

# UTILIZATION OF AGRO-WASTES IN BIOHYDROGEN FERMENTATION BY VARIOUS MICROORGANISMS

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Biohydrogen production based on agro-industrial wastes might be an attractive and effective technology for providing an energy source in the future. Dark fermentation is considered to be one of the most suitable biohydrogen formation processes. In this paper, various agro-industrial wastes as well as microorganisms applied for biohydrogen formation are reviewed.

Keywords: anaerobic process, dark fermentation, bioreactor

# 1. Introduction

Hydrogen produced biologically (biohydrogen) can be considered as a renewable source of energy [1]-[4]. Moreover, it is regarded as one of the most promising, environmentally-friendly "fuels" of the future due to its favourable characteristics, e.g., only water is formed by the combustion of H<sub>2</sub>, no environmental pollution is produced, highest energy density of 143 kJ/g. Agroindustrial residues seem to be suitable feedstocks for the production of biohydrogen [5].

Large amounts of solid and liquid waste are formed in agriculture every year, moreover, the majority of it is unutilized. However, promising technologies are available whereby agro-industrial residues are applied to produce value-added products and green energy by biological methods [6]. Liquid wastes are usually found in the form of wastewater. Solid agro-wastes can be classified into four groups [7]:

- (i) field residues
- (ii) waste of processing
- (iii) livestock waste
- (iv) chemical waste

Field residues generally originate from crop production and consist of the remains of crops, e.g., leaves, stalks, stems, seeds, straw, husks, shells, pulp, roots, woodland waste, etc.

Industrial food processing results in solid wastes which are mainly by-products of and leftovers from various plants, e.g., bagasse, sugarcane molasses, wheat bran, rice bran, groundnut shells, apple pomace, fruit peels, de oiled rice bran cake and oil cakes.

Recieved: 19 Sept 2022; Revised: 28 Sept 2022; Accepted: 28 Sept 2022

Livestock waste consists of by-products of slaughterhouses or originates from processing, e.g., bones, feathers, shells of crustaceans, as well as bedding/litter, carcasses and damaged feeders.

Chemical waste includes plant protection compounds, e.g., pesticides, insecticides and herbicides, as well as their containers and bottles. However, since these materials cannot be utilized in biological methods, this study focuses on the first three groups of solid wastes.

Biological hydrogen production from agroincludes industrial wastes dark fermentation, photofermentation and microbial electrolysis cells (MEC) [5]. Photofermentation is a process whereby organic substrates are converted into hydrogen by photosynthetic organisms under anoxic conditions. H<sub>2</sub> production in MECs is a bioelectrochemical process, where electrochemical and biological techniques are combined [8]. Given that dark fermentation is one of the most common methods used for hydrogen production from agro-wastes due to its higher hydrogen production rate, it is the focus of this mini-review.

# 2. Biohydrogen production by dark fermentation

The composition and certain characteristics of agrowastes are determining factors of biohydrogen production. Generally speaking, although these materials are rich in carbon sources, the majority of them require some sort of pretreatment. The lignin content in particular should be treated to increase the efficiency of

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biodegradation and conversion processes [5]-[7]. Pretreatments of agro-industrial residues include:

- (i) physical,
- (ii) chemical,
- (iii) physicochemical and
- (iv) biological methods.

Physical methods involve ultrasonication, ozonization, thermal- and microwave-assisted techniques as well as size-reduction processes, e.g., milling, grinding, etc.

Chemical methods include acid or alkali treatments, moreover, more recently organic solvents have been applied.

In physicochemical methods, a combination of physical and chemical techniques is used, e.g., steam

explosion, hydrothermal treatment and chemical treatment complemented with microwave irradiation.

Biological methods include enzymatic and microbial (whole cell) treatments, where mainly the degradation of biopolymers occurs.

Using these pretreatment methods, higher yields and more efficient hydrogen formation can be achieved.

Dark fermentative biohydrogen production can be carried out by facultative anaerobes and obligate microbes [1], [4]-[5]. In many cases, the suitable types of microorganisms are determined by the substrate applied and mixed microbial consortia are frequently used. Nevertheless, numerous microbes have still been used in dark fermentative biohydrogen production based on agro-wastes, examples of which are listed in *Table 1*:

Agro-waste	Microorganism	H <sub>2</sub> production rate or yield	Ref.
Sugar and rice mill wastewater	Acinetobacter junii	Rate 5.23 mLH <sub>2</sub> L <sup>-1</sup> h <sup>-1</sup>	[9]
Rice mill wastewater	Clostridium beijerinckii	Yield 214.9 mLH <sub>2</sub> L <sup>-1</sup>	[10]
Cheese whey powder	Enterobacter asburiae	Yield $1.19 \pm 0.01 \text{ molH}_2 \text{mol}^{-1}$	[11]
Sugarcane molasses	Eisenia fetida	Yield 1571.81 mLH <sub>2</sub> L <sup>-1</sup>	[12]
Cashew apple bagasse	Clostridium roseum	Yield 15 mmolH <sub>2</sub> L <sup>-1</sup> hydrolysate	[13]
Fruit and vegetable wastes + seawater	Thermotoga maritima	Rate 12.4 mmolh <sup>-1</sup> L <sup>-1</sup>	[14]
Fruit wastes (apple, banana,	consortia from biogas	Yield 523 mL/g VS (volatile solid)	[15]
grape, melon, orange)	sludge	_	
Sweet sorghum bagasse	Caldicellulosiruptor saccharolyticus	Yield 73.6 mLH <sub>2</sub> mmol <sup>-1</sup> C6 sugars	[16]
Cornstalk	Thermoanaerobacterium thermosaccharolyticum	Yield 89.3 mLH <sub>2</sub> g <sup>-1</sup> DB (dry biomass)	[16]
Distillers grains	Caldicellulosiruptor thermocellum	Yield 29.2 mLH <sub>2</sub> g <sup>-1</sup> DB	[16]
Fruit and vegetable wastes as well as corn stover	anaerobic sludge	Yield 1.91 molH <sub>2</sub> mol <sup>-1</sup> glucose	[17]
Beer lees	consortia from cow dung compost	Yield 68.6 mlH <sub>2</sub> /g TVS	[18]
Corn straw	Ethanoligenens harbinense	Yield 9 mLH <sub>2</sub> g <sup>-1</sup>	[19]
Wheat straw	consortia from cow dung compost	Yield 68 mLH <sub>2</sub> g <sup>-1</sup>	[20]
Grass silage	manure	Yield 16 mLH <sub>2</sub> g <sup>-1</sup>	[19]
Extracts of sweet lime peel	anaerobic mixed consortia	Yield 76.4 ml/g COD	[21]
Palm oil mill effluent	Thermoanaerobacterium- rich sludge	Yield 84.4 mLH <sub>2</sub> g <sup>-1</sup>	[22]
Beet pulp	mixed culture from anaerobic sludge	Yield 115.6 mLH <sub>2</sub> g <sup>-1</sup> COD	[24]
Sunflower stalks	Clostridium sp.	Yield 2.04 molH <sub>2</sub> mol <sup>-1</sup> eq. hexose	[25]
Miscanthus sinensis	Thermotoga elfii	Rate 23.99 mLH <sub>2</sub> h <sup>-1</sup>	[16]
Cassava pulp hydrolysate	mixed culture	Yield 342 mlH <sub>2</sub> g <sup>-1</sup> COD <sub>reduced</sub>	[26]
Tequila vinasse	Eisenia fetida	Yield 1246.36 mLH <sub>2</sub> L <sup>-1</sup>	[12]
Sugarcane bagasse	Eisenia fetida	Yield 232.72 mLH <sub>2</sub> L <sup>-1</sup>	[12]

As can be seen, although several types of agrowastes and microbes have been used for biohydrogen fermentation, the results regarding the yield or production rate of the processes are difficult to compare since their units vary and the data on hydrogen formation is diverse. Moreover, in this study, no information was collected about the treatment of the substrate (if any) nor which kind of bioreactor, set-up or mode of operation was used, making their comparison even more difficult. Therefore, only some special considerations can be discussed in this work.

#### 3. Special considerations

In addition to single microorganisms, mixed consortia are often applied in these processes, which originate from various sources [19], e.g., sludge from biogas production; anaerobic sludges from municipal wastewater treatment plants and cow-dung composts; cattle or dairy residue composts; sludge from palm oil mill effluent; soil, rice and straw composts; fermented soybean meal, etc. Even though these sources usually contain the microbes which are able to form biohydrogen, they need to be acclimated which includes some sort of pretreatment, e.g., heat shock, starving for a couple days, etc., to enrich the suitable microorganisms [3], [28] or sometimes bioaugmentation [10].

In hydrogen-producing consortia, a wide range of species have been isolated [19], e.g., under mesophilic conditions, *Clostridium (C. pasteurianum, C. saccharobutylicum, C. butyricum), Enterobacter (E. aerogenes) and Bacillus*, while under thermophilic conditions, the genera *Thermoanaerobacterium (T. thermosaccharolyticum), Caldicellulosiruptor (C. saccharolyticus, C. thermocellum)* and *Bacillus thermozeamaize*.

Regarding the utilization of certain fruit wastes, some flavor compounds were found to exhibit an antimicrobial effect [15], e.g., a citrus flavor, since D-limonene is able to weaken the fermentation even at very low concentrations (0.01% w/v). Therefore, the undesired effect of these substrates can be diminished by special treatments, e.g., encapsulation of the microbes in a membrane or pretreatment of the feedstock.

### 4. Conclusions

In biohydrogen production, the application of agroindustrial residues has a great potential since they are renewable, huge amounts of them are formed year by year and have not been utilized. Various types of agrowastes have been investigated with regard to biohydrogen production using a wide variety of microbes, namely single microorganisms or consortia. Although it seems that both can be effective in terms of fermentation, higher yields and/or production rates could be achieved when carbohydrates – which can be easily uptaken – are present in the initial substrate.

#### Acknowledgement

This work was supported by project no. RRF-2.3.1-21-2022-00009, entitled National Laboratory for Renewable Energy, which has been implemented with the support of the Recovery and Resilience Facility of the European Union within the framework of Program Széchenyi Plan Plus.

# REFERENCES

- Azwar, M. Y.; Hussain, M. A.; Abdul-Wahab, A. K.: Development of biohydrogen production by photobiological, fermentation and electrochemical processes: A review, *Renew. Sustain. Energy Rev.*, 2014, **31**, 158–173, DOI: 10.1016/j.rser.2013.11.022
- [2] Bharathiraja, B.; Sudharsanaa, T.; Bharghavi, A.; Jayamuthunagai, J.; Praveenkumar, R.: Biohydrogen and biogas – an overview on feedstocks and enhancement process, *Fuel*, 2016, **185**, 810–828, DOI: 10.1016/j.fuel.2016.08.030
- [3] Bakonyi, P.; Nemestóthy, N.; Simon, V.; Bélafi-Bakó, K.: Review on the start-up experiences of continuous fermentative hydrogen producing bioreactors, *Renew. Sustain. Energy Rev.*, 2014, 40, 806–813, DOI: 10.1016/j.rser.2014.08.014
- [4] Akhlaghi, N.; Najafpour-Darzi, G.: A comprehensive review on biological hydrogen production, *Int. J. Hydrogen Energy*, 2020, **45**(43), 22492–22512, DOI: 10.1016/j.ijhydene.2020.06.182
- [5] Saravanan, A.; Senthil Kumar, P.; Mat Aron, N. S.; Jeevanantham, S.; Karishma, S.; Yaashikaa, P. R.; Chew, K. W.; Show, P. L.: A review on bioconversion processes for hydrogen production from agro-industrial residues, *Int. J. Hydrogen Energy*, 2022, **47**(88), 37302–37320, DOI: 10.1016/j.ijhydene.2021.08.055
- Zakaria, Z. A.; Boopathy, R.; Dib, J. R. (Eds): Valorisation of agro-industrial residues - Volume I: Biological approaches, (Springer Nature, Switzerland), 2020, DOI: 10.1007/978-3-030-39137-9
- [7] Dey, T.; Bhattacharjee, T.; Nag, P.; Ritika; Ghati, A.; Kuila, A.: Valorization of agro-waste into value added products for sustainable development, *Bioresour. Technol. Rep.*, 2021, **16**, 100834, DOI: 10.1016/j.biteb.2021.100834
- [8] Cardena, R.; Kook, L.; Zitka, J.; Bakonyi, P.; Galajdova, B.; Otmar, M.; Nemestothy, N.; Buitron, G.: Evaluation and ranking of polymeric ion exchange membranes used in microbial electrolysis cells for biohydrogen production, *Bioresour. Technol.*, 2021, **319**, 124182, DOI: 10.1016/j.biortech.2020.124182
- [9] Murugan, R. S.; Dinesh, G. H.; Raja, R. K.; James Obeth, E. S.; Bora, A.; Samsudeen, N. M.; Pugazhendhi, A.; Arun, A.: Dark fermentative biohydrogen production by *Acinetobacter junii*-AH4 utilizing various industry wastewaters, *Int. J. Hydrogen Energy*, 2021, **46**(20), 11297–11304, DOI: 10.1016/j.ijhydene.2020.07.073

- [10] Rambabu, K.; Bharath, G.; Thanigaivelan, A.; Das, D. B.; Show, P. L.; Banat, F.: Augmented biohydrogen production from rice mill wastewater through nano-metal oxides assisted dark fermentation, *Bioresour. Technol.*, 2021, **319**, 124243, DOI: 10.1016/j.biortech.2020.124243
- [11] Alvarez-Guzman, C. L.; Cisneros-de la Cueva, S.; Balderas-Hernandez, V. E.; Smolinski, A.; De Leon Rodriguez, A.: Biohydrogen production from cheese whey powder by *Enterobacter asburiae*: effect of operating conditions on hydrogen yield and chemometric study of the fermentative metabolites, *Energy Rep.*, 2020, 6, 1170–1180, DOI: 10.1016/j.egyr.2020.04.038
- [12]Oceguera-Contreras, E.; Aguilar-Juarez, 0.: Oseguera-Galindo, D.; Macıas-Barragan, J.; Bolanos-Rosales, R.; Mena-Enriquez, M., Arias-García. A.; Montoya-Buelna, M.; Graciano-Machuca, O.; De León-Rodríguez, A.: Biohydrogen vermihumus production by associated microorganisms using agro industrial wastes as substrate, Int. J. Hydrogen Energy, 2019, 44(20), 9856-9865, DOI: 10.1016/j.ijhydene.2018.10.236
- [13] Silva, J. S.; Mendes, J. S.; Correia, J. A. C.; Rocha, M. V. P.; Micoli, L.: Cashew apple bagasse as new feedstock for the hydrogen production using dark fermentation process, *J. Biotechnol.*, 2018, 286, 71–78, DOI: 10.1016/j.jbiotec.2018.09.004
- [14] Saidi, R.; Liebgott, P. P.; Gannoun, H.; Gaida, L. B.; Miladi, B.; Hamdi, M.; Bouallagui, H.; Auria, R.: Biohydrogen production from hyperthermophilic anaerobic digestion of fruit and vegetable wastes in seawater: Simplification of the culture medium of *Thermotoga maritima*, *Waste Manage.*, 2018, **71**, 474–484, DOI: 10.1016/j.wasman.2017.09.042
- [15] Akinbomi, J.; Taherzadeh, M. J.: Evaluation of fermentative hydrogen production from single and mixed fruit wastes, *Energies*, 2015, 8(5), 4253–4272, DOI:10.3390/en8054253
- [16] Kumar, G.; Bakonyi, P.; Periyasamy, P.; Kim, S. H.; Nemestóthy, N.; Bélafi-Bakó, K.: Lignocellulose biohydrogen: Practical challenges and recent progress, *Renew. Sustain. Energy Rev.*, 2015, 44, 728–737, DOI: 10.1016/j.rser.2015.01.042
- [17] Rodriguez-Valderrama, S.; Escamilla-Alvarado, C.; Magnin, J. P.; Rivas-Garcia, P.; Valdez-Vazquez, I.; Rios-Leal, E.: Batch biohydrogen production from dilute acid hydrolyzates of fruits-and-vegetables wastes and corn stover as co-substrates, *Biomass Bioenergy*, 2020, **140**, 105666, DOI: 10.1016/j.biombioe.2020.105666
- [18] Fan, Y.-T.; Zhang, G.-S.; Guo, X.-Y.; Xing, Y.; Fan, M.-H.: Biohydrogen production from beer lees biomass by cow dung compost, *Biomass Bioenergy*, 2006, **30**(5), 493–496, DOI: 10.1016/j.biombioe.2005.10.009

- [19]Guo, X. M.; Trably, E.; Latrille, E.; Carrère, H.; Steyer, J.-P.: Hydrogen production from agricultural waste by dark fermentation: A review, *Int. J. Hydrogen Energy*, 2010, **35**(19), 10660– 10673, DOI: 10.1016/j.ijhydene.2010.03.008
- [20] Fan, Y.-T.; Zhang, Y.-H.; Zhang, S.-F.; Hou, H.-W.; Ren, B.-Z.: Efficient conversion of wheat straw wastes into biohydrogen gas by cow dung compost, *Bioresour. Technnol.*, 2006, **97**(3), 500–505, DOI: 10.1016/j.biortech.2005.02.049
- [21] Venkata Mohan, S.; Lenin Babu, M.; Venkateswar Reddy, M.; Mohanakrishna, G.; Sarma, P. N.: Harnessing of biohydrogen by acidogenic fermentation of *Citrus limetta* peelings: effect of extraction procedure and pretreatment of biocatalyst, *Int. J. Hydrogen Energy*, 2009, **34**(15), 6149–6156, DOI: 10.1016/j.ijhydene.2009.05.056
- [22]O-Thong, S.; Prasertsan, P.; Intrasungkha, N.; Dhamwichukorn, S.; Birkeland. N.-K.: Optimization simultaneous thermophilic of fermentative hydrogen production and COD reduction from palm oil mill effluent by Thermoanaerobacterium-rich sludge, Int. J. Hydrogen Energy, 2008, **33**(4), 1221-1231, DOI: 10.1016/j.ijhydene.2007.12.017
- [23] Ivanova, G.; Rákhely, G.; Kovács, K. L.: Thermophilic biohydrogen production from energy plants by *Caldicellulosiruptor saccharolyticus* and comparison with related studies, *Int. J. Hydrogen Energy*, 2009, **34**(9), 3659–3670, DOI: 10.1016/j.ijhydene.2009.02.082
- [24] Ozkan, L.; Erguder, T. H.; Demirer, G. N.: Effects of pretreatment methods on solubilization of beetpulp and bio-hydrogen production yield, *Int. J. Hydrogen Energy*, 2011, **36**(1), 382–389, DOI: 10.1016/j.ijhydene.2010.10.006
- [25] Monlau, F.; Aemig, Q.; Trably, E.; Hamelin, J.; Steyer, J.-P.; Carrere, H.: Specific inhibition of biohydrogen-producing *Clostridium* sp. after diluteacid pretreatment of sunflower stalks, *Int. J. Hydrogen Energy*, 2013, **38**(28), 12273–12282, DOI: 10.1016/j.ijhydene.2013.07.018
- [26] Phowan, P.; Danvirutai, P.: Hydrogen production from cassava pulp hydrolysate by mixed seed cultures: effects of initial pH, substrate and biomass concentrations, *Biomass Bioenergy*, 2014, 64, 1–10, DOI: 10.1016/j.biombioe.2014.03.057
- [27] Chatellard, L.; Marone, A.; Carrère, H.; Trably, E.: Trends and challenges in biohydrogen production from agricultural waste, in: Biohydrogen production: Sustainability of current technology and future perspective, A. Singh; D. Rathore (Eds.), (Springer, New Delhi), 2017, pp. 69–95, DOI: 10.1007/978-81-322-3577-4\_4
- [28] Rózsenberszki, T.; Koók, L.; Bakonyi, P.; Nemestóthy, N.; Bélafi-Bakó, K.: Comparative study on anaerobic degradation processes of pressed liquid fraction of organic solid waste, *Hung. J. Ind. Chem.*, 2021, **49**(1), 31–35, DOI: 10.33927/hjic-2021-05