# Microbial phytase supplementation for improving availability of plant phosphorus in the diet of the growing pigs

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> Abstract. The effect of a phytase supplement produced by Aspergillus niger on phytinphosphorus availability for pigs on phytase deficient maize-soybean meal diets was measured in two digestibility and balance experiments involving twelve growing pigs. Apparent digestibility of P in diets without inorganic-P supplementation or with low addition (0.18 of total P) was significantly lower than in the control diets (0.16 and 0.23 vs. 0.42; P < 0.01), respectively. Phytase supplementation improved P digestibility (P < 0.01) such that plant-P digestibility (0.40 and 0.28) rose to the same level as the control diet. P retention was also increased significantly by phytase supplementation. Calcium digestibility and retention were higher in phytase treated diets. Due to its effect on phytate-P availability, phytase treatment of feedstuffs allows a greater proportion on the pig's P requirement to be met by P of plant origin.

Index words: phytase, phosphorus, minerals, digestibility, availability, pig.

#### Introduction

In cereals and vegetable protein sources half to three quarters of the phosphorus (P) occurs organically bound with phytic acid in form of myoinositolphosphate, which is the major storage form of P in plant tissue. The availability of plant-phytate-P is lower for pigs than that of non-phytate-P in plants or inorganic P of mineral sources and is variable among different plant feeds as reviewed by CROM-WELL (1980, 1989) and JONGBLOED (1987). In order for the pigs to utilize the phosphorus from phytate it must be hydrolysed by phytate-degrading enzymes to yield phosphates. Moreover, phytic acid binds strongly to other essential dietary minerals, such as calcium, zinc, magnesium, iron and copper, reducing their availability in the digestive tract of monogastrics (ZHU et al. 1990).

Although, phytase activity occurs in microbes and microbial phytases can be produced biotechnically, there are no reports of improved plant P utilization by pigs receiving phytase supplements (SHURSON et al. 1983 JONGBLOED 1987). However, in broilers phytase supplementation of diets (KIISKINEN and PIIRONEN 1990) and in chicks the addition of phytase producing organisms to the diet has resulted in a marked improvement in the utilization of phytate-P (NELSON et al. 1971). In pigs, similar attempts to supplement the diets with a dried yeast product have been unsuccessful (CROMWELL et al. 1978).

The objective of this study was to evaluate the possibility of improving the usage of phosphorus from plant ingredients by a phytase supplement produced by *Aspergillus niger* with a new technology. A maize-soybean meal pig ration containing low naturally occurring phytase activity was chosen as the test diet. Attention was also paid to the reduction of the phosphorus content of pig excrement in order to avoid the accumulation of phosphates in soils.

#### Materials and methods

Two experiments were conducted with six castrated growing pigs, in which the effect of

added microbial phytase on the availability of plant phosphorus (P) was measured using balance assay.

The experimental diets were composed of maize (880 g/kg) and soybean meal (120 g/kg) and the daily allowances were 2500-2700 g. The control diets in both exps. were supplemented with dicalcium phosphate to give 6.5 g/kg total phosphorus in the ration. In exp. 1, diets 2 and 3 were supplemented with 3.3 g/kg of dicalcium phosphate to provide 0.50 of total P of the control diet mixture. In exp. 2 no inorganic P-supplementation was added to diets 2 and 3 (Table 2). In control diets, maize and soybean meal contributed 0.44 of the total supply of P and dicalcium phosphate provided 0.56. Calcium was added in all of the rations to provide diets with a content of 0.80 % total calcium. The trace elements and vitamins were supplemented according to Finnish recommendations (SALO et al. 1982) in all treatments.

	Dry matter	Ash	Crude protein	Ether extract	Crude carbo- hydrates	Phos- phorus	Phytic acid
Soybean meal							
Exp. 1	868	65	502	36	398	7.87	4.33
Exp. 2	895	66	497	36	401	7.29	4.38
Maize							
Exp. 1	860	13	98	50	839	2.80	2.82
Exp. 2	878	15	95	58	832	2.78	3.04
Dicalcium							
phosphate							
Exp. 1	802	782				185.44	
Exp. 2	983	853				166.27	

Table 1. Chemical composition of the experimental feed ingredients (g/kg DM).

Table 2. Feed intake of pigs. (Exp. 1, Exp. 2), (g DM/d).

Experiment		11				
Treatment	1	2	3	1	2	3
Maize	1950	1954	1955	1994	1921	1998
Soybean meal	269	269	269	277	267	278
Calcium carbonate	9	41	41	13	47	49
Dicalcium phosphate	52	9	9	57	_	
Sodium chloride	5	5	5	5	5	5
Trace element prem.	15	15	15	16	16	16
Phytase supplement	_	_	11		_	2.3

The phytase product Finase-F (produced by Alko Ltd., Rajamäki, Finland) was used in both experiments. The level of supplementation in diet 3 in Exp. 1 was 100 000 Pu/kg feed (1 Pu is the amount of enzyme that liberates 1 nmol of inorganic phosphorus from sodiumphytate in one minute,  $37^{\circ}$ C, pH 5.0) and 500 000 Pu/kg in Exp. 2, respectively. The enzyme was added to the ration in liquid form just before feeding. Finase-F preparation is produced by *Aspergillus niger* var. and the phytase is acid and heat resistant over broad pH and temperature ranges (pH 2.0—6.0 and up to  $60^{\circ}$ C).

The experiment had a  $3 \times 3$  replicated Latin square designs and the pigs had an average liveweight of 78 and 98 kg in exp. 1 and 2, respectively. Each period was comprised of 6 days of adjustment and 6 days of faeces and urine total collection. The chemical analyses of feeds and faeces were performed according to the official procedures. Phytic acid contents were analysed using the method of CA-MIRE and CLYDESDALE (1982). Phosphorus was determined after dry ashing by the method of TAYSSKY and SHORR (1953) and minerals were measured on the diet ingredients, faeces and urine with a Varian Techtron AA1000 atomic-absorption spectrophotometer.

#### **Results and discussion**

Average daily dry matter intake of the pigs was 79 g in Exp. 1 and 75 g in Exp. 2 per kg W<sup>0.75</sup>. Daily phosphorus intakes of pigs receiving the control diet were 17.2 g. Pigs on diets 2 and 3 in Exp. 1 consumed 9.3 g P/day, and in Exp. 2 consumed 7.5 g/day (Table 2). Dicalcium phosphate contributed 0.56 of the total P supply in control diets and 0.18 of total P in diets 2 and 3 of Exp. 1. No supplemental phosphorus was added in Exp. 2 on diets 2 and 3; all of the P supply was of plant origin.

Organic matter digestibility was not different between the treatments in either experiment (P>0.05, Table 3). Finase-F also contains some proteinase, amylase and pectinase activity, however, digestibility was not affected in this trial. The ash digestibility was improved significantly by phytase supplementation (P < 0.01) in Exp. 2. and showed a similar tendency in Exp. 1. Nitrogen digestibility was significantly higher in Exp. 2. in pigs receiving diets without an inorganic phosphorus supplement (P < 0.05). However, there was no effect on N balance (Table 4). A number of other investigations have pointed out that there is often strong binding between phytic acid and protein (ZHU 1990) but in this study

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	Org. matter	Crude prot.	Ether extr.	Crude carbo- hydr.	Ash
Experiment 1.					
Diet 1	0.911	0.866	0.734	0.929	0.486
Diet 2	0.905	0.856	0.721	0.924	0.525
Diet 3	0.907	0.859	0.726	0.926	0.548
SEM	0.0036	0.0046	0.0071	0.0037	0.0239
Statist. signif.	NS	NS	NS	NS	NS
Experiment 2.					
Diet 1	0.903	$0.851^{a}$	0.809	0.916	0.524a
Diet 2	0.919	$0.884^{b}$	0.833	0.931	0.560a
Diet 3	0.914	$0.873^{ab}$	0.817	0.928	0.642b
SEM	0.0043	0.0065	0.0082	0.0038	0.017
Statist. signif.	NS	*	NS	NS	

Table 3. Digestibilities of the nutrients in pigs on the experimental diets.

<sup>a-b</sup>: \* P < 0.05, \*\* P < 0.01

Table 4. Nitrogen balance and protein utilization.

Treatment	1	2	3	SEM	Statist. signif.
N intake, g/d	52.31ª	52.43ª	52.83 <sup>b</sup>	0.110	*
N excretion in faeces	7.00	7.53	7.47	0.236	NS
N absorption, g/d	45.31	44.90	45.35	0.312	NS
N digestibility	0.866	0.856	0.859	0.0046	NS
N excretion in urine	22.32	23.73	23.28	0.474	NS
N retained, g/d	22.99	21.17	22.07	0.461	NS
of intake	0.440 <sup>a</sup>	0.404 <sup>b</sup>	0.419 <sup>b</sup>	0.0084	*
of absorption	0.508	0.472	0.487	0.0097	NS
g/kg W <sup>0.75</sup>	0.790	0.731	0.770	0.015	NS
Biological value	0.601	0.567	0.581	0.0091	NS
Exp. 2.					
Treatment	1	2	3	SEM	Statist. signif.
N intake, g/d	52.29	51.62	52.39	0.433	NS
N excretion in faeces	7.76ª	5.98 <sup>b</sup>	6.66 <sup>ab</sup>	0.350	*
N absorption, g/d	44.53	45.63	45.73	0.323	NS
N digestibility	0.851ª	0.884 <sup>b</sup>	0.873ab	0.0065	*
N excretion in urine	22.13	23.65	22.96	0.774	NS
N retained, g/d	22.40	21.98	22.77	0.673	NS
of intake	0.428	0.428	0.435	0.0161	NS
of absorption	0.504	0.483	0.498	0.0151	NS
g/kg W <sup>0.75</sup>	0.72	0.71	0.73	0.023	NS
Biological value	0.603	0.581	0.596	0.0140	NS

a-b: \* P < 0.05.

phytase treatment did not result any improvement in protein digestibility.

The apparent digestibility of P on the control diet averaged 0.42 in both trials and was significantly higher (P < 0.05, 0.01) than that of the diets without inorganic phosphorus supplementation (Exp. 2) or with low (0.18 contribution of total P) phosphate addition in Exp. 1 (Table 5). Phytase supplementation of the diet resulted a significant improvement in phosphorus digestibility (P < 0.01) compared with that of the unsupplemented diet. Phytase improved plant-P digestibility such that it rose to the same level as the inorganic-P supplemented diet (Exp. 2). Also in Exp. 1 phytase enhanced apparent digestibility of P by five percentage units (P > 0.05). The apparent digestibility assay of the P in the total diet, however, poses some difficulties in the interpretation of the results, particularly when P supplies are different as in present study.

Endogenous P secretions also complicate the measurement (JONGBLOED 1987), who reported the value of 5 mg/kg metabolic weight excreted in faeces. An approximation of these daily P-losses in pig is 1.5-2.5 g/100 kg body weight (Arc 1981).

Similar low apparent P digestibilities as in present maize-soybean meal diets have been reported in cereals or cereal-based diets containing no supplemental P (PIERCE et al. 1977, CALVERT et al. 1978, JONGBLOED 1987, OKSBJERG 1988). Phosphorus from phytate cannot be absorbed in its original form by pigs and so it must first be released by hydrolysis by phytate-degrading enzymes to yield inositol and phosphoric acid. Almost all vegetable feed ingredients poses phytase, although with different levels of activity (NELSON 1969). Maize has been reported to have very low contents of natural phytase (CALVERT et al. 1978, POINTILLART et al. 1984, 1988, OKSBJERK Table 5. Phosphorus excretion, apparent absorption and retention in pigs.

Treatment	1	2	3	SEM	Statist. signif.
P intake, g/d	17.16ª	9.30 <sup>b</sup>	9.34ª	0.107	
P excretion in faeces	9.69 <sup>a</sup>	7.12 <sup>b</sup>	6.71 <sup>b</sup>	0.197	***
P absorption, g/d	7.47ª	2.18 <sup>b</sup>	2.63 <sup>b</sup>	0.207	***
P digestibility	0.43ª	0.235 <sup>b</sup>	0.284 <sup>ab</sup>	0.0216	***
P excretion in urine	5.59ª	0.38 <sup>b</sup>	0.20 <sup>b</sup>	0.317	**
P retained, g/d	2.23	1.79	2.43	0.380	NS
of intake	0.128	0.193	0.261	0.0312	NS
of absorption	0.312c	0.828ad	0.927bd	0.0315	**
g/kg W <sup>0.75</sup>	0.07ª	0.06 <sup>a</sup>	0.09 <sup>a</sup>	0.063	NS
Exp. 2.					
Treatment	1	2	3	SEM	Statist. signif.
P intake, g/d	17.19ª	7.54b	7.65 <sup>b</sup>	0.103	***
P excretion in faeces	10.07 <sup>a</sup>	6.33 <sup>b</sup>	4.59°	0.293	**
P absorption, g/d	7.11ª	1.21 <sup>b</sup>	3.06 <sup>c</sup>	0.276	***
P digestibility	0.414 <sup>a</sup>	0.162 <sup>b</sup>	0.400 <sup>a</sup>	0.0271	***
P excretion in urine	3.37ª	0.16 <sup>b</sup>	0.06 <sup>b</sup>	0.094	***
P retained, g/d	3.72ª	1.04 <sup>b</sup>	3.00 <sup>a</sup>	0.212	***
of intake	0.217ª	0.139 <sup>a</sup>	0.392 <sup>b</sup>	0.0206	***
of absorption	0.523ª	0.887 <sup>b</sup>	0.979 <sup>b</sup>	0.0283	***
g/kg W <sup>0.75</sup>	0.12 <sup>a</sup>	0.03 <sup>b</sup>	0.10 <sup>a</sup>	0.007	***

Exp. 1.

<sup>a-c</sup>: \* P<0.05, \*\* P<0.001, \*\*\* P<0.001

1988), which is why this diet was chosen to test microbial phytase supplementation even though maize is not used in pig feeding in Finland. In contrast to this the seed coats of wheat and barley have relatively high phytase activity. The optimum pH for plant phytase is about 4.0—6.0 as reviewed by IRWING (1980) therefore plant phytase probably does not survive the acid conditions of the stomach.

P digestibility in maize-based diets has been improved by treatments other than microbial phytase supplementation. FRAPE et al. (1979) and BAGHERI and GUEGUEN (1985) have shown that absorption coefficients of phosphorus in a maize SBM diet without added P were increased from 0.36 to 0.48 by introduction of 200 g/kg wheat bran into the diet. Similarly, FOURDIN et al. (1988) reported that rye bran supplementation (200 g/kg) improved P retention 40—50 % in pigs on low P maize diets. In high-moisture maize the P is 3 to 4 times more available than in dried grain (Boyd et al. 1983). The improvements with those treatments have been similar or better than that of the present study of microbial phytase supplementation.

Besides being present in the feedstuffs, phytases are produced by the microbes in the intestine and are also secreted by the intestinal tract of the pig. Intestinal alkaline phosphatase can hydrolyse some phytate (POINTIL-LARD et al. 1984). However, it has been concluded that intestinal phytase does not seem to be of great significance for the hydrolysis of phytate (POINTILLART et al. 1984). In the large intestine with its rich microbial flora phytate can be hydrolysed by bacterial phytases. However, the pig's absorption of phosphorus in the large intestine is very limited and no P or only a limited amount of P was absorbed beyond an ileal fistula (JORGENSEN et al. 1985).

The retention of phosphorus was 0.13-0.22 of P intake on the control diets and 0.14–0.19 on diets without inorganic P supplementation. This low availability is in accordance with existing literature. Microbial phytase supplementation improved retention of phosphorus to the level of 0.26-0.39 and the difference was significant (P < 0.01) compared with the untreated non P-supplemented diet. Retention of the absorbed P was very high (0.93–0.98) in diets supplemented with phytase and was 0.10 higher relative to the untreated diets (P<0.05 in the first Exp.). Some low-molecular weight myoinositol phosphates were probably absorbed but could not be utilized by the pig.

The retention of P in Exp. 1 was considerable lower compared to Exp. 2, but the reason for this remains uncertain. Average daily gain of pigs in Exp. 1 was 729 g, while it was 854 g in Exp. 2. The pigs on low-P phytase unsupplemented diet had the lowest gains as well as the lowest P-retention. The effect of phytase supplementation was different between the two experiments due to the application level, which was five-fold higher in the second trial. JONGBLOED (1987) found that in slaughter pigs on average 4.9 g P/kg live weight was retained. The phosphorus availability in most inorganic phosphate supplements approaches 0.7 to 1.0 (CROMWELL 1989, JONGBLOED 1987, DEN HARTOG et al. 1988).

Calcium digestibility was improved by the phytase supplementation and retained Ca was significantly (P < 0.01) higher compared to the untreated diet (Table 6). When the intake of Ca is adequate but that of P inadequate, the retention of Ca falls (JONGBLOED 1987). In the present study the calculated Ca/P ratios were 0.93 and 2.07 for control and experimental groups, respectively, while the optimum Ca/P ratio is from 1.2 to 1.3:1 (ARC 1981). Digestibility of magnesium also tended to be increased by the phytase treatment but retention was not affected. Iron supply was higher in the control diet due to the high content of iron in dicalciumphosphate. Fe digestibility was

not different between diets but retention in the control diet was significantly higher. Copper and zinc digestibility and retention were higher in diets without phosphate addition but phytase supplementation did not affect any difference in utilization. Phytic acid readily forms complexes with several essential minerals such as calcium, iron, zinc and manganese and also protein impairing their utilization by the animal (NELSON et al. 1971).

There are no previous reports of successful addition of microbial phytase to pig diets. However, many of the fungi, bacteria and yeasts are good sources of phytase. Attempts to improve phytate utilization in pigs by including a dried yeast product in the diet have been unsuccessful (CROMWELL et al. 1978). In contrast, several studies with chicks have demonstrated that the addition of phytaseproducing organisms to the diet can result in a marked improvement in the utilization of phytate phosphorus (NELSON et al. 1971, CROMWELL 1980). SHURSON et al. (1983) was not able to improve phytate utilization in pigs by including a yeast phytase in the diet. It is possible that the additions of phytase have been insufficient or that the phytase used has not been resistant to the low pH in the stomach of the pig. In this study 100 000 Pu/kg was not sufficient because increasing phytase activity gave a greater response in the Exp. 2. The Finase-F preparation is thermotolerant and acid-resistant to a pH value of 2. Successful treatments in poultry have been also reported. NELSON et al. (1971) added phytase produced by a culture of Aspergillus ficium to the diet of chickens and found an increase in bone ash content compared to control. KIISKINEN and PIIRONEN (1990) showed improved plant-P digestibility in broilers after phytase supplementation.

In conclusion, the results of present study demonstrate that addition of a microbial phytate degrading enzyme in maize-soybean meal diet enhances the absorption of vegetable phosphorus and other essential minerals. More of the pigs' requirement for phosphorus could be met by P in maize and SBM if Table 6. Mineral excretion, apparent absorption and retention in pigs.

T	_	_	-	
E	X	D	2	

Treatments	1	2	3	SEM	Statis
		1			signi
Calcium					
Intake, g/d	15.91	15.60	15.84	0.131	NS
Excret. in faeces, g/d	9.72ª	8.62 <sup>ab</sup>	7.64 <sup>b</sup>	0.419	•
Absorption	0.388 <sup>a</sup>	0.450 <sup>ab</sup>	0.517 <sup>b</sup>	0.0248	*
Excretion in urine, g/d	0.52ª	3.65 <sup>b</sup>	1.87°	0.186	***
Retention, g/d	5.67ª	3.33 <sup>b</sup>	6.33ª	0.426	**
of intake	0.356ª	0.216 <sup>b</sup>	0.400 <sup>a</sup>	0.0287	**
of absorption	0.922ª	0.466 <sup>b</sup>	0.775°	0.0328	***
Magnesium					
Intake, g/d	3.48	3.47	3.52	0.029	NS
Excret. in faeces, g/d	2.53ª	2.17 <sup>ab</sup>	2.07 <sup>b</sup>	0.089	*
Absorption	0.273ª	0.375 <sup>b</sup>	0.412 <sup>b</sup>	0.0243	**
Excretion in urine, g/d	0.42 <sup>a</sup>	0.74 <sup>b</sup>	0.77 <sup>b</sup>	0.032	***
Retention, g/d	0.54	0.56	0.68	0.096	NS
of intake	0.153	0.163	0.193	0.0282	NS
of absorption	0.544	0.421	0.463	0.0481	NS
ron					
Intake, mg/d	561ª	338 <sup>b</sup>	344 <sup>b</sup>	3.4	***
Excret. in faeces, mg/d	408ª	279ь	280 <sup>b</sup>	11.5	***
Absorption	0.272	0.177	0.184	0.0279	NS
Excretion in urine, mg/d	3.10 <sup>a</sup>	1.42 <sup>b</sup>	1.62 <sup>b</sup>	0.301	**
Retention, mg/d	150 <sup>a</sup>	58 <sup>b</sup>	62 <sup>b</sup>	11.2	***
of intake	0.266	0.173	0.179	0.0276	NS
of absorption	0.977	0.972	0.974	0.0038	NS
Copper					
Intake, mg/d	184	182	184	1.5	NS
Excret. in faeces, mg/d	178ª	156 <sup>b</sup>	157ª	5.7	*
Absorption	0.031ª	0.143 <sup>ab</sup>	0.145 <sup>b</sup>	0.0294	* .
Excretion in urine, mg/d	1.7	2.1	1.3	0.31	NS
Retention, mg/d	4.3ª	23.4ab	25.6 <sup>b</sup>	5.15	*
of intake	0.021ª	0.136 <sup>ab</sup>	0.138 <sup>b</sup>	0.0291	*
of absorption	0.816	0.903	0.943	0.0183	NS
Zinc					-
Intake, mg/d	189	187	190	1.6	NS
Excret. in faeces, mg/d	184ª	159ь	155ª	6.0	*
Absorption	0.019 <sup>a</sup>	0.150 <sup>b</sup>	0.181 <sup>b</sup>	0.0314	*
Excretion in urine, mg/d	10.6	13.3	13.3	2.38	NS
Retention, mg/d	$-6.4^{a}$	14.4 <sup>ab</sup>	21.2 <sup>b</sup>	5.66	*
of intake	$-0.038^{a}$	0.078 <sup>ab</sup>	0.111 <sup>b</sup>	0.0314	*
of absorption	1.467ª	0.416 <sup>b</sup>	0.531 <sup>b</sup>	0.2260	*

<sup>a-c</sup>: \* P<0.05, \*\* P<0.01, \*\*\* P<0.001.

phosphorus could be converted to an available form. Phosphorus content in pig manure could be reduced followed the improved plant-P utilization with the present phytase treatment of the feed. Accumulation of P in cultivated fields is increased by using manures with high P content. Whether the use of enzymatic treatment to release plant phosphorus is economically feasible compared to addition of other sources of phosphorus to feed remains to be determined.

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### SELOSTUS

# Fytaasilisäyksen vaikutus kasviperäisen fosforin hyväksikäyttöön lihasian ruokinnassa

## Matti Näsi

#### Helsingin yliopisto, kotieläintieteen laitos

Kahdessa sulavuus- ja tasekokeessa tutkittiin bioteknisesti tuotetun sienifytaasin tehoa kasviperäisen fytiinifosforin hyväksikäytön parantamiseen lihasikojen ruokinnassa. Maissi-soijarouhedieetin fosforin sulavuus oli alhainen, 0.16–0.23, ja se oli merkitsevästi alempi kuin fosfaatilla täydennetyn dieetin P:n sulavuus (0.42). Fytaasilisäys paransi merkitsevästi kasviperäisen fosforin sulavuutta (0.40–0.28). Fosforin pidättyminen lisääntyi myös merkitsevästi fytaasikäsittelyn ansiosta. Samoin kalsiumin sulavuutta ja pidättymistä voitiin parantaa fytaasilisäyksen avulla. Rehujen fytiinifosforin hyväksikäyttöä voitiin parantaa huomattavasti fytaasilisäyksellä ja samalla saada aikaan sonnan fosforipitoisuuden alenemista. Fosfaattien käyttöä voitaisiin ilmeisesti vähentää sikojen ruokinnassa lisäämällä orgaanisen fosforin käyttökelpoisuutta ja siten vähentää lannan mukana poistuvan fosforin määrää.