to total antioxidant activity of grapes. The higher values of antioxidant activity were recorded in the blue varieties: Blue Portugieser and Cabernet Moravia. The lower values were recorded in white varieties. The DPPH and FRAP methods show different results. Due to different extraction methods, extraction solvents or protocols and different grape varieties, the data are not comparable (Sochor *et al.*, 2010b).



**Figure 2.** Values of reducing power (FRAP)

The study by Leal *et al.* (2020) determined antioxidant activity of grape stems of five white grapevine varieties by two different methods, DPPH and ABTS. For both methods, there were significant differences between the samples. The lower antioxidant activity for both assays was  $0.63 \pm 0.03$  and  $0.37 \pm 0.03$  mmol Trolox g<sup>-1</sup> dw (dry weight), for ABTS and DPPH, respectively, with higher antioxidant activity for the ABTS method ( $1.17 \pm 0.09$  mmol Trolox g<sup>-1</sup> dw) and for the DPPH method ( $0.56 \pm 0.02$  mmol Trolox g<sup>-1</sup> dw).

A study by Domínguez-Perles (2014) presented lower values of antiradical activity in white varieties (Viosinho) using the ABTS method ( $0.049$  mmol Trolox g<sup>-1</sup> dw). The study was conducted to analyse the effect of extraction conditions on the extraction of total phenolics, flavonoids, ortho-diphenols and anthocyanins as well as to assess the ABTS<sub>+</sub> scavenging capacity, which were considered as response variables, since the antioxidant activity is positively correlated with the polyphenolic content (Anastasiadi *et al.*, 2012).

Results of the study by Domínguez-Perles (2014) suggested that optimal extraction of phenolic molecules may not reflect the highest antioxidant capacity. This may be explained by the potential for aqueous based solvents to contribute to solubilising a larger range of compounds, some of which may have little or no antioxidant activity (Anwar *et al.*, 2013).

The study by Silva *et al.* (2018) evaluated the antioxidant activity of skins, seeds and stems in the blue varieties, Preto Martinho and Touriga Nacional, using the ABTS and DPPH methods, expressed in  $\mu$ mol Trolox equivalent. Higher antioxidant capacity was found for the seed extracts, followed by stems and skins. In both varieties, a higher total polyphenols concentration was determined for the seed extracts when compared to the skin and stem extracts, which likely explains the higher antioxidant activity observed for the former.

This is in good agreement with previous studies that evaluated the antioxidant capacity and total polyphenols concentration of several grape varieties, and reported a high correlation between these parameters, pointing to the fact that the antioxidant activity of wines is mainly due to its phenolic compounds (Paixão *et al.*, 2007).

The study by Queiroza *et al.* (2017) evaluated the radical scavenging capacity of the isolated compounds from the grape stems, using the DPPH and ABTS methods. Higher antioxidant activity was determined in the coloured flavonoids (anthocyanins), malvidin-3-O-glucoside, malvidin-3-O-(6-O-caffeoyl)-glucoside and quercetin-3-O-glucoside.

*Determination of total polyphenols*

Recently, grapevines have been studied from the point of view of possible sources of polyphenolic substances because they possess antioxidant, antimicrobial and anticancerogenic activity. The total content of phenolic substances (reported in milligrams of gallic acid (GA) per gram of dry sample) in the extracts of the stems is different mainly depending on the variety of *Vitis vinifera* as well as the type of extract preparation. The influence of the extraction temperature was studied by Wenzel *et al.* (2015), and the ideal temperature ( $163 \pm 0.9$  to  $260 \pm 1.5$  °C) was determined.

Figure 3 shows the determination of total flavanol concentration on the basis of a calibration curve using epicatechin as a standard for the following varieties: Grüner Veltliner, Blauer Portugieser, Sauvignon and Cabernet Moravia. The results show that the highest concentrations of total flavanols were measured in the Blauer Portugieser ( $498.5 \text{ mg.L}^{-1}$ ) and Cabernet Moravia ( $449.4 \text{ mg.L}^{-1}$ ) varieties. The values for the white varieties of Grüner Veltliner and Sauvignon were lower, as in all previous determinations. The values of catechin equivalent were measured in the Grüner Veltliner variety ( $183.4 \text{ mg.L}^{-1}$ ) and in the Sauvignon variety ( $214.6 \text{ mg.L}^{-1}$ ).

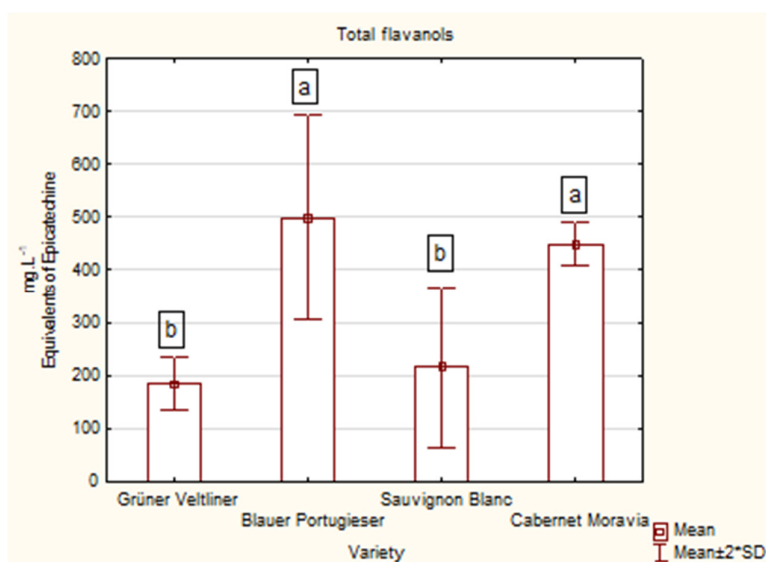


Figure 3. Concentration of total flavanols in grape stems expressed as  $\text{mg.L}^{-1}$  epicatechine

Llobera and Cañellas (2007) report the total content of phenolic substances extracted from the stems as  $116 \pm 2 \text{ mg GA g}^{-1}$  dry matter.

Anastasiadi *et al.* (2012) determined the total phenolic content of  $367\text{-}587 \text{ mg GA g}^{-1}$  dry matter, using a methanol:water:HCl (90:5:5 v/v) extraction mixture. The same methodology, using shags from other varieties of *Vitis vinifera*, was chosen by Sahnazidou *et al.* (2014), which found the total phenolic content of the extracts to be  $318\text{-}415 \text{ mg}^{-1}$  GA dry matter.

Although the content of polyphenols is associated with more colourful varieties, it turns out that even white varieties are an important source of polyphenols. The phenolic content of the grapes depends mainly on the variety and not on the colour of the grape (Yang *et al.*, 2009).

Leal *et al.* (2020) studied the phenolic content of white variety grape stem extracts. The total phenol content varied between  $94.71 \pm 4.65$  (Rabigato) and  $123.09 \pm 5.02$  (Malvasia Fina)  $\text{mg GA g}^{-1}$  dw. Also, Anastasiadi *et al.* (2012) reported a lower content of total phenols for the white varieties, Asyrtiko, Aidani and Athiri, with  $11.146$ ,  $7.220$  and  $4.808 \text{ mg GA g}^{-1}$  dw, respectively. Furthermore, high values were shown by Sahnazidou *et al.* (2014), with  $372 \text{ mg GA g}^{-1}$  dw for the white variety Assyrtiko. Concerning the ortho-diphenols, this content varied between  $80.62 \pm 3.69$  (Viosinho) and  $116.18 \pm 2.67$  (Malvasia Fina)  $\text{mg GA g}^{-1}$

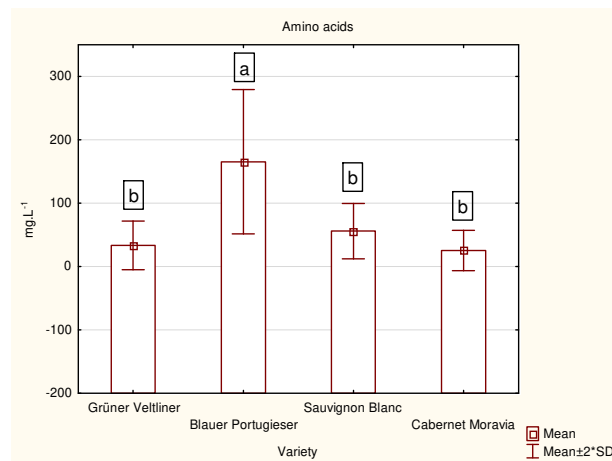
dw. The study carried out by Domínguez-Perles *et al.* (2014) showed values of 36.54 mg GA g<sup>-1</sup> dw for Viosinho.

These differences in phenolic composition can be explained by the specific characteristics of each variety, by the climate and biotic factors as well as by the viticultural practices (Portu *et al.*, 2018). The growing conditions have a great effect on the phenolic composition of plants. In lower altitude sites, samples of grape stems revealed a higher content of total phenols, ortho-diphenols and flavonoids. The low altitude region is characterised by some stress factors, such as thermal and water stress as well as the Atlantic influence on climate, resulting in abundant rain. This situation can explain the induction of secondary metabolites (Gouvinhas *et al.*, 2020). Despite these differences, grape stems contain a high concentration of phenolics compounds, in some cases, higher than the fruits (grapes) or the other by-products, revealing the importance of this waste as a rich source of phenolic compounds (Teixeira *et al.*, 2014).

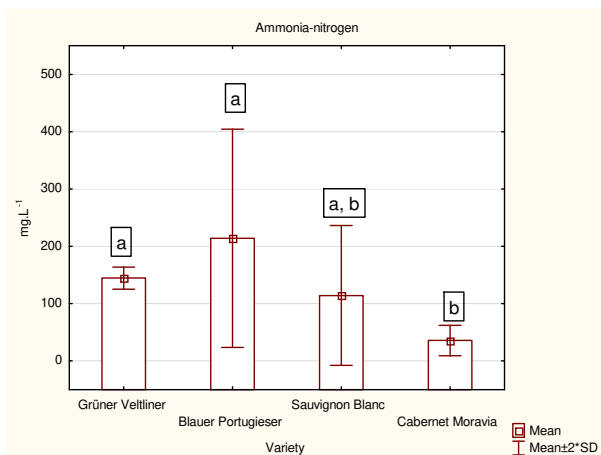
The content of polyphenols in wine also depends on the method of vinification. Their concentration can be increased by longer maceration, or in whole bunch winemaking processes (Baron *et al.*, 2017; Sanmartin *et al.*, 2019).

#### *Determination of yeast assimilable nitrogen*

In addition to phenolic substances, stems are also an important source of nitrogen substances, which serve as nutrition for yeast during fermentation. The amino acid and ammoniacal nitrogen content are shown in Figures 4 and 5. While the blue variety Blauer Portugieser had the highest content, another blue variety, Cabernet Moravia, had the lowest content. The results can be useful in whole-bunch winemaking.



**Figure 4.** Concentration of alpha-amino acids



**Figure 5.** Concentration of ammonia nitrogen

Figure 4 shows the determination of alpha-amino acids on the basis of a calibration curve, using glycine as a standard for the Grüner Veltliner, Blue Portugal, Sauvignon and Cabernet Moravia varieties. The highest values were determined for the Blauer Portugieser variety (165.3 mg.L<sup>-1</sup>). The second highest results were measured in the Sauvignon variety (55.8 mg.L<sup>-1</sup>). The lowest results were measured in the Grüner Veltliner variety (33.3 mg.L<sup>-1</sup> on average).

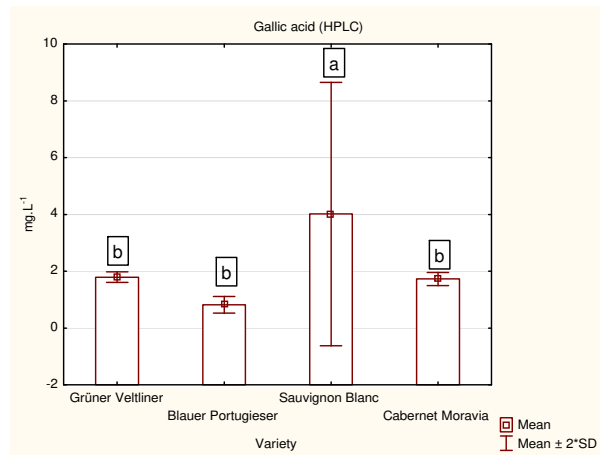
Figure 5 shows the ammonia nitrogen content of the Grüner Veltliner, Blauer Portugieser, Sauvignon and Cabernet Moravia varieties. The highest measured content was 214 mg.L<sup>-1</sup> for the Blauer Portugieser variety, with 144.5 mg.L<sup>-1</sup> ammoniacal nitrogen for the Grüner Veltliner variety, and 114.3 mg.L<sup>-1</sup> for the Sauvignon variety. The lowest content of ammonia nitrogen was measured in Cabernet Moravia (35.7 mg.L<sup>-1</sup><sup>3</sup>).

Sanmartin *et al.* (2019) studied the impact of co-fermentation on intact grape clusters and stalks. Both chemical and sensory profiles of wines were discussed soon after racking off as well as after 10 months of ageing with particular attention to phenolic and aromatic compounds. Our results show that co-fermentation of intact grape clusters and stalks can be profitably applied in order to improve the nutraceutical features of Syrah wines as well as to emphasise their aromatic expression, thus significantly speeding up their ageing phase.

#### *Determination of individual antioxidant components*

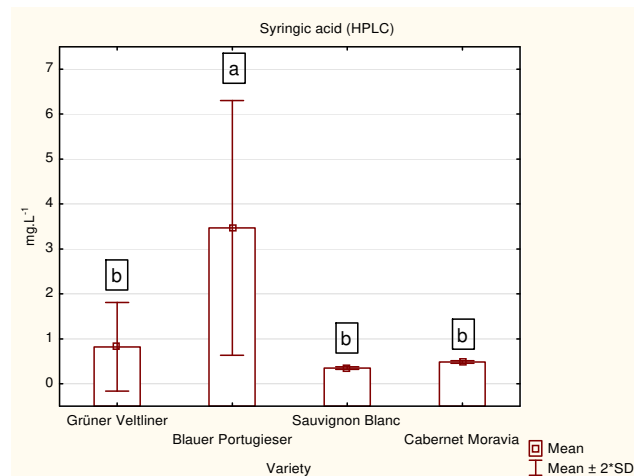
The wine industry is one of the most important agro-economic activities in the countries of southern Europe. This results in the production of a large number of by-products, rich in proanthocyanidins, flavonols, hydroxycinnamic acid derivatives and anthocyanins. These data suggest that grape stems are a rich source of healthy phytochemicals that could be used as food for animals (Barros *et al.*, 2014).

The highest content (4.015 mg.L<sup>-1</sup>) of gallic acid was found in the Sauvignon variety (Figure 6). The lowest content of gallic acid was in the Blauer Portugieser variety (0.822 mg.L<sup>-1</sup>). Figure 7 shows concentration of syringic acid, which was evaluated to be highest in the Blauer Portugieser variety (1.346 mg.L<sup>-1</sup>). The lowest concentration of this acid was found in the Sauvignon variety (0.349 mg.L<sup>-1</sup>).



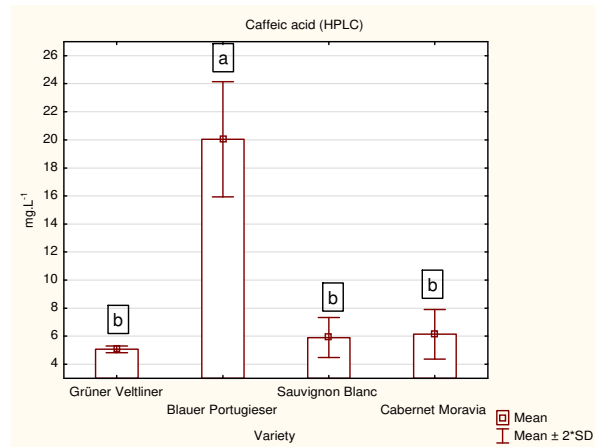
**Figure 6.** Concentration of gallic acid

Gallic acid in stems of red varieties in different studies ranged in concentration from 0.07-33.00 mg·g<sup>-1</sup>·dw, determined by HPLC-DAD (Anastasiadi *et al.*, 2012; Apostolou *et al.*, 2013; Di Lecce *et al.*, 2014). In white varieties, the concentration range was 0.01-0.03 mg·g<sup>-1</sup>·dw, determined by HPLC-DAD (Cetin *et al.*, 2011), and 1.05-22.60 µg·g<sup>-1</sup>·dw, determined by HPLC-DAD (Anastasiadi *et al.*, 2012; Apostolou *et al.*, 2013). Syringic acid in red varieties was 32.20 mg·g<sup>-1</sup>·dw, determined by HPLC-DAD and in white varieties ≤0.10 µg·g<sup>-1</sup>·dw, HPLC-DAD (Apostolou *et al.*, 2013). Results depended on extraction techniques and methods of determination.



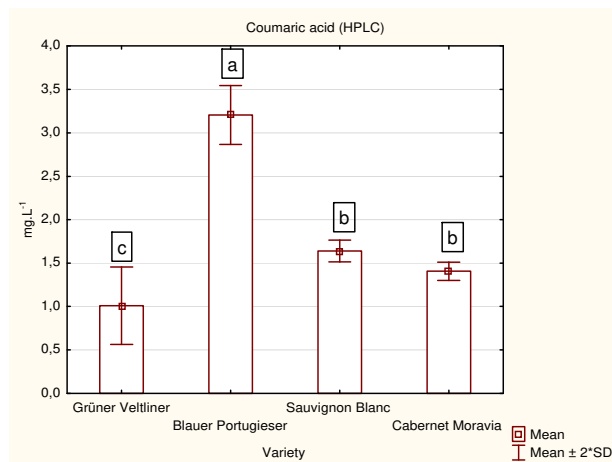
**Figure 7.** Concentration of syringic acid

As with most other acids, the concentration of caffeic acid (Figure 8) was again highest in the Blauer Portugieser variety (20 mg·L<sup>-1</sup>). The coumaric acid content of the Grüner Veltliner variety was not higher than 1 mg·L<sup>-1</sup> in all three samples tested (Figure 9).



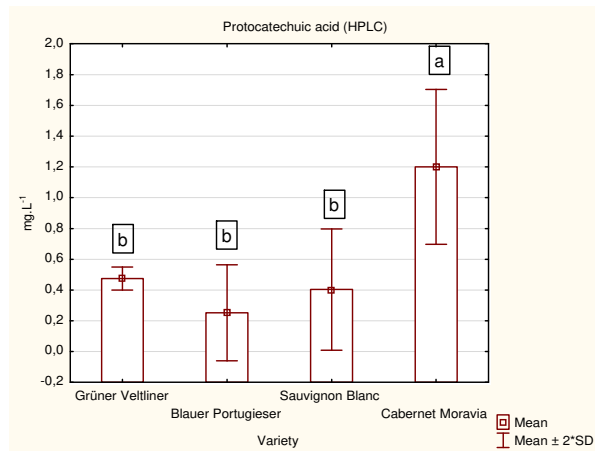
**Figure 8.** Concentration of caffeic acid

In different studies, in red varieties, caffeic acid was determined to be  $\leq 0.60 \text{ mg}\cdot\text{g}^{-1}\cdot\text{dw}$  (Apostolou *et al.*, 2013);  $0.60\text{-}1.90 \text{ }\mu\text{g}\cdot\text{g}^{-1}\cdot\text{dw}$  (Cetin *et al.*, 2011). White varieties were determined to be  $\leq 0.05 \text{ }\mu\text{g}\cdot\text{g}^{-1}\cdot\text{dw}$  (Apostolou *et al.*, 2013);  $1.00\text{-}1.50 \text{ mg}\cdot\text{g}^{-1}\cdot\text{dw}$  (Cetin *et al.*, 2011). The concentration of coumaric acid in red varieties was determined to be  $0.04\text{-}0.90 \text{ mg}\cdot\text{g}^{-1}\cdot\text{dw}$  (Apostolou *et al.*, 2013);  $0.90\text{-}2.20 \text{ }\mu\text{g}\cdot\text{g}^{-1}\cdot\text{dw}$  (Cetin *et al.*, 2011), and in white varieties in concentrations of  $0.01\text{-}0.08 \text{ }\mu\text{g}\cdot\text{g}^{-1}\cdot\text{dw}$  (Apostolou *et al.*, 2013), all determined by HPLC-DAD.



**Figure 9.** Concentration of coumaric acid

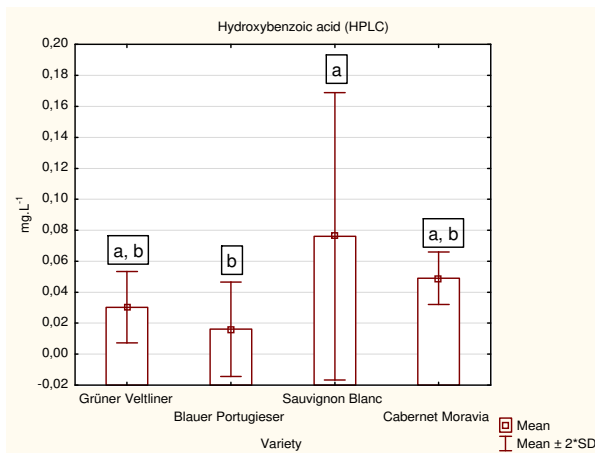
Figure 10 shows the highest concentration of protocatechuic acid was in the Cabernet Moravia variety ( $1.201 \text{ mg}\cdot\text{L}^{-1}$ ). In the Grüner Veltliner variety, the content of protocatechuic acid was  $0.426 \text{ mg}\cdot\text{dm}^{-3}$ . The lowest measured values were again in the Blue Portugal variety ( $0.253 \text{ mg}\cdot\text{L}^{-1}$ ). The content of 4-hydroxybenzoic acid was evaluated as very low in the stems of the Grüner Veltliner, Blauer Portugieser, Sauvignon and Cabernet Moravia varieties (Figure 11). The highest content of this acid was measured in the Sauvignon variety, on average  $0.076 \text{ mg}\cdot\text{L}^{-1}$ .



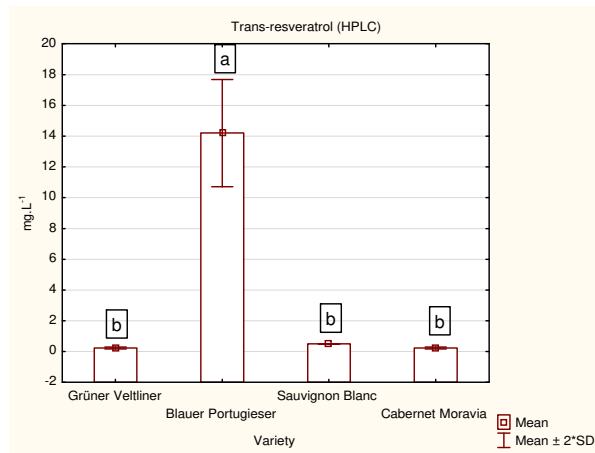
**Figure 10.** Concentration of protocatechuic acid

The higher concentration of trans-resveratrol was in Blauer Portugieser, the lowest concentrations were measured in both green varieties and also in the blue Cabernet Moravia variety (Figure 12). The lowest content of ferulic acid was measured in the Grüner Veltliner variety ( $0.005 \text{ mg.L}^{-1}$ ) (Figure 13). Ferulic acid in stems of the red variety was  $\leq 2.50 \text{ (mg.g}^{-1}\text{.dw HPLC-DAD)}$  (Apostolou *et al.*, 2013).

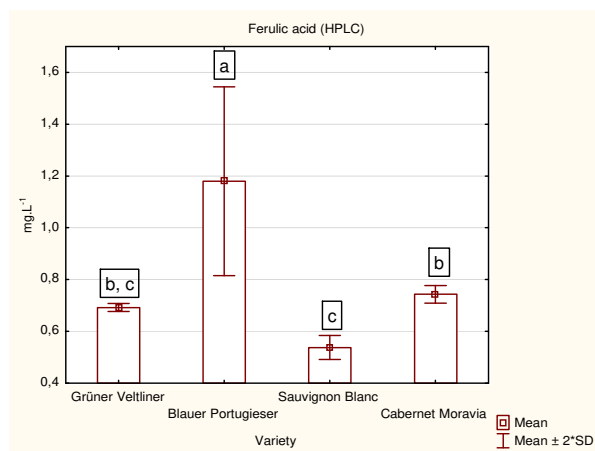
Trans-resveratrol in stems of red varieties in different studies was determined to be in the range of  $\leq 0.09$ - $124.10 \text{ mg.g}^{-1}\text{.dw}$ , and in white varieties was  $\leq 0.02 \text{ mg.g}^{-1}\text{.dw}$  (Cetin *et al.*, 2011; Anastasiadi *et al.*, 2012; Apostolou *et al.*, 2013).



**Figure 11.** Concentration of hydroxybenzoic acid

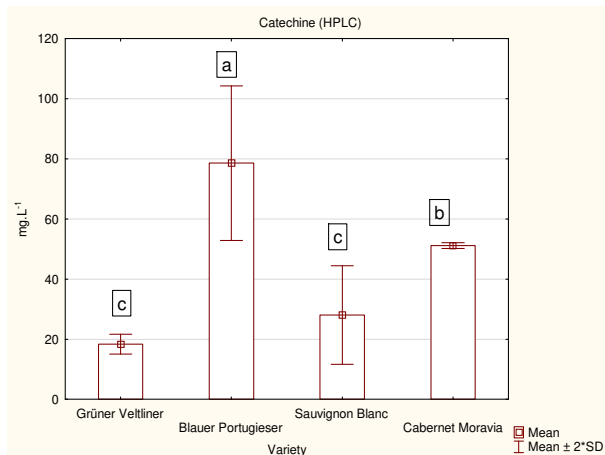


**Figure 12.** Concentration of trans-resveratrol

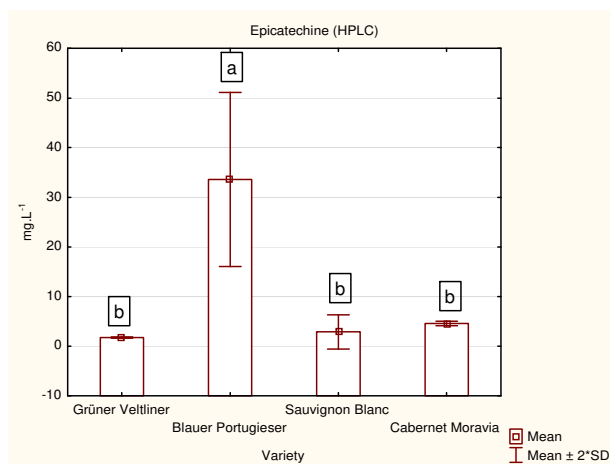


**Figure 13.** Concentration of ferulic acid

The values of catechin were evaluated as the highest by the chromatographic method in comparison with the other substances. Its highest concentration was measured in the Blauer Portugieser variety (Figure 14). The high levels of epicatechin were measured in the Blauer Portugieser variety, and were more than ten times higher than those of the other varieties, Grüner Veltliner, Sauvignon and Cabernet Moravia (Figure 15). The highest content of epicatechine was also in the variety Blue Portugal.



**Figure 14.** Concentration of catechine



**Figure 15.** Concentration of epicatechine

Catechin concentrations in stems of red varieties in different studies using HPLC\_DAD or HPLC-UV were determined to be 0.71-85.80 mg·g<sup>-1</sup>·dw; 1.24-2.58 µg·g<sup>-1</sup>·dw; and 0.12-1.27 mg·g<sup>-1</sup>·dw. White variety stem concentrations ranged from 46.50-98.30 µg·g<sup>-1</sup>·dw; 0.13-2.89 mg·g<sup>-1</sup>·dw; 3.85-18.58 µg·g<sup>-1</sup>·dw and 9.30-133.90 mg·g<sup>-1</sup>·dw (Cetin *et al.*, 2011; Anastasiadi *et al.*, 2012; Gonzáles-Centeno *et al.*, 2012; Apostolou *et al.*, 2013; Spatafora *et al.*, 2013; Sá *et al.*, 2014).

Epicatechin concentrations in stems of red varieties were determined to be ≤1.00-13.30 mg·g<sup>-1</sup>·dw, and ≤0.11 mg·g<sup>-1</sup>·dw. White variety stem concentrations ranged from ≤4.00-0.58 µg·g<sup>-1</sup>·dw; 0.04-1.13 mg·g<sup>-1</sup>·dw and 0.50-5.80 mg·g<sup>-1</sup>·dw (Cetin *et al.*, 2011; Anastasiadi *et al.*, 2012; Gonzáles-Centeno *et al.*, 2012; Apostolou *et al.*, 2013; Spatafora *et al.*, 2013; Sá *et al.*, 2014).

## Conclusions

The result of this study is a comparison of the concentrations of all-important phenolic substances within different grape varieties. Large differences can be observed not only between white and red varieties, but also within red varieties. These results can serve as a basis for further research and experiments with coniferous fermentation. This is exactly what the study according to (Pascual *et al.*, 2016) dealt with, which evaluated the influence of grape seeds and cones on the composition, color and astringency of wine. Grape stems are also a

source of yeast assimilable nitrogen. The concentration of nitrogen in amino acids ranged from 33.3 mg.L<sup>-1</sup> in white varieties to 165 mg.L<sup>-1</sup> in blue varieties. The concentration of ammonia nitrogen ranged from 35.7 mg.L<sup>-1</sup> in the blue variety Cabernet Moravia to 214 mg.L<sup>-1</sup> in Blauer Portugieser variety. These concentrations are not negligible and can partially contribute to the total amount of YAN in the case of whole-bunch fermentation.

The results of HPLC analysis showed the highest measured values of the tested substances in the Blauer Portugieser variety. Only the content of 4-hydroxybenzoic acid and gallic acid was highest in the Sauvignon variety, and the highest content of protocatechuic acid was detected in the Cabernet Moravia variety.

### Authors' Contributions

BP - determination of antioxidant activity and total flavanols, writing of article; JL - sample preparation, statistical analysis; MK - determination of antioxidants by HPLC; MB - determination of basic analytical parameters, writing of article; JS - head of experiment, supervision of the article.

All authors read and approved the final manuscript.

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### Conflict of Interests

The authors declare that there are no conflicts of interest related to this article.

### References

- Anastasiadi M, Chorianopoulos NG, Nycha GJE, Haroutounian SA (2009). Antilisterial activities of polyphenol-rich extracts of grapes and vinification byproducts. *Journal of Agricultural and Food Chemistry* 57:457-463. <https://doi.org/10.1021/jf8024979>
- Anastasiadi M, Pratsinis H, Kletsas D, Skaltsounis AL, Haroutounian SA (2012). Grape stem extracts: Polyphenolic content and assessments of their *in vitro* antioxidant properties. *LWT Food Science and Technology* 48:316-322. [10.1016/j.lwt.2012.04.006](https://doi.org/10.1016/j.lwt.2012.04.006)
- Anwar F, Kalsoom U, Sultana B, Mushtaq M, Mehmood T, Arshad HA (2013). Effect of drying method and extraction solvent on the total phenolics and antioxidant activity of cauliflower (*Brassica oleracea L.*) extracts. *International Food Research Journal* 20:653-659.
- Apostolou A, Stagos D, Galitsiou E, Spyrou A, Haroutounian S, Portesis N, ... Kouretas D (2013). Assessment of polyphenolic content, antioxidant activity, protection against ROS-induced DNA damage and anticancer activity of *Vitis vinifera* stem extracts. *Food of Chemistry and Toxicology* 61:60-68. [10.1016/j.fct.2013.01.029](https://doi.org/10.1016/j.fct.2013.01.029)
- Bang SH, Hyun YJ, Shim J, Hong SW, Kim DH (2015). Metabolism of rutin and poncirin by human intestinal microbiota and cloning of their metabolizing  $\alpha$ -L-rhamnosidase from *Bifidobacterium dentium*. *Journal of Microbiology and Biotechnology* 25:18-25. <https://doi.org/10.4014/jmb.1404.04060>
- Baron M, Kumsta M, Prusova B, Tomaskova L, Sochor J (2017). Effect of pre-fermentation maceration on the content of antioxidant compounds in grapevine juice. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca* 45(1):105-111. <https://doi.org/10.15835/nbha45110531>

- Barros A, Girones-Vilaplana A, Teixeira A, Collado J, Moreno DA, Rosa E, Dominigues-Perles R (2014). Evaluation of grape (*Vitis vinifera L.*) stems from Portuguese varieties as a resource of (poly)phenolic compounds: A comparative study. *Food Research International* 65:375-384. <https://doi.org/10.1016/j.sjbs.2020.02.013>
- Cabanis JC (2000). Ácidos orgánicos, sustancias minerales, vitaminas y lípidos in *Enología: Fundamentos científicos y tecnológicos [Organic acids, minerals, vitamins and lipids in enology: Scientific fundamentals and technological]*. France: Mundi Prensa and AMV. <http://faostat.fao.org/site/339/default.aspx>.
- Çetin ES, Altinöz D, Tarçan E, Baydar NG (2011). Chemical composition of grape canes. *Industrial Crops and Products* 34:994-998. <https://doi.org/10.1016/j.indcrop.2011.03.004>
- Chafer A, Pascual-Martí MC, Salvador A, Berna A (2005). Supercritical fluid extraction and HPLC determination of relevant polyphenolic compounds in grape skin. *Journal of Separation Science* 28:2050-2056. <https://doi.org/10.1002/jssc.200500128>
- Di Donato P, Taurisano V, Tommonaro G, Pasquale V, Jiménez JMS, de Pascual-Teresa S, ... Nicolaus B (2017). Biological properties of polyphenolic extracts from agro industry's wastes. *Waste and Biomass Valorization* 18(1):336-341. <https://doi.org/10.1080/1828051X.2018.1529544>
- Di Lecce G, Arranz S, Jáuregui O, Tresserra-Rimbau A, Quifer-Rada P, Lamuela-Raventós RM (2014). Phenolic profiling of the skin, pulp and seeds of Albariño grapes using hybrid quadrupole time-of-flight and triple-quadrupole mass spectrometry. *Food Chemistry* 145:874-882. [10.1016/j.foodchem.2013.08.115](https://doi.org/10.1016/j.foodchem.2013.08.115)
- Dineiro-García Y, Suarez Valles B, Picinelli Lobo A (2009). Phenolic and antioxidant composition of by-products from the cider industry: apple pomace. *Food Chemistry* 117:731-738. <https://doi.org/10.1016/j.foodchem.2009.04.049>
- Domínguez-Perles R, Teixeira AI, Rosa E, Barros AI (2014). Assessment of (poly)phenols in grape (*Vitis vinifera L.*) stems by using food/pharma industry compatible solvents and response surface methodology. *Food Chemistry* 164:339-346. <https://doi.org/10.1016/j.foodchem.2014.05.020>
- Garrido-Banuelos G, Buica A, Schuckel J, Zietsman AJJ, Willats WGT, Moore JP, Du Toit WJ (2019). Investigating the relationship between cell wall polysaccharide composition and the extractability of grape phenolic compounds into Shiraz wines. Part II: Extractability during fermentation into wines made from grapes of different ripeness levels. *Food Chemistry* 278:26-35. <https://doi.org/10.1016/j.foodchem.2018.10.136>
- González-Centeno MR, Jourdes M, Fermentia A, Simal S, Rosselló C, Teissedre PL (2012). Proanthocyanidin composition and antioxidant potential of the stem winemaking byproducts from 10 different grape varieties (*Vitis vinifera L.*). *Journal of Agricultural and Food Chemistry* 60:11850-11858. <https://doi.org/10.1021/jf303047k>
- Gouvinhas I, Pinto R, Santos R, Saavedra MJ, Barros AI (2020). Enhanced phytochemical composition and biological activities of grape (*Vitis vinifera L.*) stems growing in low altitude regions. *Scientia Horticulturae* 265:109-248. <https://doi.org/10.1016/j.scienta.2020.109248>.
- Kabir F, Tow WW, Hamauzu Y, Katayama S, Tanaka S, Nakamura S (2015). Antioxidant and cytoprotective activities of extracts prepared from fruit and vegetable wastes and by-products. *Food Chemistry* 167:358-362. <https://doi.org/10.1016/j.foodchem.2014.06.099>
- Karvela E, Makris DP, Kalogeropoulos N, Karathanos VT (2009). Deployment of response surface methodology to optimise recovery of grape (*Vitis vinifera*) stem polyphenols. *Talanta* 79:1311-1321. <https://doi.org/10.1016/j.profoo.2011.09.249>
- Leal C, Santos AR, Pinto R, Queiroz M, Rodrigues M, Saavedra MJ, ... Gouvinhas I (2020). Recovery of bioactive compounds from white grape (*Vitis vinifera L.*) stems as potential antimicrobial agents for human health. *Saudi Journal of Biological Sciences* 27:1009-1015. doi: <https://doi.org/10.1016/j.sjbs.2020.02.013>
- Llobera A, Canellas J (2007). Dietary fibre content and antioxidant activity of Manto Negro red grape (*Vitis vinifera*): Pomace and stem. *Food Chemistry* 101:659-666. [10.1016/j.foodchem.2006.02.025](https://doi.org/10.1016/j.foodchem.2006.02.025)
- Lu Y, Foo LY (1999). The polyphenol constituents of grape pomace. *Food Chemistry* 65:1-8. [https://doi.org/10.1016/S0308-8146\(98\)00245-3](https://doi.org/10.1016/S0308-8146(98)00245-3)
- Maier T, Schieber A, Kammerer DR, Carle R (2009). Residues of grape (*Vitis vinifera L.*) seed oil production as a valuable source of phenolic antioxidants. *Food Chemistry* 112(3):551-559.
- Makris DP, Boskou G, Andrikopoulos NK, Kefalas P (2008). Characterisation of certain major polyphenolic antioxidants in grape (*Vitis vinifera cv. Roditis*) stems by liquid chromatography-mass spectrometry. *European Food Research Technology* 226:1075-1079. <https://doi.org/10.1007/s00217-007-0633-9>

- Okolie C, Akanbi T, Mason B, Udenigwe C, Aryee A (2019). Influence of conventional and recent extraction technologies on physicochemical properties of bioactive macromolecules from natural sources: a review. *Food Research International* 116:827-839. <https://doi.org/10.1016/j.foodres.2018.09.018>
- Paixão N, Perestrelo R, Marques JC, Câmara JS (2007). Relationship between antioxidant capacity and total phenolic content of red, rosé and white wines. *Food Chemistry* 105:204-214. <https://doi.org/10.1016/j.foodchem.2007.04.017>.
- Pascual O, Gonzalez-Royo E (2016). Influence of grape seeds and stems on wine composition and astringency. *Journal of Agricultural and Food Chemistry* 64(34):6555-6566. <https://doi.org/10.1021/acs.jafc.6b01806>
- Pintač D, Majkic T, Torovic L, Orcica D, Beara I, Simin N, ... Lesjak M (2018). Solvent selection for efficient extraction of bioactive compounds from grape Pomace. *Industrial Crops and Products* 111:379-390. <https://doi.org/10.1016/j.indcrop.2017.10.038>
- Portu J, López R, Santamaría P, Garde-Cerdán T (2018). Methyl jasmonate treatment to increase grape and wine phenolic content in Tempranillo and Graciano varieties during two growing seasons. *Scientia Horticulturae (Amsterdam)* 240:378-386. <https://doi.org/10.1016/j.scienta.2018.06.019>.
- Pulido R, Bravo L, Saura-Calixto F (2000). Antioxidant activity of dietary polyphenols as determined by a modified ferric reducing/antioxidant power assay. *Journal of Agricultural and Food Chemistry* 48:3396-3402. <https://doi.org/10.1021/jf9913458>
- Queiroz M, Oppolzer D, Gouvinhas I, Silva AM, Barros AIRNA, Raúl Domínguez-Perles R (2017). New grape stems' isolated phenolic compounds modulate reactive oxygen species, glutathione, and lipid peroxidation *in vitro*: Combined formulations with vitamins C and E. *Fitoterapia* 120:146-157. <http://dx.doi.org/10.1016/j.fitote.2017.06.010>
- Ruiz-Moreno MJ, Raposo R (2015). Efficacy of olive oil mill extract in replacing sulfur dioxide in wine model. *Lwt-Food Science and Technology* 61(1):117-123.
- Sahpazidou D, Geromichalos GD, Stagos D, Apostolou A, Haroutounian SA, Tsatsakis AM, ... Kouretas D (2014). Anticarcinogenic activity of polyphenolic extracts from grape stems against breast, colon, renal and thyroid cancer cells. *Toxicology Letters* 230:218-224. <https://doi.org/10.1016/j.toxlet.2014.01.042>.
- Sanmartin C, Taglieri I, Venturi F, Ferroni G, Flamini G, Macaluso M, ... Zinnai A (2019). Co-fermentation of intact grape clusters and stalk: a natural and economical strategy to modulate nutraceutical and sensory features of Syrah variety. *Agrochimica* 63(2):197-207. [10.12871/00021857201927](https://doi.org/10.12871/00021857201927)
- Sá M, Justino V, Spranger MI, Zhao YQ, Han L, Suna BS (2014). Extraction yields and anti-oxidant activity of proanthocyanidins from different parts of grape pomace: Effect of mechanical treatments. *Phytochemical Analysis* 25:134-140.
- Sepúlveda L, Romani A, Aguilar CN, Teixeira J (2018). Valorization of pineapple waste for the extraction of bioactive compounds and glycosides using autohydrolysis. *Innovative Food Science and Emerging Technologies* 47:38-45. <https://doi.org/10.1016/j.ifset.2018.01.012>
- Silva V, Igrejas G, Falco V, Santos TP, Torres C, Oliveira AMP, ... Poeta P (2018). Chemical composition, antioxidant and antimicrobial activity of phenolic compounds extracted from wine industry by-products. *Food Control* 92:516-522. <https://doi.org/10.1016/j.foodcont.2018.05.031>
- Sochor J, Zitka O, Skutkova H, Pavlik D, Babula P, Krska B, ... Kizek R (2010a). Content of phenolic compounds and antioxidant capacity in fruits of apricot genotypes. *Molecules* 15(9):6285-6305. <https://doi.org/10.3390/molecules15096285>
- Sochor J, Ryvolova M, Krystofova O, Salas P, Hubalek J, Adam V, ... Kizek R (2010b). Fully automated spectrometric protocols for determination of antioxidant activity: advantages and disadvantages. *Molecules* 15:8618-8640. <https://doi.org/10.3390/molecules15128618>
- Spatafora C, Barbagallo E, Amico V, Tringali C (2013). Grape stems from Sicilian *Vitis vinifera* cultivars as a source of polyphenol-enriched fractions with enhanced antioxidant activity. *Food Science and Technology* 54:542-548. <https://doi.org/10.1016/j.lwt.2013.06.007>
- Teixeira A, Baenas N, Dominguez-Perles R, Barros A, Rosa E, Moreno DA, Garcia-Viguera C (2014). Natural bioactive compounds from winery byproducts as health promoters: A review. *International Journal of Molecular Science* 15:15638-15678. <https://doi.org/10.3390/ijms150915638>.
- Wenzel J, Samaniego CS (2015). Superheated liquid and supercritical denatured ethanol extraction of antioxidants from Crimson red grape stems. *Food Science and Nutrition* 3(6):569-576. <https://doi.org/10.1002/fsn3.246>

- Yacco RS, Watrelot AA, Kennedy JA (2016). Red wine tannin structure- activity relationships during fermentation and maceration. *Journal of Agricultural and Food Chemistry* 64:860-869. <https://doi.org/10.1021/acs.jafc.5b05058>
- Yang J, Martinson TE, Hai R (2009). Phytochemical profiles and antioxidant activities of wine grapes. *Food Chemistry* 116(1):332-339. <https://doi.org/10.1016/j.foodchem.2009.02.021>



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