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Novel organophosphorus Schiff base ligands: Synthesis, characterization, ligational aspects, XRD and biological activity studies

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ABSTRACT: Six complexes have been synthesized from Cu(II), Ni(II), and Co(II) with new bidentate N_2 donor Schiff base ligand (2-methoxybenzalidene-1-phenylsemicarbazide L₁) and tridentate N_2O donor organophosphorus Schiff base ligand (2-methoxybenzalidenediphenylphosphate-1-

phenylsemicarbazide L_2). Both ligands were synthesized and characterized by metal analysis, infrared (IR), ultraviolet visible (UV-Vis), and nuclear magnetic resonance (NMR) spectral studies. The chemical structures of the synthesized complexes were characterized using their metal analysis, magnetic susceptibility, molar conductance, IR, and UV-Vis spectra. According to molar ratio studies, the complexes have the composition of ML_2 for L_1 and ML for L_2 . The X-ray diffraction (XRD) studies showed that the particle size of ligands and L_1 complexes were in nano-range. The ligands and their metal complexes have been screened for their antioxidant, antibacterial and antifungal activity.





1. Introduction

Schiff base compounds are commonly used as ligands in inorganic chemistry to create stable complexes with different transition metal ions (Abu-Dief et al., 2021). They are prepared by a condensation reaction of aromatic/aliphatic aldehydes and amines to form an azomethine group (CH=N) (Abu-Yamin et al., 2022). In previous research, many biological activities of Schiff base compounds have been documented including antibacterial, antifungal, antimalarial, antiproliferative, analgesic, anti-inflammatory, antiviral, antipyretic, and anticancer properties (Camellia et al., 2022; Ceramella et al., 2022; Ibeji et al., 2022; Jamil et al., 2023; Kaczmarek et al., 2018: Kafi-Ahmadi and Mariani, 2019: Song et al., 2020). A survey of the literature reveals a work devoted to the synthesis, characterization, and biological activities of many metal complexes of Schiff base from 2-methoxybenzaldyde (Camellia et al., 2022; Ibeji et al., 2022; Sani and Iliyasu, 2018; Yusof et al., 2015) In addition, organophosphorus compounds with transition metal ions have been the subject of several studies since complexes of such compounds exhibit biological activity (Abd El-Wahab and El-Sarrag, 2004; Brzezińska-Błaszczyk et al., 1996; El-khazandar, 1997; Ochocki et al., 1995). Hence, in the present study, a new Schiff base of 2-methoxybenzaldyde and 1-phenylsemicarbazide, its organophosphorus Schiff base, and their complexes with Cu²⁺, Ni²⁺, and Co²⁺ were prepared and characterized. The biological activities were investigated for these compounds, such as their antibacterial and antioxidant.

2. Experimental

2.1 Materials

The chemicals, 1-phenylsemicarbazide (Riedel-de Haën), 2-methoxybenzaldyde (Aldrich), diphenyl chlorophosphate (Riedel-de Haën), Triethyl amine (Riedel-de Haën), bipyridine (Aldrich), ferric chloride (Aldrich), ascorbic acid (BDH), sodium acetate trihydrate (BDH), glacial acetic acid (BDH) and metals chloride hydrate (CuCl₂.2H₂O, NiCl₂.6H₂O, and CoCl₂.6H₂O) were purchased from BDH. The solvents used were of spectroscopic grade.

2.2 Physical measurements

Elemental analysis for C, H, and N was performed with a Vario EL Fab. CHN Nr, at Central Laboratory, Faculty of Science, Cairo University, Giza, Egypt. Conductivity measurements were done with a Jenway conductivity meter model 4510 in dimethylformamide (DMF). The magnetic moments were determined by Gouy's method using a magnetic susceptibility balance from Johnson Metthey and Sherwood model. Ultraviolet visible (UV-vis) spectra were measured with (Specord200, Analytilk Jena, Germany) in the range of 200–900 nm, at Sana'a University. Fourier-transform infrared (FT-IR) spectra were recorded in transmission mode bv an (FT/IR-140. Jasco. Japan) spectrophotometer in the wavenumber range of 4000-400 cm⁻¹ (KBr was used as a matrix material for pellets). ¹H and ¹³C NMR spectra for ligands were performed in d₆-DMSO solvent using tetramethylsilane as an internal standard, at Cairo University, Giza, Egypt. All melting points reported for the compounds are measured in glass capillary tubes in Celsius degrees. Chloride was determined gravimetrically bv silver nitrate of coordinated (Vogel, 1961). The amount and molecules uncoordinated water was determined gravimetrically using the weight loss method (Vogel, 1961). The X-ray diffraction (XRD) patterns were obtained using XD-2 (Shimadzu ED-720) powder X-ray diffractometer at a voltage of 35 kV and a current of 20 mA using CuK(α) radiation in the range of 5° < 2 θ $< 70^{\circ}$ at 1° min⁻¹ scanning rate and a wavelength 1.54056 Å at Yemen Geological Survey and Mineral Resources Board.

2.3 Synthesis of ligands

2.3.1 Synthesis of Schiff base (2methoxybenzalidene-1-phenylsemicarbazide L_1)

The reaction mixture, including an ethanolic solution of amine (1-phenylsemicarbazide) (7.55 g, 0.05 mol) with an ethanolic solution of aldehyde (2methoxybenzaldyde) (6.80 g, 0.05 mol) was refluxed on hot reflux under constant stirring for 2-3 h, then the product was separated on cooling, it was filtered, washed several times with ethanol and ether, then the obtained crystals product was recrystallized by ethanol to give the corresponding Schiff base. The reaction is given in Fig. 1.





2.3.2 Synthesis of organophosphorus Schiff base (2methoxybenzalidenediphenylphosphate-1phenylsemicarbazide L_2)

The organophosphorus Schiff base L_2 was prepared by condensation between a solution of Schiff base (4.03 g, 0.015 mol) in dry benzene (50 mL) with a solution of diphenyl chlorophosphate (3.69 g, 0.015 mol) in dry benzene (50 mL) in 1:1 molar ratio in presence of triethyl amine (Fig. 2). After complete addition, the reaction mixture was heated under reflux for 2 h. The formed solid (triethyl amine hydrochloride) was filtered and the product was obtained after evaporation in a water bath.



2.4 Synthesis of metal complexes

2.4.1 Synthesis of Schiff base metal complexes

A hot ethanolic solution of metal chloride was added dropwise to an ethanolic solution of Schiff base $(2 \text{ L}:1 \text{ mol } \text{L}^{-1})$ molar ratio. The mixture was refluxed on a hot plate with stirring for 2–3 h. After evaporation of the solvent, the solid product was washed several times with ethanol and dried over anhydrous CaCl₂.

2.4.2 Synthesis of organophosphorus Schiff base metal complexes

A solution of metal chloride in 20 mL absolute ethanol was added dropwise to the solution of organophosphorus Schiff base in warm absolute ethanol (1 L: 1 mol L^{-1}). After the addition, the mixture was heated under hot reflux for three hours. The solution was evaporated, and the solid complexes were collected using ether.

2.5 Determination of the stoichiometry of the formed complexes of molecular structure (Molar ratio method)

The molar ratio method was described by the Molar ratio method (Yoe and Jones, 1944). The concentrations of metal ions (Cu^{2+} , Ni^{2+} , and Co^{2+}) were kept constant at (1×10^{-3} mol L⁻¹) in methanol and the concentration of ligands (L_1 and L_2) was regularly varied in methanol. The absorbance of the prepared solutions was measured at the constant wavelength (λ_{max}). The absorbance values were plotted versus the molar ratio [ligand] / [metal ion]. The intersections of the obtained straight lines indicate the molar ratio of the stable complexes.

2.6 Crystallinity and particle size from XRD

The percentage of crystallinity, XC (%) was calculated based on the integrated peak areas of the principal peaks (Shah *et al.*, 2006). The crystallinity of the complexes is calculated relative to the crystallinity of the ligands as a ratio (Eq. 1):

$$X_{C} (\%) = \frac{A_{complex}}{A_{ligand}} \times 100$$
 (1)

where $A_{complex}$ and A_{ligand} are the areas under the principal peaks of the complex and ligand sample, respectively.

X-ray diffraction was also used to determine the average particle size (D) which was estimated by the Scherrer equation (Akhtar *et al.*, 2015; Patterson, 1939) (Eq. 2):

$$D = \frac{\kappa \lambda}{\beta \cos\theta} \tag{2}$$

where **K** is Scherrer constant and equals 0.94, λ is the X-ray wavelength of Cu-K α radiations (1.5405 Å), β is full width at half maximum (FWHM) and θ is Bragg diffraction angle in degrees.

2.7 Antioxidant activity

The total antioxidant activity of the compounds has ferric-bipyridine been studied using reducing capacity (FBRC) (Al-Azab et al., 2023). It was taken (1 mL, 10⁻² mol L⁻¹ FeCl₃.6H₂O) to 10 mL volumetric flask different volumes of the standard antioxidant ascorbic acid (0.1 g L^{-1}) were added (0.01, 0.02, 0.04, 0.06, 0.08, 0.1, 0.2 mL). This was followed by 2.0 mL 0.3 mol L⁻¹ acetate buffer (pH 4) and 1.0 mL bipyridine $(6.4 \times 10^{-3} \text{ mol } \text{L}^{-1})$. The volume was completed to the mark with deionized water. After 10 min of incubation at room temperature, the absorbance was recorded against a blank at 535 nm. The absorbance values were plotted against the concentration of the various antioxidant solutions (Fig. 3). Similarly, 0.02 mL of (0.1 mg mL⁻¹ methanol) of each tested compound was reacted with 1 mL, 10⁻² mol L⁻¹ FeCl₃.6H₂O solution, $(6.4 \times 10^{-3} \text{ mol } \text{L}^{-1})$ bipyridine and 2.0 mL 0.3 mol L⁻¹ acetate buffer (pH 4). This mixture was diluted to 10 mL with deionized water for the antioxidant assay. The fixed reaction time and fixed state measurement method were used to find out the antioxidant activity of these in methanol compounds as a solvent. All spectrophotometric measurements were measured using UV-vis spectrophotometer (Specord200, Analytikjena, Germany) at Sana'a University.



Figure 3. Calibration curve of ascorbic acid with Fe-Bp complex.

2.8 Antimicrobial activity

The synthesis compounds were exanimated for their antimicrobial activity in the Laboratory of Microbiology at Sana'a University. They Germinated four types of bacteria (Staphylococcus aureus, Bacillus subtilis, Escherichia coli, and Pseudomonas aeruginosa) and one fungus (Candida albicans) on nutrient agar and Sabouraud agar solid media respectively. Using the filter paper disc method (Ericsson et al., 1960), it was prepared one concentration from each compound (1000 μ g mL⁻¹) in DMSO. Then added (100 µL) from each prepared compound on a filter paper (Whatman No. 1 filter paper, 5 mm diameter) which contained the bacteria with agar solid media, all the Petri dishes were put in an incubation period at 37 °C for 24 h. Gentamicin 120 µg mL⁻¹ was used as a reference substance for bacteria and Mycostatin 30 μ g mL⁻¹ for fungi. The results were registered by calculating the diameter of the inhibition zone (mm).

3. Results and discussion

The Schiff base ligand (L_1) was prepared by the condensation of 1-phenylsemicarbazide with 2methoxybenzaldyde in the molar ratio 1:1. The organophosphorus ligand (L2) was prepared by the condensation of diphenylchlorophosphate with the prepared Schiff base (L_1) in the molar ratio 1:1. The ligand ratio of (L_1) with Cu(II), Ni(II), and Co(II) complexes was found to be 1:2, and the ligand ratio of (L_2) with Cu(II), Ni(II), and Co(II) complexes were found to be 1:1. The molar conductance data of the complexes of L₁ are 18.44, 20.31, and 21.09 Ω^{-1} cm² mol⁻¹ in DMF $(1 \times 10^{-3} \text{ mol } \text{L}^{-1})$. These results show that these are non-electrolytes complexes (Aderoju and Sherifah, 2015; Geary, 1971), so the complexes of L_1 are insoluble in water and common organic solvents but soluble in polar solvents such as DMF and DMSO. The non-electrolytic nature of these complexes suggests that the chloride anions of the three salts have coordinated with the L_1 chelate of metal complexes. The reaction of the synthesized L_1 complexes with silver nitrate have been not given a white precipitate, and after digestion with nitric acid, the silver nitrate test gave a positive result. Furthermore, the results of molar conductance of

 L_2 complexes (73.75, 84.93, and 77.79) show that these complexes are 1:1 electrolytes in DMF (1 × 10⁻³ mol L⁻¹) (Geary, 1971; Shakdofa *et al.*, 2017). The complexes of L_2 are slightly soluble in water and soluble in common organic solvents. In the case of L_2 complexes, the precipitation was observed upon the addition of silver nitrate to the solution of the complexes (Vogel, 1961).

 L_1 and L_2 and their complexes were characterized by elemental analyses, FT-IR, ¹HNMR, and electronic spectra which are compliant with the proposed structures as shown in Fig. 4a and b. Table 1 lists some physical and analytical data on the ligands and their complexes.



Figure 4. Proposed structure of (a) L_1 complexes; (b) L_2 complexes.

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Compound	Color	M.P.		F.Wt	Elementa calculated		mental anal ulated% (fo	ysis und)	
•	(Yield)	(°C)	$(\Omega^{-1} \operatorname{cm}^2 \operatorname{mol}^{-1})$	(g mol ⁻¹)	С	Н	N	Р	М
$C_{12}H_{11}N_3O_2(L_1)$	White (70%)	174	-	269.30	66.90 (67.04)	5.61 (5.78)	15.60 (15.27)	-	-
C ₂₃ H ₁₀ N ₃ O ₅ P(L ₂)	Brown (75%)	153	-	479.36	65.15 (64.96)	2.94 (3.19)	8.76 (8.90)	7.05 (6.89)	-
[Cu(L1)2Cl2]·2H2O	Green (70%)	240	18.44	709.08	50.81 (51.30)	4.83 (4.88)	11.85 (12.08)	-	8.96 (9.00)
$[Ni(L_1)_2Cl_2]\cdot 2H_2O$	Green (50%)	250	20.31	704.22	51.16 (50.99)	4.87 (5.02)	11.93 (12.20)	-	8.33 (8.75)
$[Co(L_1)_2Cl_2]\cdot 2H_2O$	Dark blue (57%)	220	21.09	704.46	51.14 (51.36)	4.86 (4.33)	11.92 (11.68)	-	8.36 (8.13)
[Cu(L ₂)Cl (H ₂ O) ₂].Cl	Dark red (17%)	130	73.75	649.84	48.05 (47.85)	2.79 (2.86)	12.93 (13.18)	4.76 (4.55)	9.77 (9.53)
[Ni(L2)Cl (H2O)2].Cl	Red (19%)	108	84.93	644.98	48.41 (48.47)	2.82 (3.00)	13.02 (12.79)	4.80 (4.70)	9.09 (9.00)
[Co(L ₂)Cl (H ₂ O) ₂].Cl	Red (23%)	112	77.79	645.22	48.39 (48.01)	2.81 (2.57)	13.02 (13.24)	4.79 (4.99)	9.13 (8.98)

3.1 Nuclear magnetic resonance spectral studies

3.1.1 ¹HNMR spectra of the ligands

The ¹HNMR spectra of the ligands L_1 and L_2 (Figs. S1) and S2 see Supplementary Material) exhibited signals which are consistent with the proposed structure. The peaks at 6.019, 6.713, and 5.997, 6.692 ppm are assignable to the protons of the NH groups in the ligands L_1 and L_2 respectively (El-Tabl *et al.*, 2007; Shakdofa *et* al., 2017). Azomethine proton appeared as a singlet signal at 8.656 ppm (s, 1H) in L_1 (Cakir et al., 2003) and this signal disappeared in the L_2 spectrum due to a replacement reaction on this group with eliminated HCl and forming a P-C bond. The signals at 3.908 and 3.050 were assigned to (s, 3H, OCH₃) L_1 and L_2 , respectively (Cakır *et al.*, 2003). The peaks appearing in the range 6.742-7.831 and 6.717-7.842 ppm may correspond to protons of the aromatic hydrogen of L_1 and L_2 respectively (Galil et al., 2015).

3.1.2¹³CNMR Spectra of the ligands

The ¹³CNMR spectra (Figs. S3 and S4 see Supplementary Material) showed singlet signals at 161.688 and 160.421 ppm that attributed to carbonyl carbon C=O of the ligands L_1 and L_2 respectively (Carrasco *et al.*, 2020; Galil *et al.*, 2015). Also, the spectra exhibited two peaks at δ 149.517 and δ 149.623 ppm that were able to be appointed to azomethine carbon C=N of the ligands L_1 and L_2 , respectively (Shakdofa *et al.*, 2017). The peaks between 111.241–130.917 and 112.189–130.143 ppm is attributed to aromatic carbon in L_1 and L_2 , respectively (Galil *et al.*, 2015). The carbon of the methoxy group appeared at 55.534 and 45.381 ppm in ligands L_1 and L_2 , respectively (Yusof *et al.*, 2015).

3.2 IR spectra of the ligands and their complexes

Important IR bands of the ligands and their complexes are given in Table 2 and Fig. 5a. The presence of IR bands at 1,655 and 1,602 cm⁻¹ of ligand L₁ assigned to v(C=O)and v(C=N) respectively (Hossain *et al.*, 2019). The strong bands at 3,334 and 3,397 cm⁻¹ are assignable and attributed to v(NH) of this ligand (Nakamoto, 1998). The bands showed at 3,057, 2,958, and 1,267 cm⁻¹ belong to aromatic v(C-H), aliphatic v(C-H), and v(-C-N) stretching mode of vibrations, respectively (Jassem *et al.*, 2013). The band at 2,853 cm⁻¹ in its spectrum can be appointed to $v(-O-CH_3)$ (Mohapatra *et al.*, 2011).

The IR spectra of the complexes (Fig. 5a) are characterized by the appearance of broad bands at 3,424, 3,346, and 3,324 cm⁻¹ due to the water molecules in $L_1(Cu)$, $L_1(Ni)$, and $L_1(Co)$ complexes, respectively (González-García *et al.*, 2016). In the spectra of all the complexes, no change in the frequency of (C=O) but the 1,602 cm⁻¹ band was shifted to a lower frequency indicating the involvement of the N atom of the azomethine group in coordination (Table 2). The bands of v(NH, NH) disappeared in complexes due to the broad bands of water. New vibrations at 400–600 cm⁻¹ that are not present in the free ligand are attributed to the existence of v(M-N) and v(M-NH) (Yassin *et al.*, 2020). Based on the observations, it has been suggested that the ligand L_1 acts as bidentate.

Although the presence evidence of v(M-Cl) could not be brought in the IR data due to instrumental limitation, the insolubility of L_1 complexes in water and their nonelectrolytic nature evinced that $2Cl^{-}$ coordinated with metal in L_1 complexes.

In the ligand L_2 spectrum (Fig. 5b), the bands observed at 1,654 and 1,591 cm⁻¹ corresponds to v(C=O) and v(C=N), respectively (Hossain *et al.*, 2019). The presence of v(P=O) and v(P-O-C) is indicated by the appearance of bands at 1,188 and 1,072 cm⁻¹, respectively (Elkhazandar, 1997). The coordinated water molecules with metal ions in the complexes were proved by the bands 3,347, 3,407, and 3,312 (Aderoju and Sherifah, 2015). The IR spectra of the metal complexes of ligand L_2 (Fig. 5b) proposed that this ligand tridentate with the phosphoryl-oxygen, aminenitrogen, and azomethine-nitrogen due to the shift in the position of v(P=O), v(-NH) and v(C=N) (Table 2).

Table 2. The main IR absorption bands of the ligands and their complexes.

Compound	v(H2O)	υ(NH, NH)	v(-O-CH3)	υ(C=N)	υ(C=O)	υ(P=O)	υ(P-O-C)	υ(M-N)	υ(M-NH)
$C_{16}H_{14}N_4O(L_1)$	-	3398 3335	2853	1602	1655	-	-	-	-
C ₂₇ H ₁₃ N ₄ O ₄ P (L ₂)	-	3450 3347	2878	1591	1654	1188	1072	-	-
$[Cu(L_1)_2Cl_2]\cdot 2H_2O$	3424	3424	2850	1580	1651	-	-	527	428
[Ni(L1)2Cl2]·2H2O	3346	3346	2849	1571	1649	-	-	558	414
$[Co(L_1)_2Cl_2]\cdot 2H_2O$	3324	3324	2855	1577	1653	-	-	574	424
[Cu(L2)Cl (H2O)2].Cl	3421	3467 3452	2866	1560	1650	1165	1072	516	410
[Ni(L ₂)Cl (H ₂ O) ₂].Cl	3407	3407	2850	1617	1655	1160	1067	520	458
[Co(L ₂)Cl (H ₂ O) ₂].Cl	3312	3365 3312	2846	1603	1651	1161	1068	517	467



Figure 5. Infrared spectra. (a) L_1 and its complexes; (b) L_2 and its complexes.

3.3 Electronic spectra and magnetic measurements

The absorption spectra of the ligands L_1 and L_2 and their metal complexes were recorded in DMSO in the range of 200–900 nm (Table 3).

The ligands L_1 and L_2 exhibited two absorption bands (Figs. 6a and b) at 37,037.03, 35,714.28 and 33,333.33, 32,258.06 cm⁻¹ that have been allocated for the π - π * and n- π * transitions, respectively (Mohammed *et al.*, 2019). In the metal complexes, the transition of n- π * has been seen but they are shifted to another range. The shift in the transitions has been accounted for by the complexation between ligands with the metal ions. The Cu complexes of L_1 and L_2 exhibited single broad absorption peaks at 13,513.51 and 13,333.33 cm⁻¹ (Figs. 6a and b), respectively, attributable to the ${}^2Eg \rightarrow {}^2T_2g$ transition, indicating a distorted octahedral structure (Al-Maydama *et al.*, 2008; Gup and Kirkan, 2005).

The electronic spectra of the $L_1(Ni)$ and $L_2(Ni)$ complexes (Figs. 5a and b) showed absorption bands at 18,181.81, 15,151.51 and 17,543.85, 15,151.51 cm⁻¹, respectively, attributable to the ${}^{3}A_{2}g \rightarrow {}^{3}T_{1}g(P)$ and ${}^{3}A_{2}g \rightarrow {}^{3}T_{1}g(F)$ transitions, which is compatible with this complex having the octahedral structure (Gup and Kirkan, 2005).

The electronic spectra of the L₁ and L₂ complexes with Co (Figs. 6a and b) exhibited characteristic bands at 20,833.33, 15,873.01, 13,888.88 and 21,739.13, 20,000, 17,543.85 cm⁻¹ assigned to ${}^{4}T_{1}g \rightarrow {}^{4}T_{1}g(P)$, ${}^{4}T_{1}g \rightarrow {}^{4}A_{2}g$ and ${}^{4}T_{1}g \rightarrow {}^{4}T_{2}g(F)$ transitions. These bands are associated with the octahedral structures (Gup and Kirkan, 2005). This information with the effective magnetic moment (μeff) data (Table 3) of all complexes helped to support the suggested octahedral geometry (Al-Hakimi *et al.*, 2011; Fouda *et al.*, 2008).

Table 3.	Electronic s	pectral and	magnetic	moment	data of th	e ligands	s and their	· comp	olexes
			0			0			

	*	U		0		
Compound	μ _{eff} (B.M.)	$(\pi \rightarrow \pi^*),$ transition	$(n \rightarrow \pi^*)$ transition	d-d transition band (cm ⁻¹)	Assignments	Supposed structure
$C_{16}H_{14}N_4O(L_1)$	-	37,037.03	33,333.33	-	-	-
$C_{27}H_{13}N_4O_4P(L_2)$	-	35,714.28	32,258.06	-	-	-
[Cu(L1)2Cl2].2H2O	1.88	34,482.75	30,303.03	13,513.51	$^{2}\text{Eg}\rightarrow^{2}\text{T}_{2}\text{g}$	Distorted Octahedral
$[Ni(L_1)_2Cl_2]\cdot 2H_2O$	2.75	35,714.28	32,258.06	18,181.81 15,151.51	${}^{3}A_{2}g \rightarrow {}^{3}T_{1}g(P)$ ${}^{3}A_{2}g \rightarrow {}^{3}T_{1}g(F)$	Octahedral
$[Co(L_1)_2Cl_2]\cdot 2H_2O$	4.19	35,714.28	32,258.06	20,833.33 15,873.01 13,888.88		Octahedral
[Cu(L ₂)Cl (H ₂ O) ₂].Cl	1.61	37,037.03	33,333.33	13,333.33	$^{2}\text{Eg}{\rightarrow}^{2}\text{T}_{2}\text{g}$	Distorted Octahedral
[Ni(L2)Cl (H2O)2].Cl	2.39	35,714.28	31,250.00	17,543.85 15,151.51	${}^{3}A_{2}g \rightarrow {}^{3}T_{1}g(P)$ ${}^{3}A_{2}g \rightarrow {}^{3}T_{1}g(F)$	Octahedral
[Co(L2)Cl (H2O)2].Cl	4.77	35,714.28	31,250.00	21,739.13 20,000.00 17,543.85		Octahedral



Figure 6. UV-Vis spectra. (a) L_1 ligand and its complexes; (b) L_2 ligand and its complexes.

3.4 Molar ratio (stoichiometry) of the studied complexes

Investigation of the molecular structure of the complexes formed between the metal ions of $(Cu^{2+}, Ni^{2+}, and Co^{2+})$ with ligands in L_1 and L_2 in methanol using molar ratio (Figs. 7 and 8) revealed the formation of (1:2) (M:L) complexes for L_1 and (1:1) (M:L) complexes for L_2 under investigation. The results are depicted in Table 4.



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Figure 7. Molar ratio plot for the complexes between metal ions of (a) Cu, (b) Ni and (c) Co with L_1 in methanol.





Figure 8. Molar ratio plot for the complexes between metal ions of (a) Cu, (b) Ni and (c) Co with L_2 in methanol.

Table 4. Molar ratios for determination of stoichiometry of the L_1 and L_2 complexes in methanol.

\mathbf{L}_1									
	Absor	bance of com	plexes						
[L]/[M]	Cu(II)	Ni(II)	Co(II)						
	$\lambda = 330$	$\lambda = 310$	$\lambda = 310$						
0.5:1	0.121	0.067	0.043						
1:1	0.170	0.099	0.077						
1.5:1	0.195	0.143	0.117						
2:1	0.230	0.181	0.159						
2.5:1	0.232	0.185	0.174						
3:1	0.222	0.187	0.184						
3.5:1	0.218	0.182	0.190						
4:1	0.220	0.184	0.196						
	L_2								
	Absor	bance of com	plexes						
[L]/[M]	Cu(II)	Ni(II)	Co(II)						
	$\lambda = 300$	$\lambda = 320$	$\lambda = 320$						
0.25:1	0.290	0.200	0.313						
0.5:1	0.369	0.326	0.399						
0.75:1	0.450	0.480	0.483						
0.75:1	0.450 0.495	0.480 0.531	0.483 0.511						
0.75:1 1:1 1.25:1	0.450 0.495 0.492	0.480 0.531 0.555	0.483 0.511 0.530						
0.75:1 1:1 1.25:1 1.5:1	0.450 0.495 0.492 0.499	0.480 0.531 0.555 0.558	0.483 0.511 0.530 0.541						

3.5 X-ray diffraction

2:1

The XRD patterns of the L_1 and its complexes are shown in Fig. S5a–d (see Supplementary Material). The decrease of the XRD peak intensities of L_1 complexes can be attributed to the reduction in crystallinities due to the complexation. This is in good agreement with the lowering in relative crystallinity calculated for these complexes (the crystallinity calculation in Table 5 is based on the integrated area of principal peaks of the complex to the L_1 ligand obtaining a relative

0.509

0.571

0.544

crystallinity) (Shah *et al.*, 2006). The particle size calculated for L_1 and its complexes (2.074–5.552 nm) in the range of nanoparticle size—the nanoparticle size reported in the literature ranges between 1–100 nm (Boverhof *et al.*, 2015).

The particle size of L_2 ligand was found within nanometric range (10.555 nm), nevertheless its complexes appeared as an amorphous character (Fig. S5e-h, see Supplementary Material). From this change, it is expected to improved properties of L_2 complexes as compared with L_2 ligand.

Table 5. Data of the principal	l values of intensity of	f the ligands and L_1 c	complexes from XRD spectra.
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Compound	20	β	D (nm)	Mean D	X c (%)
	6.780	0.340	4.266		
	13.720	0.419	3.480		
	18.880	0.459	3.154		
L ₁	19.960	0.405	3.630	3.991	100
	23.920	0.379	3.905		
	28.220	0.295	5.061		
	29.900	0.337	4.447		
	12.799	0.120	12.142		
т	16.740	0.153	9.464	10 555	100
L_2	25.441	0.121	12.268	10.555	
	31.040	0.180	8.349		
	6.719	0.352	4.120		
	13.460	0.562	2.576		10.392
$[Cu(L_1)_2Cl_2]\cdot 2H_2O$	18.599	0.229	6.407	5.552	
_	20.919	0.237	6.213		
	31.659	0.178	8.455		
	16.279	0.439	3.332		
$[Ni(L_1)_2Cl_2]\cdot 2H_2O$	32.761	0.240	6.033	5.202	18.506
2 (-/= -] 2-	33.579	0.232	6.241		
	14.600	0.603	2.421		
$[C_{\alpha}(L_{\alpha}), C_{\alpha}] = 1.211 O$	35.520	0.762	1.995	2.074	2 100
$[CO(L_1)_2CI_2]^{-2}H_2O$	57.199	0.745	2.213	2.074	3.488
	57.579	0.868	1.668		

3.6 Antioxidant activity

The ligands and L_2 complexes were investigated using FBRC technique to determine the antioxidant activity (Al-Azab *et al.*, 2023).

Fe(II)-bipyridine complex produced by the reaction of

Fe(III) with antioxidant followed by bipyridine exhibited maximum absorption at 535 nm (Fig. 9). Total antioxidant activity is based on the redox reaction between compounds and Fe(III) at room temperature. The initial antioxidant concentration is indicated by the concentration of the oxidizing Fe(III).





Fe(II)- bipyridine complex Pink color $\lambda = 535 \text{ nm}$

Figure 9. Stoichiometry outline of FBRC method.

The results in Table 6 showed that these compounds were found to possess high potent antioxidant activity due presence of a combination of donor sites such as amide oxygen and imine nitrogen (Kostova and Saso, 2013).

Table 6. FBRC	values express	antioxidant	activity	as mg
L ⁻¹ for prepared	compounds to	$1 \text{ mg } L^{-1} \text{ of}$	ascorbic	e acid

L'in propurou compound	b to 1 mg L of useofficie uera
FBRC (mg L ⁻¹)	Sample
0.0248 ± 0.0001	L_1
0.0281 ± 0.0004	L_2
0.0201 ± 0.0002	[Cu(L ₂)Cl (H ₂ O) ₂].Cl
0.0183 ± 0.0002	[Ni(L ₂)Cl (H ₂ O) ₂].Cl [Ni(L ₂)Cl·(H ₂ O) ₂].Cl
0.0187 ± 0.0005	$[Co(L_2)Cl (H_2O)_2].Cl \\ [Co(L_2)Cl (H_2O)_2].Cl$

3.7 Antibacterial and antifungal screening for L_1 , L_2 and their complexes

The synthetic ligands and their metal complexes havbeen examined for their antibacterial and antifungal activities using the disk diffusion method (Ericsson *et al.*, 1960) against four types of Bacteria (*Staphylococcus aureus*, *Bacillus subtilis*, *Escherichia coli*, and *Pseudomonas aeruginosa*) and one fungus (*Candida albicans*) on nutrient agar and Sabouraud dextrose agar (SDA) solid media, respectively. The inhibition zone diameter of the compounds is shown in Table 7 and Fig. 10a–d. From these results, it was noted that:

- 1- No inhibition zone was observed for ligands and their metal complexes against the fungus (*Candida albicans*);
- 2- The metal complexes are more potent bactericides than the ligands;
- 3- The order of antibacterial activity for the compounds with *Bacillus subtilis* was $L_1(Cu) = L_2(Ni) > L_1(Ni) > L_2(Cu) > L_2 > L_1 > L_1(Co) = L_2(Co)$, with *Staphylococcus aureus* was $L_2(Ni) > L_1(Cu) > L_1(Co) > L_2(Co) > L_2 > L_1 > L_1(Ni) = L_2(Cu)$, with *Escherichia Coli* was $L_1(Co) > L_1(Cu) > L_2(Cu) > L_2(Co) > L_2 > L_1 > L_1(Ni) = L_2(Ni)$ and with *Pseudomonas aeruginosa* was $L_1(Co) = L_2(Co) > L_1(Cu) > L_2(Co) > L_2 > L_1 > L_1(Ni) = L_2(Ni)$ and with *Pseudomonas aeruginosa* was $L_1(Co) = L_2(Co) > L_1(Cu) > L_2(Ni) > L_2 > L_1 > L_1(Ni) = L_2(Ni) = L_2(Co) > L_1(Cu) > L_2(Ni) > L_2 > L_1 > L_1(Ni) = L_2(Cu)$.

In general, the complexes showed good antibacterial compared to the free ligands. Based on the chelation theory (Franklin and Snow, 1989), the increased inhibition activity of the metal complexes may be explained on the basis that their structures mainly possess C=N bonds. Besides, the coordination decreases the polarity of the metal ion due to the partial sharing of its positive charge with donor groups and possible π -electron delocalization inside the chelate ring-shaped during coordination that makes the complexes more lipophilic. This increased lipophilicity enhances the penetration of the metal complexes into lipid membranes, blocks the metal binding sites in the enzymes, and limits the further development of the organisms.

Table 7. Biological activities of the ligands and their metal complexes against bacteria and fungus (zone of inhibition in mm).

		Fungi			
Compound	grar	n-positive	gram-	negative	
(1000/mL ⁻¹)	Bacillus subtilis	Staphylococcus aureus	Escherichia coli	Pseudomonas aeruginosa	Candida albicans
L ₁	5	3	4	3	0
L ₂	6	5	6	5	0
$[Cu(L_1)_2Cl_2].2H_2O$	11	8	9	10	0
[Ni(L1)2Cl2].2H2O	10	0	0	0	0
[Co(L1)2Cl2].2H2O	0	7	10	15	0
[Cu(L ₂)Cl (H ₂ O) ₂].Cl [Cu(L ₂)Cl·(H ₂ O) ₂].Cl	7	0	8	0	0
[Ni(L ₂)Cl (H ₂ O) ₂].Cl [Ni(L ₂)Cl·(H ₂ O) ₂].Cl	11	10	0	7	0
[Co(L ₂)Cl (H ₂ O) ₂].Cl [Co(L ₂)Cl·(H ₂ O) ₂].Cl	0	6	7	15	0
Gentamicin (120 µg/ml)	29	25	23	28	-
Mycostatin (30 µg/ml)	-	_	-	-	18



Figure 10. Antibacterial activity of the ligands and their metal complexes against gram-positive bacterium (a) *Bacillus subtilis*; (b) *Staphylococcus aureus*; (c) *Pseudomonas aeruginosa*; and gram-negative bacterium (d) *Escherichia coli*.

4. Conclusions

In summary, the synthesis and physicochemical analysis were done for new ligands L_1 and L_2 with their complexes. In complexes, the ligand L_1 acts as neutralbidentate and L_2 acts as neutral-tridentate around the metallic ion. Electronic properties and magnetic susceptibility suggested octahedral geometry for all complexes. According to XRD, the ligands and L_1 complexes were in the nanoscale (2.074–10.555 nm). The ligands and L_2 complexes gave high antioxidant activity using FBRC technique. All compounds have been screened for their antibacterial and antifungal activity.

Authors' contribution

Conceptualization: Jamil, Y. M.; Al-Azab, F. M. Data curation: Al-Selwi, N. A. Formal Analysis: Jamil, Y. M.; Al-Selwi, N. A. Funding acquisition: Not applicable. Investigation: Jamil, Y. M.; Al-Azab, F. M.; Al-Selwi, N. A. Methodology: Jamil, Y. M.; Al-Azab, F. M.; Al-Selwi, N. A. Project administration: Jamil, Y. M.; Al-Azab, F. M. Resources: Jamil, Y. M.; Al-Azab, F. M.; Al-Selwi, N. A. Software: Al-Selwi N. A. Supervision: Jamil, Y. M.; Al-Azab, F. M. Validation: Jamil, Y. M.; Al-Azab, F. M. Visualization: Jamil, Y. M.; Al-Azab, F. M.; Al-Selwi, N. A. Writing - original draft: Al-Selwi, N. A. Writing – review & editing: Jamil, Y. M.

Data availability statement

All data sets were generated or analyzed in the current study

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Supplementary Materials



Figure S1. ¹HNMR spectrum of L₁.



Figure S2. ¹HNMR spectrum of L₂.



Figure S3. ¹³CNMR spectrum of L₁.





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Figure S5. XRD pattern of (a) L₁; (b) L₁(Cu); (c) L₁(Ni); (d) L₁(Co); (e) L₂; (f) L₂(Cu); (g) L₂(Ni); (h) L₂(Co);