

## Co-existence and interaction between myxomycetes and other organisms in shared niches

IRINA O. DUDKA and KATERINA O. ROMANENKO

Department of Mycology, M.G. Kholodny Institute of Botany  
National Academy of Sciences of Ukraine  
Tereshchenkivska 2, MSP 1 Kyiv, UA 01 001

Dudka I.O., Romanenko K.O. *Co existence and interaction between myxomycetes and other organisms in shared niches*. Acta Mycol. 41(1): 99 112, 2006.

The ecology of myxomycetes co existing with the *Latridiidae* (Coleoptera), bryophytes and ascomycetous, basidiomycetous and anamorphic fungi were studied in Crimea and at different locations on the left bank of Ukraine. Results from the left bank indicate that the *Latridiidae* feed on myxomycetes. Colonies of the most common 13 myxomycete species (which included *Stemonitis axifera* (Bull.) Macbr., *S. fusca* Roth, *S. splendens* Rost., *Fuligo septica* (L.) Wigg. and *Mucilago crustacea* Wigg.) were inhabited by 5 species of the *Latridiidae*. Myxomycete spores were present in guts of 19 of the 25 beetle specimens investigated. Beetles *Latridius hirtus*, *Enicmus rugosus* and *E fungicola* seem to be obligate myxomycete feeders, while *Corticarina truncatella* was clearly facultative. 13 species of myxomycetes were recorded on 9 species of moss and 3 species of liverwort developing on decaying wood or bark in the Crimean Nature Reserve. Relations between myxomycetes and bryophytes on woody substrata are spatial rather than trophic, and are possibly regulated by specific microclimatic conditions inside bryophyte thallomes. 69 species of myxomycetes were found co existing with 36 species of ascomycetes, 21 species of basidiomycetes and 9 species of anamorphic fungi in the Crimean Nature Reserve. Associations formed by myxomycetes and fungi on different woody substrata are analyzed.

**Key words:** myxomycetes, ascomycetes, basidiomycetes, anamorphic fungi, bryophytes, *Latridiidae* beetles

### INTRODUCTION

In terrestrial ecosystems, certain patterns can be found governing the spatial distribution of living organisms. These are connected with trophic, topic and another relations. Autotrophic plants are usually considered the foundation for the resulting consortia (ecological communities originating through co-development of heterogeneous groups tightly joined by common activities), since they provide a prime food source, and a substratum on which other organisms can grow. Myxomycetes (also known as slime moulds) often form a part of those consortia, and may also make associations with various other groups of organisms (Dudka et al. 1976). Research on

the co-existence of slime moulds with beetles (Coleoptera), bryophytes, ascomycetous, basidiomycetous and anamorphic fungi is briefly reviewed below.

### BRIEF HISTORICAL REVIEW

Myxomycetes are frequently associated with insects, especially beetles (Coleoptera). Stephenson et al. (1994) reviewed slime mould / beetle associations recorded since the end of the nineteenth century, and listed the beetle families most frequently forming these associations as follows: *Rhizodidae*, *Leiodidae*, *Staphylinidae*, *Clambidae*, *Eucinetidae*, *Sphindidae*, *Cerylonidae*, *Latridiidae*. The numerous and repeated records of these beetles on the same species of myxomycetes suggested an obligate connexion. Since examination of beetle gut contents usually reveals abundant myxomycete spores, it is clear the beetles feed on them. In fact, some beetle species use not only spores but also plasmodia of slime moulds as a source of nutrition (Lawrence, Newton 1980; Wheeler 1980). The term myxomycetophagy has been proposed to describe the phenomenon of beetles deriving nutrition from slime moulds (Newton 1984; Newton, Stephenson 1990).

Despite their rather diverse taxonomic range, slime moulds used by beetles as food share a number of morphological, developmental and ecological characters as follows: 1) abundant spores in large simple sporophores or large aethalia which comprise a group of sporangia covered with a common cortex and pillow-like; 2) a long period of sporophore formation during the growing season; 3) long-lasting sporophores; 4) abundance; 5) development on woody substrata; 6) spores with spiny, warty or net-like walls, 4 (5-9)-15 µm diam. (Blackwell 1984). *Arcyria incarnata* (Pers.) Pers., *Stemonitis axifera* (Bull.) Macbr., *S. fusca* Roth, *S. splendens* Rost., *Lycogala epidendrum* (L.) Fr., *Stemonaria longa* (Peck) Nann.-Brem., *Fuligo septica* (L.) Wigg., *Tubifera ferruginosa* (Batsch) Gmel. and others largely match those criteria. Stephenson et al. (1994) have shown that myxomycetes of the genus *Stemonitis* Roth are used as food by 10 beetle genera, *Fuligo* Hall. by 9, and *Arcyria* Wigg. and *Tubifera* Gmel. by 7 each.

The feeding preferences of some beetles for myxomycetes explains their constant association with slime moulds. There is, however, evidence that these beetles also play some part in disseminating myxomycete spores. Observation of *Fuligo septica* spores from *Sphindidae* beetle guts has shown that some spores pass through insect intestines without damage and can subsequently germinate, while transfer of spores of *Physarum straminipes* Lister and *F. septica* from the exoskeleton surface of *Latridiidae* beetles to agar medium has also been observed, particularly when there are large numbers of such spores (Blackwell, Laman, Gilbertson 1982).

Representatives of the family *Latridiidae* (Coleoptera) are small beetles, no more than 3 mm long, occurring under bark of fallen trunks, in decaying wood and in forest litter. Worldwide about 750 species are known, 55 in Ukraine. Typically they are associated with fungi, in particular anamorphic fungi, ascomycetes and basidiomycetes, predominantly with basidiomata of wood-destroying fungi, where they feed on the spores (Howard, Currie 1932; Newton, Stephenson 1990; Stephenson et al. 1994). Information about associations between the *Latridiidae* and myxomycetes has been summarized (Dudka, Trikhleb, Romanenko 2002).

The co-existence of myxomycetes and bryophytes has been studied much less than that of slide moulds and beetles. Myxomycete sporophores at the thallome

surface of mosses and liverworts are rather widespread in nature (Stephenson, Stempen 1994; Härkönen et al. 2002; Stojanowska, Panek 2004). In most cases, however, associations of slime moulds and bryophytes arise accidentally since both components tend to occur on the same substratum – decaying and decomposing wood of deciduous and/or coniferous trees (Stephenson, Studlar 1985). It has also been surmised that myxomycetes develop more intensively and occur more frequently on fallen decaying logs overgrown with bryophytes because of their high humidity (Stojanowska, Panek 2004). Only a few myxomycetes, in particular *Barbeyella minutissima* Meylan, *Colloderma oculatum* (Lipp.) G. Lister and *Lepidoderma tigrinum* (Schr.) Rost. seem to be true bryophilous species. The first of these species is adapted to rich communities of liverworts, especially *Novellia curvifolia* and species of the genus *Cephalozia* which sometimes completely cover the surface of fallen decaying coniferous logs by (Schnittler, Novozhilov 1998; Schnittler, Stephenson, Novozhilov 2000; Novozhilov 2005).

In forest ecosystems many organisms are involved in decomposition of woody substrata. Apart from slime moulds, wood-inhabiting fungi are also active at various stages of succession. Some fungal species are known to develop and sporulate on the surface of myxomycete fruitbodies, using them as a feeding substratum (Rogerson, Stephenson 1993), but that is a case of fungal parasitism on slime moulds and an example of trophic relations which are probably negative for the myxomycete. Other interrelations between myxomycetes and these fungi so far remain almost totally unstudied, although side-by-side co-existence of myxomycetes and fungi on the same woody substratum is rather commonly observed in nature. Such relations appear to be indifferent, but that is only an assumption: exact information is absent not only about the nature of those relations but about also the slime mould and fungal species compositions on which they are based. Co-existence of myxomycetes and true fungi has therefore been studied least of all (Rogerson, Stephenson 1993).

## OBJECTIVES

It is clear that the characteristics of the co-existence and of interrelations between myxomycetes and other organisms is still poorly understood. The aim of the present work has therefore been to contribute to that research by identifying myxomycetes associated with *Latrindiidae* family beetles, liverworts, mosses and wood-inhabiting anamorphic, ascomycetous and basidiomycetous fungi collected in different parts of southern Ukraine. This study has also been designed to begin analysis of relations between component members in the associations.

## MATERIAL AND METHODS

Fieldwork was carried out from April to September 2000 on the left bank of Ukraine within three geobotanical regions (Donetsk, Leftbank and Starobilsk grass-meadow steppes), and in the Crimean peninsula within the Crimean Nature Reserve (Montane Crimea geobotanical region) during the growing seasons of 2000 and 2001. Left bank myxomycete collections were predominantly from forest plantations near Donetsk, and flood-plane forests around Stanychno-Luganske settlement in the valley of the Siverski Donets river (Table 1) where oak (*Quercus robur* L.), ash (*Fraxinus*

*excelsior* L.) and aspen (*Populus tremula* L.) are dominant tree species. Beech (*Fagus sylvatica* L.) and Crimean pine (*Pinus pallasiana* D. Don) dominate other tree species in the study areas of forest communities in the Crimean Nature Reserve.

Myxomycete colonies (groups of sporophores and aethalia) were sampled in the field from fallen trunks and logs of trees in various stages of wood decay, from fallen branches of various diameters and from decaying stumps. 53 field samples of myxomycetes were collected on woody substrata the left bank study areas and 450 from the Crimean Nature Reserve. 300 samples of woody substrata from that reserve, mainly tree bark, were collected for laboratory detection of myxomycetes, using the moist chamber method (Gilbert, Martin 1933; Stephenson 1985; Nannenga-Bremekamp 1991). For myxomycete identification, the main manuals on the group were used (Nannenga-Bremekamp 1991; Novozhilov 1993; Stephenson, Stempen 1994). Slime mould nomenclature follows Ing (1999).

Of the 53 samples collected from left bank forest plantations, 25 were examined for the presence of *Latridiidae* in the myxomycete colonies. The biggest colonies were selected from the field samples, then divided into three more or less equal parts and placed in 90 mm sterile Petri dishes with filter paper at the bottom. The dark bodies of beetles emerging from the colony were easily observed against the white surface of the paper. These individuals were then identified, and their gut contents sampled. For identification of beetles the appropriate volume of Fauna Hungariae was used (Rücker 1983). Preparations of gut contents for identification of myxomycetous spores were made in Fore liquid (50 ml distilled water, 20 g chloralhydrate, 40 ml glycerine, 30 g gum-arabic).

In addition to gut content studies of *Latridiidae* specimens found on slime moulds collected in the field, living *Latridius hirtus* were kept in the laboratory on aethalia of *Fuligo intermedia*, to see if *L. hirtus* can complete its life cycle with only a single feed source present. Pillow-like aethalia of *F. intermedia*, 0.5-4 cm long, up to 10 mm thick and undamaged by insects were collected in the field. These aethalia were checked for other insect inhabitants using a stereo microscope, then placed in sterile Petri dishes and stored for two weeks to ensure absence of insects. Adult *Latridius hirtus* individuals were then added, and the dishes were regularly examined to follow development of the successive beetle stages.

Associations between myxomycetes and bryophytes were studied in the Crimean Nature Reserve. Special attention was paid to associations where slime moulds developed sporophores, aethalia or plasmodia directly on the surface of bryophyte thallus or inside it.

Associations between myxomycetes and wood-inhabiting fungi were studied in the Crimean Nature Reserve to identify the component species and assess the influence of substratum size on species diversity.

## RESULTS

### **Interactions between myxomycetes and *Latridiidae* beetles on woody substrata**

Associations between myxomycetes and the *Latridiidae* are shown in Table 1. These associations were found to be rather widespread on the left bank of Ukraine,

Table 1  
Myxomycetes associated with *Latridiidae* (Coleoptera)  
at the left bank of Ukraine

Myxomycete species, by order and family	Beetle species	Place and date of association collection	Beetle gut content
<b>Liceales, Reticulariaceae</b>			
<i>Tubifera ferruginosa</i> (Batsch) Gmel.	<i>Enicmus rugosus</i>	Lugansk oblast, s. Stan ychno Luganske, 1 July 2000	Spores of different myxomycete species including <i>T. ferruginosa</i>
<i>Reticularia lycoperdon</i> Bull.	<i>Enicmus fungicola</i>	Donetsk area, 6 June 2000	Spores of <i>R. lycoperdon</i>
	<i>E. rugosus</i>	Donetsk area, 26 May 2000	Spores of <i>R. lycoperdon</i>
	<i>Melanophthalma</i> sp.	Lugansk oblast, s. Stanychno Luganske, 29 June 2000	Spores of <i>Alternaria alternata</i> (Hyphomycetes)
<b>Trichiales, Trichiaceae</b>			
<i>Arcyria obvelata</i> (Oeder) Onsberg	<i>E. rugosus</i>	Lugansk oblast, s. Stan ychno Luganske, 30 June 2000	Spores of <i>A. obvelata</i>
<b>Stemonitales, Stemonitaceae</b>			
<i>Comatricha nigra</i> (Pers.) Schroet.	<i>E. rugosus</i>	Donetsk area, 18 August 2000	Spores of <i>C. nigra</i>
<i>Enerthenema papillatum</i> (Pers.) Rost.	<i>E. rugosus</i>	Donetsk area, 3 September 2000	Spores of <i>E. papillatum</i>
<i>Stemonaria longa</i> (Peck) Nann. Brem.	<i>E. rugosus</i>	Donetsk oblast, Volodarski district, nature reserve Kamiani Mogyly, 22 July 2000	Spores of <i>S. longa</i>
<i>Stemonitis axifera</i> (Bull.) Macbr.	<i>E. rugosus</i>	Donetsk oblast, Novo azovski district, Azovske forestry, 3 July 2000	Spores of <i>S. axifera</i>
<i>Stemonitis fusca</i> Roth	<i>Corticarina truncatella</i>	Lugansk oblast, s. Stan ychno Luganske, flood plain forest, 2 July 2000	Spores of <i>Alternaria alternata</i> (Hyphomycetes)
	<i>E. rugosus</i>	Lugansk oblast, s. Stan ychno Luganske, flood plain forest, 2 July 2000	Spores of <i>S. fusca</i>
<i>Stemonitis splendens</i> Rost.	<i>C. truncatella</i>	Donetsk, 3d city pond, 8 August 2000	Spores of <i>S. splendens</i>
	<i>E. rugosus</i>	Donetsk, 3d city pond, 8 August 2000	Spores of <i>S. splendens</i>
	<i>Latridius hirtus</i>	Donetsk, 3d city pond, 8 August 2000	Spores of <i>S. splendens</i>
<i>Symphytocarpus amaurochaetoides</i> Nann. Brem.	<i>E. rugosus</i>	Donetsk, Putylyvski forest, 18 July 2000	Spores of <i>S. amaurochaetoides</i>

Tab. 1 cont.

<b>Physarales, Physaraceae</b>			
<i>Fuligo intermedia</i> Macbr.	<i>E. rugosus</i>	Donetsk area, 3 September 2000	Spores of <i>F. intermedia</i>
	<i>L. hirtus</i>	Donetsk area, 3 September 2000	Spores of <i>F. intermedia</i>
<i>Fuligo septica</i> (L.) Wigg.	<i>E. rugosus</i>	Donetsk, Putyivski forest, 18 July 2000	Spores of different myxomycete species including <i>F. septica</i>
	<i>L. hirtus</i>	Donetsk, valley of the river Bakhmutka, 27 June 2000	Spores of <i>F. septica</i>
<b>Physarales, Didymiaceae</b>			
<i>Mucilago crustacea</i> Wigg.	<i>L. rugosus</i>	Donetsk area, 10 July 2000	Spores of different myxomycete species including <i>M. crustacea</i>
	<i>L. hirtus</i>	Donetsk area, 3 September 2000	Spores of <i>M. crustacea</i>

with 5 species from 4 *Latridiidae* genera being were found in the colonies of 13 species of myxomycetes from 10 genera.

Gut content analysis (from 5 of the *Latridiidae* species encountered in colonies of the 25 investigated samples of 13 myxomycete species) revealed slime moulds spores in the guts of 19 individuals (Tab. 1). In 16 of those cases, the spores found belonged to the slime mould species on which the beetle was collected. Spores of *Tubifera ferruginosa*, *Fuligo septica* and *Mucilago crustacea* were found in the guts of *Enicmus rugosus* specimens not originating from colonies of those species. Other spores distinct in morphology and size were also present in the guts of those *Enicmus rugosus* specimens. These may also have been myxomycete spores, but their taxonomic position was not established. No slime mould spores were found in the guts of female *Melanophthalma* sp. individuals found on *Reticularia lycoperdon* or from *Corticarina truncatella* individuals collected on *Stemonitis fusca*; instead, the guts of those beetles were filled with spores of the anamorphic fungus *Alternation alternata* (Fr.) Keisler.

Study of the living *Latridius hirtus*, kept in the laboratory on aethalia of *Fuligo intermedia*, resulted in the following sequence of observations: adults, laying eggs in the aethalia, larvae, moult, pupae, young adults emerging from pupae.

### **Co-existence of myxomycetes and bryophytes on woody substrata**

Altogether 13 species of myxomycetes were recorded on 9 species of moss and 3 species of liverwort associated with woody substrata in the Crimean Nature Reserve (Table 2). *Arcyria cinerea* (Bull.) Pers., *Echinostelium arboreum* Keller et Brooks, *E. minutum* de Bary, *Macbrideola cornea* (G. Lister et Cran) Alexop., *Perichaena vermicularis* (Schw.) Rost., *Physarum cinereum* (Batsch) Pers. were most commonly encountered in association with the following bryophytes widespread in montane Crimea: *Hypnum cupressiforme*, *Leucodon sciuroides* and *Porella platyphylla*. *Didymium*

*trachysporum* G. Lister on *Ctenidium molluscum*, *Licea minima* Fr. on *Hypnum cupressiforme*, *Perichaena chrysosperma* (Currey) Lister on *Frullania dilatata*, *Stemonitis fusca* on *Leucodon sciuroides*, *Symphytocarpus amaurochaetoides* on *Pterigynandrum filiforme*, *S. impexus* Ing. et Nann.-Brem. on *Porella platyphylla* and *Trichia varia* (Pers.) Pers. on *Anomodon viticulosus* were more rare in bryophytes communities of the reserve (Tab. 2).

Augmenting information about woody substrata in Table 2, *Echinostelium minutum* was collected on bark of *Acer stevenii* Pojark., *Alnus glutinosa* (L.) Gaertn., *Carpinus betulus* L., *Fagus sylvatica*, *Pinus kochiana* Klotzsch., *P. pallasiana*, *P. sylvestris* L., *Quercus petraea* (Mattuschka) Liebl., *Q. robur*, *Sorbus torminalis* (L.) Crantz., *Macbrideola cornea* was collected on bark of *Acer campestre* L., *Acer stevenii*, *Carpinus betulus*, *Cornus mas* L., *Crataegus* sp., *Fagus sylvatica*, *Fraxinus excelsior*, *Taxus baccata* L., *Tilia cordata* Mill. and *Perichaena vermicularis* was collected on bark of *Acer campestre*, *Alnus glutinosa*, *Cornus mas*, *Fraxinus excelsior*, *Pyrus communis* L., *Q. petraea*, *Taxus baccata*, *Tilia cordata*. On the reserve *Didymium trachysporum* was associated with *Ctenidium molluscum* but there are additional records of it on fallen conifer needles, pieces of small bushes and herbivore dung. *Physarum cinereum* was found on fallen leaves and decaying wood, but was most frequent in bryophyte communities as listed in Table 2.

Table 2  
Myxomycetes collected in the Crimean nature reserve on bryophytes

Species of myxomycetes	Species of bryophytes	Woody substrata
<i>Arcyria cinerea</i> (Bull.) Pers.	<i>Frullania dilatata</i> <i>Hypnum cupressiforme</i> <i>Porella platyphylla</i>	absence
<i>Didymium trachysporum</i> G. Lister	<i>Ctenidium molluscum</i>	
<i>Echinostelium arboreum</i> Keller et Brooks	<i>Leucodon sciuroides</i> <i>Hypnum cupressiforme</i>	
<i>Echinostelium minutum</i> de Bary	<i>Hypnum cupressiforme</i> <i>Leucodon sciuroides</i> <i>Pterigynandrum filiforme</i> <i>Racomitrium canescens</i>	absence
<i>Licea minima</i> Fr.	<i>Hypnum cupressiforme</i>	decaying wood
<i>Macbrideola cornea</i> (G. Lister et Cran) Alexop.	<i>Leucodon sciuroides</i> <i>Porella platyphylla</i> <i>Pterigynandrum filiforme</i> <i>Hypnum cupressiforme</i> <i>Metzgeria furcata</i>	absence
<i>Perichaena chrysosperma</i> (Currey) Lister	<i>Frullania dilatata</i>	
<i>Perichaena vermicularis</i> (Schw.) Rost.	<i>Hypnum vaucheri</i> <i>Porella platyphylla</i>	absence
<i>Physarum cinereum</i> (Batsch.) Pers.	<i>Anomodon viticulosus</i> <i>Homolothecium sericeum</i> <i>Hypnum cupressiforme</i> <i>Porella platyphylla</i> <i>Pseudoleskeella nervosa</i>	absence
<i>Stemonitis fusca</i>	<i>Leucodon sciuroides</i>	decaying wood
<i>Symphytocarpus amaurochaetoides</i>	<i>Pterigynandrum filiforme</i>	decaying wood
<i>Symphytocarpus impexus</i> Ing et Nann. Brem.	<i>Porella platyphylla</i>	
<i>Trichia varia</i> (Pers.) Pers.	<i>Anomodon viticulosus</i>	decaying wood

### Co-existence of myxomycetes and fungi on woody substrata

In the Crimean Nature Reserve, 69 species of myxomycetes and 66 species of fungi from the Ascomycota (36), Basidiomycota (21) and anamorphic fungi (9) were collected on decaying woody substrata of coniferous and broadleaf trees. All fungi recorded were species usually regarded as wood-inhabiting. Various complexes could be distinguished according to substratum type (Tab. 3) and range of associated species (Tab. 4).

Altogether 52 associations (Tab. 3) were encountered in the Crimean Nature Reserve between slime moulds and wood-inhabiting fungi, including 7 on large fallen trunks of trees, 34 on fallen branches of various diameters and 11 on decaying stumps. The highest numbers of myxomycete and the fungal species (from 13 to 35) occurred in associations on large fallen tree trunks. Species diversity was significantly less (from 2 to 8) in associations on fallen branches (from 5 to 15 cm in diameter) and on decaying stumps.

The following fungi were dominant (Tab. 4) in associations on small woody remnants and on large fallen decaying tree trunks. Pyrenomycetes: *Ceratostomella cirrhosa* (Pers.) Sacc., *Lasiosphaeria hirsuta* (Fr.) Ces. et de Not., *Melanomma pulvis-pyrius* (Pers.) Fuckel, *Nectria peziza* (Tode) Fr., *Rosellinia conglobata* (Fr. et Fuckel) Sacc.; discomycetes: *Bisporella citrina* (Batsch) Korf et S.E. Carp., *Mollisia melaleuca* (Fr.) Sacc., *Orbilia coccinella* (Sommerf.) Fr. These were associated with the following myxomycetes: *Arcyria denudata* (L.) Wettst., *Ceratiomyxa fruticulosa* (Mull.) Macbr., *Hemitrichia clavata* (Pers.) Rost., *Lycogala epidendrum*, *Metatrichia vesparium* (Batsch) Nann-Brem., *Physarum nutans* Pers., *Stemonitis fusca*, *Stemonitopsis typhina* (Wigg.) Nann-Brem., *Trichia decipiens* (Pers.) Macbr., *T. scabra* Rost. and *T. varia* (Pers.) Pers. The basidiomycete *Calocera cornea* (Batsch) Fr. and the anamorphic fungus *Brachysporium nigrum* (Link: Fr.) S. Hughes often joined those associations. *Fomes fomentarius* (L.: Fr.) Fr., *Stereum hirsutum* (Willd.) Fr., some species of the genus *Xylaria* Hill ex Schrank [particularly *X. longipes* Nitschke and *X. polymorpha* (Pers.) Grev.] were also found in those associations, but only on large fallen trunks. All listed species of myxomycetes and fungi formed the main body of associations on the woody substrata in the reserve. Frequency of occurrence in associations was high throughout the period of study.

Sometimes pyrenomycetes *Amphisphaeria umbrina* (Fr.) de Not., *Strickeria kochii* Koerb., *Lasiosphaeria ovina* (Pers.) Ces. et de Not., discomycetes *Mollisia cine-*

Table 3

Distribution of associated complexes between myxomycetes and xylotrophic fungi on the different types of woody substrates

Type of woody substrate	Number of associated complexes between myxomycetes and wood inhabiting fungi	Range of species from both groups in associated complexes
Large fallen tree trunks	7	From 13 to 35
Fallen branches of trees	34	From 2 to 6
Stumps	11	From 2 to 8
Total	52	133



Table 4  
Some complexes of myxomycetes and wood inhabiting fungi on woody substrata differentiated by size

Myxomycete species	Species of wood inhabiting fungi
Associated complexes on large fallen tree trunks	
<i>Arcyria denudata</i> <i>Fuligo septica</i> <i>Lycogala epidendrum</i> <i>Stemonitis fusca</i> <i>Stemonitis flavogenita</i> Jahn <i>Trichia scabra</i> <i>Trichia varia</i>	<i>Ceratostomella cirrhosa</i> , <i>C. ampullasca</i> (Cooke) Sacc., <i>Nectria coccinea</i> (Pers.) Fr., <i>N. peziza</i> , <i>Lasiochaeta hirsuta</i> , <i>L. ovina</i> , <i>Melanomma pulvis pyrius</i> , <i>Coniochaeta ligniaria</i> (Grev.) Masec, <i>Rosellinia conglobata</i> , <i>Xylaria longipes</i> , <i>Strickeria kochii</i> , <i>Dasyscyphus virgineus</i> S.F. Gray, <i>Bispora citrina</i> , <i>Mollisia melaleuca</i> , <i>Orbilium coccinella</i> , <i>Trichophaeopsis bicuspis</i> , <i>Calocera cornea</i> , <i>Fibuloporia mucida</i> , <i>Stereum hirsutum</i> , <i>Fomes fomentarius</i> , <i>Marasmius alliaceus</i> (Jacq.) Fr., <i>Oudemansiella</i> sp., <i>Armillaria mellea</i> (Vahl.) P. Kumm., <i>Pleurothecium recurvatum</i> , <i>Pseudosporopetes subuliferus</i> (Corda) M.B. Ellis, <i>Acrogenospora sphaerocephala</i> (Berk. et Broome) M.B. Ellis, <i>Graphium calicioides</i> (Berk.) Cooke et Masec
<i>Lycogala epidendrum</i> <i>Metatrachia vesparium</i> <i>Stemonitis fusca</i> <i>Trichia varia</i>	<i>Lasiochaeta hirsuta</i> , <i>Nectria peziza</i> , <i>Melanomma mastoidea</i> (Fr.) Schroet., <i>Melanomma pulvis pyrius</i> , <i>Chaetosphaeria ovoidea</i> Sacc., <i>Mollisia melaleuca</i> , <i>Bispora citrina</i> , <i>Xylaria polymorpha</i> , <i>Calocera cornea</i> , <i>Bjerkandera adusta</i> (Willd. Ex Fr.) P. Karst., <i>Stereum hirsutum</i> , <i>Fomes fomentarius</i> , <i>Pleurotus ostreatus</i> (Jacq.) P. Kumm., <i>Pleurothecium recurvatum</i> , <i>Periconia cambrensis</i>
<i>Arcyria denudata</i> <i>Comatrachia laxa</i> Rost. <i>Hemitrichia calyculata</i> (Speg.) Farr <i>Hemitrichia clavata</i> (Pers.) Rost. <i>Lycogala epidendrum</i> <i>Stemonitis fusca</i> <i>Symphytocarpus amaurochaetoides</i> <i>Trichia scabra</i> <i>Trichia varia</i>	<i>Melanomma pulvis pyrius</i> <i>Rosellinia conglobata</i> <i>Lasiochaeta hirsuta</i> <i>Bispora citrina</i> <i>Trichophaeopsis bicuspis</i> <i>Fomes fomentarius</i>
Associated complexes on fallen branches of trees	
<i>Trichia decipiens</i>	<i>Ceratostomella cirrhosa</i> , <i>Mollisia melaleuca</i> , <i>Orbilium coccinella</i> , <i>Microthelia incrustans</i> (El. et Ev.) Corlet et S. Hughes as anamorph <i>Dendryphiopsis</i> sp.
<i>Arcyria cinerea</i> (Bull.) Pers.	<i>Nectria coccinea</i> , <i>N. peziza</i> , <i>Eutypa lata</i> (Pers.) Tul. et C. Tul., <i>Orbilium coccinella</i> , <i>Trichophaeopsis bicuspis</i>
<i>Hemitrichia clavata</i> <i>Trichia decipiens</i>	<i>Rosellinia conglobata</i> , <i>Chaetosphaeria ovoidea</i> , <i>Mollisia melaleuca</i> , <i>Calocera cornea</i>
Associated complexes on the stumps	
<i>Comatrachia nigra</i> <i>Enerthenema papillatum</i>	<i>Hypocrea citrina</i> (Pers.) Fr., <i>Mollisia lignicola</i> Phill., <i>Stereum hirsutum</i>
<i>Ceratiomyxa fruticulosa</i> <i>Trichia decipiens</i>	<i>Lasiochaeta hirsuta</i> , <i>Amphisphaeria umbrina</i> , <i>Mollisia cinerea</i>
<i>Lycogala epidendrum</i>	<i>Bispora citrina</i> , <i>Stereum hirsutum</i> , <i>Brachysporium nigrum</i>

*rea* (Batsch) P. Karst., *Trichophaeopsis bicuspis* (Boud.) Korf et Erb., basidiomycetes *Schizopora paradoxa* (Schrad.: Fr.) Donk, *Phellinus ferruginosus* (Schrad.: Fr.) Pat., *Fibuloporia mucida* (Pers.: Fr.) Niemelä, anamorphic fungi *Pleurothecium recurvatum* (Morgan) Höhn., *Periconia cambrensis* E.W. Mason et M.B. Ellis, *Heteroconium chaetospora* (Grove) M.B. Ellis were also present on the numerous woody remnants

of various sizes, but these fungi were rare in comparison with the dominant species: they were often represented by a single record in the reserve.

## DISCUSSION

### Interactions between myxomycetes and *LatrIDIIDAE* beetles on woody substrata

Outside Ukraine, *Reticularia lycoperdon* (Liceales, *Reticulariaceae*), various species of the genus *Stemonitis* (Stemonitales, *Stemonitaceae*), in particular *S. fusca*, and more rarely *Fuligo septica* (Physarales, *Physaraceae*) are the most prevalent slime moulds in associations with the *LatrIDIIDAE*, while the genera *Enicmus* and *Dienerella* are the most wide-spread representatives of the family *LatrIDIIDAE* to be involved in associations with slime moulds (Newton, Stephenson 1990; Stephenson et al. 1994).

Within Ukraine, the same myxomycete and *LatrIDIIDAE* taxa were dominant in associations on the left bank. The myxomycete family *Stemonitaceae* (Stemonitales) was best represented, with 7 species from 5 genera, associated with 3 species of the *LatrIDIIDAE*, *Enicmus rugosus* being dominant (Tab. 1). Myxomycetes from the order Physarales were present in left bank Ukraine associations with the *LatrIDIIDAE* as follows (3 species from 2 genera and 2 families): *Fuligo intermedia*, *F. septica* (*Physaraceae*) and *Mucilago crustacea* (*Didymiaceae*). They were associated with 2 species of the *LatrIDIIDAE* – *E. rugosus* and *LatrIDIUS hirtus*. Both beetle species were associated with each of the 3 Physarales species. *Reticularia lycoperdon* (Liceales) was found from 3 locations in association with *Enicmus fungicola*, *E. rugosus* and *Melanophthalma* sp. (*distinguenda*-group - an exact species identification was not possible from the females collected). Interestingly, *R. lycoperdon* has been widely reported as the basic myxomycete component in associations with the *LatrIDIIDAE* in many parts of the world.

Among the myxomycetes connected with *LatrIDIIDAE* beetles in our left bank sampling sites, large aethalia (0.5-20 cm long and 0.1-3 cm thick) are typical for *Reticularia lycoperdon*, *Fuligo intermedia*, *F. septica* and *Mucilago crustacea*. Sporophores closely grouped in large and dense colonies are observed in species of the genus *Stemonitis* (especially *S. fusca* and *S. splendens*). Sporophores of *Symphytocarpus amaurochaetoides* tend to coalesce resulting in the formation of complex fruiting structures. The sporophores and aethalia of these slime moulds therefore provide sufficient spores to attract feeding by *LatrIDIIDAE* beetles. Together with *Stemonaria longa*, *Comatricha nigra* and some other species, they all have a long period of sporophore formation and the sporophores so formed then tend to be long-lasting. Furthermore, all of these slime mould species develop on woody substrata and have spores from 4-15 µm diam. with ornamented walls. They therefore have morphological, developmental and ecological characters which fit the criteria proposed by Blackwell (1984) for slime moulds associated with beetles.

The presence of myxomycete spores in *LatrIDIIDAE* gut contents shows that these beetles eat not only true fungi but also myxomycetes, while our observations of *LatrIDIUS hirtus* kept alive in the laboratory in defined conditions showed that at least one species is able to complete its life cycle, apparently normally, with only aethalia of one myxomycete species, *F. intermedia*, as food.

There are very few reports from other parts of the world of myxomycete feeding by *Latridiidae* beetles. A few individuals of an unknown species of *Enicmus* were collected on colonies of *Reticularia lycoperdon* from four locations in Himachal Pradesh (India): the gut contents of two were analysed and spores of *R. lycoperdon* found in both. A few individuals of a previously undescribed species of *Dienerella* were discovered on colonies of *Stemonitis fusca* from two locations in Himachal Pradesh: the guts content of an individual from each location were examined, and in both cases the guts were filled with spores of *S. fusca* (Newton, Stephenson 1990). Our results on for *Latridiidae* beetles and myxomycetes in Ukraine are entirely compatible with those observations. Our work therefore confirms that, in addition to the *Cerylonidae*, *Leiodidae*, *Scaphidiinae* (*Staphylinidae*), *Sphindidae* and some other beetle families, the *Latridiidae* also feed on myxomycetes.

There is some question about whether the *Latridiidae* are obligate or facultative feeders on myxomycetes. Newton and Stephenson (1990) considered their new species of *Dienerella* from India to be a facultative feeder. Results from the left bank of Ukraine suggest that *Latridius hirtus*, *Enicmus rugosus* and *E. fungicola* may be obligate feeders: *L. hirtus*, in particular, has been shown to be capable of completing its life cycle with only myxomycetes as a food supply. *Corticarina truncatella*, however, is more likely to be facultative: records on myxomycetes are rather rare, and it was usually found in forest litter, and on hay and other plant remains; furthermore gut content analysis shows this species can feed on fungal spores as well as those of slime moulds.

Use of myxomycetes by *Latridiidae* beetles extends beyond feeding: they may lay their eggs in sporophores, aethalia and plasmodia; large downy colonies of myxomycetes may provide good cover for larvae and adults. The absence of myxomycete spores in guts of *Melanophthalma* sp. *distinguenda*-group beetles from colonies of *Reticularia lycoperdon* suggests that the association between these beetles and myxomycetes is based on something other than a food chain - perhaps a spatial relationship - although that needs further study.

### Co-existence of myxomycetes with bryophytes on woody substrata

Decaying fallen trunks, logs, large branches, decomposing stumps are good substrata for simultaneous development of a diverse biota, including slime moulds, liverworts and mosses. Myxomycetes often occur on wood overgrown with bryophytes (Stephenson, Studlar 1985; Stephenson, Stempen 1994; Schnittler, Novozhilov 1998; Schnittler, Stephenson, Novozhilov 2000; Härkönen et al. 2002). Woody substrata covered with bryophytes provide favourable conditions for development of slime moulds, particularly on account of their high humidity (Stojanowska, Panek 2004; Novozhilov 2005).

All slime mould species found in the reserve associated with mosses and liverworts are usually considered lignicolous or corticolous: *Licea minima*, *Stemonitis fusca*, *Symphytocarpus amaurochaetoides* and *Trichia varia*, usually collected from decaying wood (Ing 1999), were also found on that substratum in the reserve. *Arcyria cinerea*, *Echinostelium arboreum*, *E. minutum*, *Macbrideola cornea*, *Perichaena vermicularis* and *Physarum cinereum* are known as typical corticolous species (Mitchell 1980). In our study area most of these corticolous species also were more often noted on

bark of various trees (Tab. 2). *Echinostelium arboreum* was the exception. In the reserve, every collection was made on moss, predominantly *Leucodon sciuroides* but once *Hypnum cupressiforme*.

*Didymium trachysporum* and *Physarum cinereum* are well documented as species developing on plant remains in the litter (Ing 1999), but there is also a report of their occurrence in association with bryophytes (Stephenson, Studlar 1985). Our results from Crimea match this pattern exactly. Some slime mould species (*Arctyria cinerea*, *Echinostelium arboreum*, *E. minutum*, *Macbrideola cornea*, *Perichaena vermicularis* and *Physarum cinereum*) developed on more than one bryophyte species. This may be evidence that some associations between myxomycetes and bryophytes are purely accidental, arising because slime moulds and bryophytes are both well-adapted to develop on the same substrata (bark or wood). The associations may, however, alternatively arise because bryophytes provide ideal protection and other ecological conditions in which slime moulds can complete their full life cycle from plasmodia to sporophores.

The relation between myxomycetes and bryophytes can be regarded as one of shared space. The bryophytes ensure temperature and humidity conditions suitable for slime mould development. Fluctuations of temperature and humidity are less inside the bryophyte thallomes than outside, so that the bryophytes function like a moist chamber for the slime moulds, providing favourable conditions for their development. This function benefits not only the myxomycetes but also the bacteria, algae and protozoa on which they feed (Stephenson, Stempen 1994). The myxomycetes produce sporophores and aethalia from plasmodia in the drier areas on the bryophyte open surfaces. Bryophytes associated with myxomycetes thus seem neither to benefit nor be damaged by them (Stephenson, Studlar 1985): bryophytes associated with slime moulds continue normal growth and reproduction, while the myxomycetes have a favourable biotope in the bryophytes communities. Interrelations between the bryophytes and the slime moulds are thus generally positive and may be classified as commensal: unilateral use of one species by another without damage.

### **Co-existence of myxomycetes and fungi on woody substrata**

Large fallen tree trunks might be thought to provide the richest associations of slime moulds and wood-inhabiting fungi, but in fact most associations (34) were recorded on fallen branches (Tab. 3). Direct contact of fallen branches with soil allows water to accumulate in the wood. Shade and grass cover makes loss of humidity from fallen branches and temperature fluctuations in the lower layer of forest vegetation less than from large fallen trunks and stumps. The hydrothermal regime of the fallen branches and their location positively influenced formation of a significant number of associations between myxomycetes and wood-inhabiting fungi. Such associations on woody substrata of various sizes and volumes are not directly connected and can exist independently. The biotic relations within those associations can therefore be characterized as neutral.

Species diversity in myxomycete / fungal associations on woody substrata can be compared according to size and volume of the substrata sampled (Tab. 4). One possible explanation for the correlation between species diversity and size (diameter)

of the woody substratum may be that larger woody remnants conserve water longer thus providing both myxomycetes and wood inhabiting fungi with better conditions for development. Substratum size may also determine the amount and availability of nutrients necessary for growth and formation of fruitbodies.

## CONCLUSIONS

Co-existence and interaction between slime moulds, beetles, bryophytes and fungi have specific features for every pair of interacting organisms: at one end, the *Latridiidae* may be either obligate or facultative feeders on slime moulds; at the other, relations between slime moulds and wood-inhabiting fungi are neutral. Further studies are necessary to clarify the mechanisms regulating these biotic relations.

**Acknowledgements:** We would like to thank Dr Tetiana Trikhleb from the National University in Donetsk (Ukraine) for identification of *Latridiidae*, and Dr Svitlana Nyporko from the Department of Lichenology and Bryology, M.G. Kholodny Institute of Botany NASU in Kyiv (Ukraine) who helped with identification of bryophytes. Our special thanks to David Minter (Egham) for his critical reading the paper and checking English.

## REFERENCES

- Blackwell M. 1984. Myxomycetes and their arthropod associates. *Fungus Insect Relationship: Perspectives in Ecology and Evolution*. N.Y. Columbia Univ. Press: 67-90.
- Blackwell M., Laman T.G., Gilbertson R.L. 1982. Spore dispersal of *Fuligo septica* (Myxomycetes) by latridiid beetles. *Mycotaxon* 14:58-60.
- Dudka I.O., Smitska M.F., Smyk L.V., Merezko T.O. 1976. Some theoretical problems of mycozoology. II. Consortia and the role of fungi in consortive relations. *Ukr. Bot. J.* 33 (2): 113-124 (in Ukrainian).
- Dudka I.O., Trikhleb T.A., Romanenko K.O. 2002. Associations of myxomycetes with the beetles (Coleoptera, *Latridiidae*). *Ecol. and Noospherol.* 12 (3-4): 54-64 (in Ukrainian).
- Gilbert H., Martin G. 1933. Myxomycetes found on bark of living trees. *Univ. Iowa Stud. Nat. Hist.* 15: 3-8.
- Härkönen M., Rikkinen J., Ukkola T., Enroth J., Virtanen V. et al. 2002. The communities of bryophytes, lichens and myxomycetes on bark of living trees in Hunan, Southern China. 4th Intern. Congress on Systematics and Ecology of Myxomycetes (August 4-9, 2002). *Abstract. Scripta Bot. Belg.* 22: 36-37.
- Howard E.L., Currie M.E. 1932. Parasitism of myxomycete plasmodia on the sporophores of hymenomycetes. *J. Arnold Arboretum* 13: 270-281.
- Ing B. 1999. *The Myxomycetes of Britain and Ireland. An Identification Handbook*. The Richmond Publishing Co. Ltd. 374 pp.
- Lawrence J.F., Newton A.F. 1980. Coleoptera associated with the fruiting bodies of slime molds (Myxomycetes). *Coleopterists Bull.* 14: 129-143.
- Mitchell D.W. 1980. *A Key to the Corticolous Myxomycetes*. Cambridge, England: The Brit. Mycol. Soc. 63 pp.
- Nannenga-Bremekamp N.E. 1991. *A Guide to Temperate Myxomycetes*. Bristol, England: Bion Press Ltd. 409 pp.
- Newton A.F. 1984. Mycophagy in Staphylinoida (Coleoptera). *Fungus Insect Relationship: Perspectives in Ecology and Evolution*. N.Y. Columbia Univ. Press: 302-353.
- Newton A.F., Stephenson S.L. 1990. A beetle/slime mold assemblage from Northern India (Coleoptera; Myxomycetes). *Oriental Insects* 24: 197-218.
- Novozhilov Yu.K. 1993. *An Identification Handbook of Russia. Division Slime Moulds. I. Class Myxomycetes*. SPb: Science. 288 pp. (in Russian).
- Novozhilov Yu.K. 2005. Myxomycetes (class Myxomycetes) of Russia: taxonomic composition, ecology and geography. *Autorev. of Dr of biol. thesis.* 48 pp. (in Russian).

- Rogerson C.T., Stephenson S.L. 1993. Myxomyceticolous fungi. *Mycologia* 85 (3): 456-469.
- Rücker W.H. 1983. Bunkócsápi Bogarak VII, Clavicornia VII: Merophysidiidae, *Latridiidae*, Dasyce-  
ridae. Magyarország Állatvilága (Fauna Hungariae) 158. Budapest. 68 pp.
- Schnittler M., Novozhilov Yu.K. 1998. Late autumn myxomycetes of the northern Ammer-  
gauer Alps. *Nova Hedwigia*. 66 (1-2): 205-222.
- Schnittler M., Stephenson S.L., Novozhilov Yu.K. 2000. Ecology and world distribu-  
tion of *Barbeyella minutissima* (Myxomycetes). *Mycol. Res.* 104: 1518-1523.
- Stephenson S.L. 1985. Slime molds in the laboratory II. Moist chamber cultures. *Amer. Biol. Teach-  
er* 47: 487-489.
- Stephenson S.L., Stempfen H. 1994. *Myxomycetes: A Handbook for Slime Molds*. Portland,  
Oregon: Timber Press. 183 pp.
- Stephenson S.L., Studlar S.M. 1985. Myxomycetes fruiting upon bryophytes: coincidence or  
preference? *J. Bryol.* 13: 537-548.
- Stephenson S.L., Wheeler Q.D., McHugh J.V., Frassinetti P.R. 1994. New North  
American associations of Coleoptera with Myxomycetes. *J. Nat. Hist.* 28: 921-936.
- Stojanowska W., Panek E. 2004. Myxomycetes of the nature reserve near Wałbrzych (SW Po-  
land) Part II. Dependence on the substrate and seasonality. *Acta Mycol.* 39 (2): 147-159.
- Wheeler Q.D. 1980. Studies on Neotropical slime mold/beetle relationships. Part I. Natural history  
and description of a new species of *Anisotoma* from Panama (Coleoptera: *Leiodidae*). *Proc. Ento-  
molog. Soc. Washington* 82: 493-498.

## Współistnienie i interakcje pomiędzy śluzowcami a innymi organizmami

### Streszczenie

W artykule podane są wyniki badań mikoekologicznych na terenie Krymu i Ukrainie Le-  
wobrzeżnej. Określono status taksonomiczny śluzowców, które współistnieją z owadami z ro-  
dziny *Latridiidae* (Coleoptera), mchami, grzybami: Ascomycota, Basidiomycota i grzybami  
anamorficznymi. Badania stosunków śluzowców i owadów z rodziny *Latridiidae* w stepowych  
regionach Ukrainy Lewobrzeżnej ukazały istnienie troficznego związku owadów i śluzowców.  
W koloniach 13 gatunków śluzowców, wśród których przeważały: *Stemonitis axifera*, *S. fusca*,  
*S. splendens*, *Fuligo septica* i *Mucilago crustacea*, występowało 5 gatunków owadów z rodziny  
*Latridiidae*. Analiza treści żołądkowej ukazała istnienie zarodników śluzowców w przewodach  
pokarmowych 19 z 25 przebadanych osobników. Stwierdzono, że owady *Latridium hirtus*, *En-  
cimus rugosus*, *E. fungicola* są myxomycetofagami obligatoryjnymi, a *Corticarina truncatella*  
może być uważany za myxomycetofaga fakultatywnego. 12 gatunków śluzowców podanych  
było z powierzchni 12 gatunków mchów, które rozwijały się na butwiejącym drewnie lub na  
korze w Krymskim Rezerwacie Przyrody. Stosunki śluzowców i mchów, wspólnie rosnących  
w Rezerwacie, na jednym drzewnym substracie są scharakteryzowane jako topiczne. Są one  
regulowane przez specyficzne mikroklimatyczne warunki, które tworzą się w środku mchów  
z wysoką wilgotnością. Stwierdzono wspólne występowanie 69 gatunków śluzowców z 36 ga-  
tunkami workowców, 21 gatunkami podstawczaków, 9 gatunkami anamorficznych grzybów.  
Analizowany skład asocjacji, które tworzą śluzowce z grzybami zależy od rozmiarów i objęto-  
ści substratów drzewnych.