SALINE STRESS EFFECT ON COWPEA BEANS GROWTH UNDER BIOFERTILIZER CORRECTION

EFEITO DO ESTRESSE SALINO NO CRESCIMENTO DO FEIJÃO-CAUPI SOB A CORREÇÃO DE BIOFERTILIZANTE

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ABSTRACT: New strategies to increase growth and food crops productivity in saline soils are priorities in the research. The experiment was carried out in a pot to investigate the growth of cowpea cv. Canapu in response to saline stress under the correction of a mixed biofertilizer. Five irrigation water salinity levels and three doses of the mixed biofertilizer were tested, with four replicates at each level. The increase in salinity level resulted in reductions in stem diameter, leaves number, shoot biomass and roots. The mixed biofertilizer use was able to minimize the effect of salinity in all analyzed variables, improving the cowpea growth and development, protecting from saline stress negative impacts. In addition, the mixed biofertilizer use may be an option to improve crop growth and productivity in salt affected soils.

KEYWORDS: Environmental stress. Water quality. *Vigna unguiculata*.L. Walp.

INTRODUCTION

Vigna unguiculata L. Walp is one of the main economically planted crops, being a cereal used as food for people and animals, with high nutritional value (QADOS, 2011).

It's considered moderately tolerant to salinity and may be cultivated in arid and semi-arid regions, such as the Brazilian northeast, which is limiting to other species. In these regions, most crops are grown under irrigation and inadequate irrigation management leads to secondary salinization, affecting 20% of irrigated land worldwide (GLICK et al., 2007; SHRIVASTAVA; KUMAR, 2015).

Salinity may affect plants in various ways such as drought stress, ionic toxicity, nutritional disorders, oxidative stress, altered metabolic processes, membrane disorganization, and reduced cell division and expansion (ZHU, 2007; SIDARI et al., 2008; MUSCOLO et al., 2013).

A saline soil is defined when the electrical conductivity (EC) of the saturation extract (CEe) in the root zone exceeds 4 dS m⁻¹ (approximately 40 mM NaCl) at 25 °C and exchangeable sodium of 15%, reducing yield of most of the plants grown in this CEe (MUNNS, 2005; JAMIL et al., 2011).

About 20 percent of total cultivated and 33 percent of irrigated farmland worldwide is affected

by high salinity (SHAHBAZ; ASHRAF, 2013). Salinized areas increase at a rate of 10% a year due to low rainfall, high surface evaporation, rocks weathering, saltwater irrigation and poor cultural practices, which can result in more than 50% salinized arable land by 2050 (JAMIL et al., 2011).

biofertilizer is any substance or A microorganism applied to plants with the aim of improve nutritional efficiency, tolerance to abiotic stress and/or quality characteristics of the crop, regardless of its nutrient content (DU JARDIM, 2015). This definition represents the clearest and most concise way to define biofertilizers. Currently, there are several biofertilizers, both industrial produced and those produced from waste. The latter prepared with animal, vegetable are and agroindustrial residues applied via soil, via irrigation systems or spread on the plants (YAKHIN et al., 2017).

The relevance of the liquid biofertilizers use as management practice in the form of simple or enriched microbial ferments is in the quantitative, diversity and in the nutrients availability by the biological activity, since these substances increase the efficiency of the nutrients use, reduce fertilizer consumption, stimulate plant development and growth (CALVO et al., 2014; HALPERN et al., 2015) and counterbalance stress factors, eventually

increasing crop quality and productivity (VAN OOSTEN et al., 2017).

The biofertilizer, when applied via soil in liquid form, improves the water infiltration rate and releases humic substances in the soil, increasing the plants osmotic adjustment by the accumulation of these substances, facilitating the water and nutrients absorption in saline environments (AYDIN et al., 2012; SOUTO et al., 2013).

Limited freshwater, growing food needs, the climate change pressure, and the land available reduction for cultivation are threats to agricultural sustainability (SHAHBAZ; ASHRAF, 2013). Practices for managing the saline water resources use may be an important resource for water scarcity (ZAHIR et al., 2008).

The aim of this study was to evaluate the salinity effect on the *V. unguiculata* vegetative growth, with and without mixed biofertilizer applications in saline soils.

MATERIAL AND METHODS

The work was carred out at the Piroás Experimental Farm, belonging to the Universidade da Integração Internacional da Lusofonia Afro-Brasileira (UNILAB), Piroás/ Redenção – Ceará, Brazil (04°15'55'S, 38°79'37''W; 240 m). The region climate is hot, dry semi-arid, with low cloudiness, high evaporation pattern and strong insolation (KÖPPEN, 1928). Rainfall is concentrated between January and April with average precipitation between 380 and 760 mm, distributed irregularly during the year (CELENTANO et al., 2017).

V. unguiculata cv. Canapu was cultivated in open-air in pots of 39.5 L in single rows with spacing of 0.5 x 0.5 m and planting density of 28,570 plants ha⁻¹. The vessels were filled with a 0.05 m layer of gravel and a substrate composed of sand and local soil in a 2:1 ratio.

The mean of irrigation water salinity was 0.4 dS m⁻¹, being considered the control treatment. Other concentrations were obtained by the addition of the NaCl, CaCl₂.2H₂O and MgCl₂.6H₂O salts, in the ratio 7:2:1 (RHOADES et al., 2000). Irrigation was started 15 days after emergence (DAE) and maintained until the end of the cycle by drip irrigation. The irrigation shift was of two days, whose water amount to be applied was calculated according to the evaporation of the class "A" tank.

The experimental design was in randomized blocks in the subdivided plots scheme, with four blocks. The plots were composed of water with five levels of salinity, 0.4; 1.4, 2.4, 3.4 and 4.4 dS m⁻¹, respectively, without leaching fraction and the

subplots consisted of three doses of mixed biofertilizer equivalent to 0, 400 and 800 mL plant⁻¹.

Mixed biofertilizer application was manually through graduated containers and divided twice a week, starting at the same irrigation application period. The biofertilizer was prepared with 100 L of fresh bovine manure, 30 L of chicken manure and 5 L of wood ash, diluted in 270 L of fresh water. The biofertilizer decomposition time was 30 days and monitored daily with EC, pH and temperature measurements. Cultural dealings followed the recommendations for the culture.

The biofertilizer after 30 days of decomposition had the following characteristics: N = 1.20; P = 0.38; K = 0.03; Ca²⁺ = 2.82; Mg²⁺ = 0.63 and S = 0.01 g L⁻¹. Na = 611; Fe = 91.54; Mn = 10, 56; Zn = 6.4 and Cu = 3.06 mg L⁻¹.

The stem diameter (DS) and leaves number (NL) were determined every 7 days between 15 and 57 DAE. The stem diameter was measured with digital caliper at the base of the stem 2 cm from the soil surface. The leaves number was quantified through weekly direct counting. The harvest occurred between 59 and 103 DAE and, at the cycle end, the fresh and dry mass of the underground and abovegorund parts were evaluated.

Fresh mass of aerial part (FMAP) and root (FRM) were determined with precision scale (model S1002 - BEL). Dry mass of aerial part (DMAP) and root (DRM) were obtained by drying in full sun until constant weight.

Statistical analysis, for all observed variables, was performed by analysis of variance (F test, 5% significance). When significant, regression analysis was performed. In the regression analysis, the equations that best fit the data were chosen based on the significance of the regression coefficients at the significance level of 5% (*) by the F test, and the highest coefficient of determination (\mathbb{R}^2). The software used was ASSISTAT - Statistical Assistance, version 7.7 beta (2016).

RESULTS AND DISCUSSION

The treatments effects on stem diameter (SD) and leaf number (NL) of cowpea beans as a evaluation times function, salinity levels and mixed biofertilizer doses indicated a significant effect ($p \le 0.05$) on the two analyzed variables, with exception of the leaves number that was not significantly affected by the biofertilizer doses. The interaction between the factors was significant ($p \le 0.05$) among all the treatments applied, except for the leaves number where the interaction times and

mixed biofertilizer doses were not significant (Table 1).

The treatments effect on SD and NL of common bean as a evaluation times function,

salinity levels and mixed biofertilizer doses suggests that plant growth was variable according to season, salinity level and mixed biofertilizer doses as reported for culture (SOUZA et al., 2014).

Table 1	1. Summary	of variance	analysis f	for stem	diameter	(SD) a	nd leaves	number	(NL) (of cowpea	plants
submitted to irrigation with different salinized water levels and mixed biofertilizer doses											

Source of Variation	đ	Mean squares				
Source of variation	ai	SD	NL			
Blocks	3	2.60210*	43.11609*			
Epoch (a)	6	80.82617*	908.62057*			
Error - a	18	0.13123	8.44498			
Salinity (b)	4	4.91688*	57.80803*			
Interaction a x b	24	1.35045*	39.96648*			
Error - b	84	0.34759	6.20613			
Biofertilizer (c)	2	8.36000*	11.19299 ^{ns}			
Interaction a x c	12	0.74877*	5.90504 ^{ns}			
Interaction b x c	8	1.21299*	18.76311*			
Interaction a x b x c	48	0.15352*	5.12509 ^{ns}			
Error - c	210	0.41337	6.55610			
Total	419	-	-			

* significant at 5% by the F test; (ns) not significant by the F test; df - Degree of freedom.

The stem diameter as a function of days after emergence, for each salinity and mixed biofertilizer level doses were adjusted to the increasing linear model. The largest stem diameter (6.74 mm) occurred at 57 DAE at 0.4 dS m⁻¹ and decreased with increasing salinity (Figure 1A-B).

The largest stem diameter at 0.4 dS m⁻¹ is due to the potentially toxic ions (Na⁺ and Cl⁻) low concentration in water and foliar tissues, without affecting the photosynthetic system and ensuring the CO₂ diffusion into the cells and consequently not harming the plant growth (LACERDA et al., 2003). The *V. unguiculata* reduction stem diameter occurred when irrigated with water at 1.55 dS m⁻¹ (PRAZERES et al., 2015). *V. unguiculata*,

SD (0.4) = 1.001 + 0.1006 DAE: R² = 0.97

SD (1,4) = 1,558 + 0,0832 DAE; R² = 0,97 SD (2,4) = 2,1657 + 0,0747 DAE; R² = 0,98

SD (3.4) = 2.9396 + 0.0589 DAF: R² = 0.91

0,065 DAE; R²

43

= 0,93

50

= 2,7344 +

36

Days after emergency (DAE)

8

7

6

5

4

3

2

15

Stem diameter (mm)

.

22

'Quarentinha' cultivar, had a decline in stem diameter under saline stress at 35 days after sowing (ANDRADE et al., 2013).

The highest stem diameters obtained with 400 and 800 mL plant⁻¹ week⁻¹ mixed biofertilizer doses are due to the nutritional availability of this fertilizer for the crop. The microbial and bioactive activity present in this input has the capacity to provide protection and resistance to the plant (GROVER et al., 2011). The stem growth increase provided by biofertilizers in saline environments was also observed in *Arachis hypogaea* L. and *Passiflora edulis f. flavicarpa* (SOUSA et al., 2012, OLIVEIRA et al., 2015).





Α

57

The leaves number reduced with increasing of saline water irrigation. The maximum

29

leaves number was 15.91 with 1.4 dS m⁻¹ (Table 1, Figure 2).



Figure 2. Number of cowpea leaves submitted to irrigation with different levels of salinized water.

The leaves number decrease with increasing irrigation water salinity probably occurred due to the sodium chloride accumulation in the cell walls and in the older leaves cytoplasm. At the same time, the vacuole sap can not accumulate more salt, reducing the concentration inside the cells, leading to rapid death and leaf fall (MUNNS, 2002). The leaf numbers reduction by saline stress impairs plant growth and development due to lower leaf area and net photosynthesis rate (GOMES et al., 2011; NUNES et al., 2012). The leaves number decreased in V. unguiculata plants irrigated with salt water, in soil with common bovine biofertilizer and enriched crab biofertilizer (SOUSA et al., 2014). The biofertilizer excess reduce plant biomass by toxicity could be the direct consequence of the chlorophyll synthesis inhibition and photosynthesis (KONG et al., 2017). Excessive salt and nitrogen amount from biofertilizer may cause decreased uptake of nutrient elements, inhibition of various enzyme activities,

induction of oxidative stress including alterations in enzymes of the antioxidant defense system and (WINGLER; ROITSCH. leaves fall 2008: ZIMMERMANN; ZENTGRAF, 2015). In addition to genetic control, senescence is regulated by joined actions of external (e.g., N availability, light) and internal (e.g., regulating metabolites, C/N ratio) signals (BAO et al., 2015; KONG et al., 2017) Sugar regulation of leaf senescence is also dependent on plant N status and soil N content. Therefore, excessive N and the desynchronization of N application with crop demand will acelerate leaves fall.

The FMAP, DMAP, FRM and DRM were significantly influenced by irrigation water salinity and mixed biofertilizer, except for DRM, which was significantly influenced by the mixed biofertilizer. There was no interaction between the factors (Table 2).

Source of variation GL		Mean squares					
		FMAP	DMAP	FRM	DRM		
Blocks	3	1276.90796 ^{ns}	9.86095 ^{ns}	9.04229 ^{ns}	0.99187 ^{ns}		
Salinity (a)	4	4518.95910*	77.29303*	10.12492*	0.53663 ^{ns}		
Error-a	12	389.67810	15.08145	2.98314	0.44052		
Biofertilizer (b)	2	3867.97993*	80.71499*	26.41949*	1.28499*		
Interaction - a x b	8	361.12130 ^{ns}	9.16537 ^{ns}	4.49979 ^{ns}	0.08366 ^{ns}		
Error-b	30	441.26835	9.91480	4.02946	0.15394		
Total	59	-	-	-	-		

Table 2. Summary of variance analysis for FMAP, DMAP, FMR, and DMR of cowpea plants submitted to irrigation with different levels of saline water and mixed biofertilizer doses

* significant at 5% by the F test; (ns) not significant by the F test; GL - Degree of freedom.

FMAP decreased linearly with increasing saline concentration of irrigation water (Figure 3A). The highest value of FMAP (69.15 g) was obtained in the control (0.4 dS m^{-1}) and the lowest value

 $(23.93\ g)$ in the highest salinity level was 4.4 dS m⁻¹. The FMAP increased linearly with the mixed biofertilizer doses (R² 0.87), with higher value

(59.55 g) obtained for the dose of 800 mL plant⁻¹

week⁻¹ (Figure 3B).



Figure 3. Fresh mass of aerial part (g) of cowpea plants submitted to irrigation with esulto f levels of salinized water (A) and mixed doses of biofertilizer (B).

The decrease in *V. unguiculata* plant growth is an energy metabolic cost reflection associated with an attempt to adapt the plant to salinity (DANTAS et al., 2002; MUNNS, 2002). The common bean lowest growth of an adaptive mechanism to the saline condition, so the plant maintains its vital activities, even to a limited extent (COELHO et al., 2013). The biofertilizer positive effect on the aerial part may be associated with the root growth stimulation by the phytohormones production, which allows the water roots uptake, improving plant nutrition and increasing nutrient availability (JOSKO et al., 2013).

DMAP decreased linearly with increasing irrigation water salinity ($R^2 = 0.82$) and increased linearly as a result of the increase in mixed biofertilizer doses ($R^2 = 0.76$) (Figure 4A-B).



Figure 4. Dry mass of aerial part (g) of cowpea plants submitted to irrigation with different levels of saline water (A) and mixed doses of biofertilizer (B).

The saline concentration increase in the soil solution causes a decrease in dry mass, which consequently increases the osmotic pressure, decreasing the absorption and the water supply, which are one of the main factors for a dry matter or biomass production (MACHADO; SERRALHEIRO, 2017). The reduction in dry matter may be due to the stomatal closure caused by salinity that limits CO₂ assimilation and inhibits leaf expansion, accelerating the mature leaves senescence, reducing the area destined to the photosynthetic process and the photoassimilates

production (LACERDA et al., 2003). The dry mass increase of the aerial part with the use of mixed biofertilizer prove the attenuation effect of this compound in *V. unguiculata* plants under salt stress due to slow nutrients release of the biofertilizer (FACHINI et al., 2004), besides promoting the soil aggregation and porosity, retention and water infiltration in the soil (RODRIGUES et al., 2013). The dry aerial part biomass of vigna bean cultivated in two soils classes (French-sandy and loamy clay) artificially salinized reduced 64.5% and 60.7% (COELHO et al., 2013), similar to the data found in

this research. The cultivar Quarentinha of *V. unguiculata* also had its dry biomass aerial part reduced by 66.94% with the increase of soil salinity irrigated with water at 5.0 dS m⁻¹ (LIMA et al., 2007), similar to the data found in this research.

The fresh root mass decreased with increasing salinity. The maximum fresh root mass

value was 5.95 g for the salinity level 0.4 dS m⁻¹ (Figure 5A). The fresh root mass increased linearly with the mixed biofertilizer doses, reaching a maximum value of 6.01 g with the dose of 800 ml plant⁻¹ week⁻¹ (Figure 5B).



Figure 5. Fresh root mass (g) of cowpea plants submitted to irrigation with different levels of saline water (A) and mixed doses of biofertilizer (B).

The fresh root mass decrease by saline stress is associated with the ions accumulation that concentrate in the soil (SHRIVASTAVA; KUMAR, 2015). The soil salt concentration increase by irrigation reduced root growth and consequently the weight of this variable in *V. unguiculata* plants (SILVA et al., 2011). Reduction in root growth and elongation in *Chenopodium quinoa* was caused by high salt levels at various development stages, reducing the supply of photosynthates from the shoot and morphological changes in the root, which should not be considered as a simple growth reduction, but rather as an induced growth reorientation to avoid stress (PANUCCIO et al., 2014).

The fresh mass increase of the roots with the mixed biofertilizer application of demonstrates the imput effect attenuation and may be considered an important management strategy for *V. unguiculata* plants cultivated in saline environment. The application of biofertilizer in the soil can induce an increase in the osmotic adjustment to the plants by the organic solutes accumulation, promoting the water and nutrients absorption under saline stress, improving the conditions for emergence, vegetative growth and biomass production of plants (BAALOUSHA et al., 2006).

Dry mass of the root increased linearly with the mixed biofertilizer application, with a maximum value of 1.32 g in the dose 800 mL plant⁻¹ week⁻¹ (Figure 6).





The increased linearly of root dry mass with the mixed biofertilizer application on V. unguiculata irrigated with saline water levels, is related to the humic substances increase of, improving soil quality, providing a source of nutrients and increasing cation exchange capacity and nutrient availability (JINDO et al., 2012). Biofertilizers improve soil structure and water retention (OLMO et al., 2016), while improving biological properties favoring plant-microorganism interactions. increasing water and nutrient uptake by plants (HAMMER et al., 2014). Although it was not included in the scope of the present study, a larger nodules number of nitrogen-fixing bacteria and with a higher finer roots number were observed in the plants root system that received the mixed biofertilizer.

CONCLUSIONS

V. unguiculata cv. Canapu proved to be very sensitive to saline stress, its growth and development being affected from 1.4 ds.m^{-1} .

The use of mixed biofertilizer was able to minimize the effect of salinity in all analyzed variables, improving the growth and development of cowpea cv. Canapu, protecting from negative impacts of saline stress. The incorporation of mixed biofertilizer may be a promising strategy to improve crop growth and productivity in salt affected soils.

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RESUMO: Novas estratégias para aumentar o crescimento e a produtividade de plantas cultivadas em solos salinos representam prioridades na pesquisa. O experimento foi conduzido em vaso, para investigar o crescimento do Feijão-Caupi em resposta ao estresse salino sob a correção de um biofertilizante misto. Cinco níveis de salinidade da água de irrigação e três doses do biofertilizante misto foram testados, com quatro repetições cada nível. O biofertilizante misto foi capaz de minimizar o estresse salino. O aumento do nível de salinidade resultou em reduções no diâmetro do caule, número de folhas, biomassa da parte aérea e das raízes. O uso do biofertilizante misto minimizou o efeito da salinidade em todas as variáveis analisadas, melhorando o crescimento e desenvolvimento do feijão caupi, protegendo-o dos impactos negativos do estresse salino. O uso de biofertilizante misto pode ser uma opção para melhorar o crescimento da cultura e a produtividade em solos afetados por sais.

PALAVRAS-CHAVE: Estresse ambiental. Qualidade da água. Vigna unguiculata L. Walp.

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