Original Article

Treatment Failure in Cutaneous Leishmaniasis Patients Referred to the School of Public Health, Tehran University of Medical Sciences during 2008–2017

Zahra Kakooei¹; Homa Hajjaran¹; *Behnaz Akhoundi¹; Sorour Charehdar¹; Samira Elikaee¹; Zahra Shafeghat¹; Hamid Hassanpour^{1,3}; Mohammad Taghi Satvat¹; Elham Kazemi-Rad¹; *Mehdi Mohebali^{1,2}

¹Department of Medical Parasitology and Mycology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran

²Center for Research of Endemic Parasites of Iran (CREPI), Tehran University of Medical Sciences, Tehran, Iran ³Department of Medical Parasitology, School of Medicine, Ilam University of Medical Sciences, Ilam, Iran

*Corresponding authors: Dr Behnaz Akhoundi, E-mail: behnazakhoundi@yahoo.com, Dr Mehdi Mohebali, Email: mohebali@tums.ac.ir

(Received 05 Nov 2019; accepted 03 Dec 2020)

Abstract

Background: Cutaneous leishmaniasis (CL) is a vector borne disease predominantly found in tropical and subtropical countries, including Iran. For more than 6 decades, pentavalent antimonials have been used successfully worldwide for the treatment of leishmaniasis, but over the past few years, clinical resistance to these medications has increased. In this study, we evaluated CL patients who did not show any desirable responses to the anti-leishmanial treatment within a 10-year period (2008 to 2017).

Methods: All patients from different parts of Iran suspected of having cutaneous leishmaniasis, who were referred to the laboratory of leishmaniosis in Tehran University of Medical Sciences from 2008–2017 were parasitological examined. **Results:** During this period, a total of 1480 suspected CL patients were referred to the laboratory of leishmaniosis. Samples from 655 patients (70.8%) suspected of having CL were positive microscopically. The failure rate in patients treated with anti-leishmaniasis medications for a minimum of three complete treatment periods was 1.83% (12 cases).

There was no association between the number and size of skin lesions and patient characteristics. Also, the route of drug administration had no significant effect on the number and size of lesions.

Conclusion: In the present study, treatment failure was found in some confirmed CL patients treated with meglumine antimoniate. Over the past few years, it seems that had been increased in resistance to these medications. So, a review of the correct implementation of the treatment protocol and/or a combination therapy may be helpful in preventing an increase in the rate of treatment failure.

Keywords: Cutaneous leishmaniasis; Anti-Leishmania drug; Treatment failure; Iran

Introduction

Protozoan parasites of the genus *Leishmania* cause a wide spectrum of clinical manifestations known as leishmaniases. Cutaneous leishmaniasis (CL) is the most common forms of this disease in the world, with more than 350 million people at risk. There is an estimated incidence of 0.9–1.7 million new cases each year (1-2). In Iran, two forms of CL have been reported: zoonotic cutaneous leishmaniasis (ZCL) and anthroponotic cutaneous leishmaniasis (ACL). The main pathogen of ZCL in Iran is *Leishmania major* (*L. major*), whereas ACL is mainly due to *Leishmania tropica* (*L. tropica*) infections (3). For more than 6 decades, pentavalent antimonial (SbV) compounds, such as meglumine antimoniate (Glucantime®) and sodium stiboglucunate (Pentostam®), have been successfully used as the first-line treatment for all forms of leishmaniasis (4). In Iran, the national treatment protocol for cutaneous leishmaniasis recommends intramuscularly administered 20mg SbV5/kg body weight per

Copyright © 2020 The Authors. Published by Tehran University of Medical Sciences.

This work is licensed under a Creative Commons Attribution-NonCommercial 4.0 International license (https://creativecommons.org/licenses/by-nc/4.0/). Non-commercial uses of the work are permitted, provided the original work is properly cited.

day. The recommended treatment duration is 14 days for ZCL and 21 days for ACL (5). However, over the past few years, there has been increasing concern about the resistance of parasites to Glucantime® (6). Anti-Leishmania resistance and treatment failure is a major challenge in new and old-world countries (7-9). Resistance of *L. tropica* to Glucantime® was first reported in Iran in 2006 (10). Previously identified risk factors for the failure of treatment with Glucantime® are body weight above 68kg, previous anti-Leishmania treatment, having \geq 3 skin lesions, and failure to complete a course of treatment (11). In this study, we evaluated cases of treatment failure in CL patients who were referred to the leishmaniosis laboratory of Tehran University of Medical Sciences (TUMS) between 2008–2017 (10 years) (12, 13).

Materials and Methods

Patients

All patients suspected of having cutaneous leishmaniasis, who were referred to the leishmaniosis laboratory of TUMS between 2008-2017 were considered eligible for inclusion in this study. Parasitological confirmation was performed after gaining accurate information about the place of living of patients. After sterilizing the skin around the lesions/nodules with Ethanol 70%, a small incision was made in the margin of the lesion with a disposable lancet, and some tissue and exudates were removed by scraping. The scrapings from the margins of the lesions were air dried, fixed in absolute methanol, and stained with Giemsa 10%. The specimens were then examined for amastigotes demonstration by light microscopy with high magnification (14).

Design

The following data were obtained and recorded for each patient: age, sex, history of travel to endemic areas of leishmaniasis, number of lesions, size of the largest lesion, duration of infection, and adverse events. In this study, we focused more on CL patients who failed to respond to treatment, and who remained positive for smear prepared from the lesions after receiving at least two complete treatment courses. The necessary criteria for investigating cases of relapse, treatment failure, and clinical resistance were as follows:

Relapse: patients who received a topical or systemic treatment courses, whose outcome was improvement but the symptoms (any active lesion) reappeared in the original site of the lesion (15).

Treatment failure: cases in which the lesion remained active after four weeks of complete topical or systemic treatment course (15).

Clinical resistance: cases of relapse and treatment failure, in which active lesions persisted for weeks after at least two complete courses of systemic treatment (15).

According to the criteria above, 40 patients overall did not show any desirable therapeutic response to the following regimen of anti-*Leishmania* drugs:

- Glucantime®: systemic injection (20mg SbV5/kg for two weeks): (No= 8)

- Glucantime®: local injection 1–2ml of Glucantime® intralesionally injection around each skin lesion weekly for 4–6 weeks): (No= 0)

- Glucantime®: systemic and local injection (20mg per kg for two weeks): (No= 3)

- Miltefosine®: oral (2.5mg per kg daily for 28 days) and Glucantime®: (No= 4)

- Antibacterial compounds(Antibiotics): (No= 25)

Data analysis

The data were analyzed using SPSS software version 24. Chi-square and Fisher's exact test were used, and $p \le 0.05$ was considered significant for differences between groups.

Results

During the 10-year study period, 1480 individuals suspected of having CL were referred to the leishmaniosis laboratory. Of

these, 655 cases were confirmed positive for leishmaniasis by microscopic examination of samples (amastigotes were seen in microscopic examination). The maximum number of positive cases was recorded in 2018 (116 patients, 17.7 %), whereas the minimum number of positive cases was recorded in 2013 and 2016 (45 cases in each year, 6.9%) (Fig. 1). Out of the 655 positive cases, 464 (70.8%) were males and 191 (29.2%) were females. The youngest patients with confirmed infection was 2 months old and highest age among the positive cases was 80 years old (Fig. 2). About the nationality of the patients, 572 patients (87.3%) were Iranians and 83 (12.7%) were from Afghanistan. Overall, the majority of the patients, 397 individuals (60.6%), lived in Tehran but had a history of trip to endemic regions. Karaj (the capitol of Alborz Province) had the second highest number of cases (48 patients, 7.3%). On the other hands, the Highest number of lesions were one to four (Fig. 3) and also the most common site of the lesions was on the hands with 42/3 % (304) (Fig. 4). Among the patients who were referred to the leishmaniosis laboratory, 139 cases (21.2%) had relapse and were referred to the laboratory for re-examination. In the next step, for followup and understanding the phase of the disease, the patients were followed up. In 2008 and 2009, 20 patients with positive microscopic test were lost to follow-up due lack of contact information.

During 2010–2017, 40 cases were initially identified as treatment failure, but only 34 patients could be followed up due to lack of contact information. Out of the 34 patients who were followed up, 22 patients improved by modifying the treatment regimen, such as reinjection of Glucantime®, use of supplementary drugs such as Miltefosine® capsule, use of ointments, and other traditional drugs. Among the remaining 12 cases, even with modification of diet and increase in drug dosage, 7 cases did not show any desirable response to treatment (58.3%) and five cases had treatment failure (41.7%) (Table 1, Fig. 5). Among these 12 individuals with treatment failure, the voungest and oldest were 16 and 62 years old, respectively. Among the patients with treatment failure, the shortest duration of disease was six and the longest was 312 months, respectively.

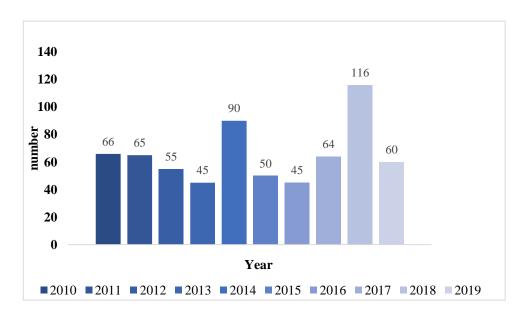


Fig. 1. Distribution of confirmed CL cases referring to leishmaniasis laboratory Tehran University of Medical Sciences during 2008–2017

J Arthropod-Borne D)is, December 1	2020, 14(4): 363–375

No	Age (year)	Living place	Travel place during two past years	Duration of persis- tence of lesion	Site of lesion	Type of administered drug and the type of injection
1	62	Tehran	Firoozkuh	3 years	leg	21 systemic Glucantime®
2	22	Iranshahr	Unknown	6 years	leg, ankle, toes	200 systemic Glucantime® 60 local Glucantime® 56 oral Miltefosine®
3	47	Hesarak Ka- raj	Qom	4 years	hand	76 systemic Glucantime®
4	30	Qom	Kashan	3 years	hand	100 systemic Glucantime®
5	27	Neyshabour	Unknown	13 years	Face, nose	300 systemic Glucantime® 100 local Glucantime® 70 oral Miltefosine®
6	36	Tabriz	Isfahan	3 years	Ear, eyes	25 systemic Glucantime® 25 local Glucantime® oral Miltefosine®
7	30	Dehloran	Unknown	13 years	back	23 systemic Glucantime®
8	20	Karaj	Afghanistan	6 years	face	200 systemic Glucantime® 100 local Glucantime®
9	16	Mashhad	Unknown	9 years	In nasal mucosa	84 systemic Glucantime® 90 oral Miltefosine®
10	25	Tehran	Afghanistan	26 years	The entire body	21 systemic Glucantime®
11	16	Tehran	Afghanistan	1 year	face	21 systemic Glucantime®
12	43	Unknown	Pakistan	6 months	face	42 systemic Glucantime®

Table 1. The characteristics of patients with no responses to anti-*Leishmania* drugs referring to leishmanisis laboratory Tehran University of Medical Sciences during 2008–2017

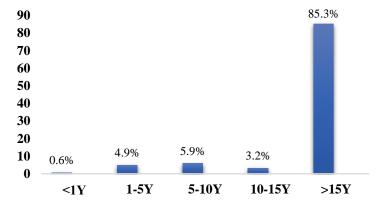


Fig. 2. Frequency of confirmed CL patients referring to leishmaniasis laboratory Tehran University of Medical Sciences by age groups during 2008–2017

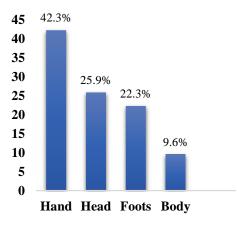


Fig. 3. Distribution of skin lesions on the bodies of the confirmed CL cases referring to leishmaniasis laboratory Tehran University of Medical Sciences during 2008–2017

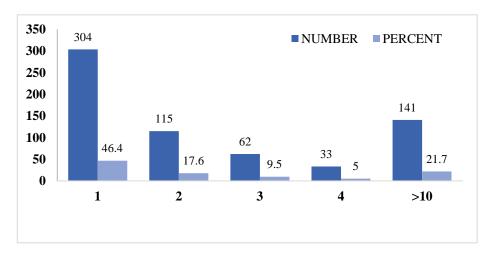


Fig. 4. Distribution of the number of lesions in confirmed CL patients referring to leishmaniasis laboratory Tehran University of Medical Sciences during 2008–2017



Fig. 5. Skin lesions with no responses to anti-*Leishmania* drugs in two CL patients referred to leishmaniasis laboratory Tehran University of Medical Sciences

Discussion

In this study, 40 cases were initially confirmed as treatment failure, but only 34 could be followed up. Out of this, 22 improved by changing therapeutic regimen such as reinjection of Glucantime[®], use of supplementary drugs such as Miltefosine[®] capsule, use of ointments, and other unknown drugs (12, 13, 15). However, in the remaining 12 cases, even by changing the therapeutic regimen and raising the drug dose, they did not show any desirable response and treatment failure was observed (Table 1).

Furthermore, in the present study, among treatment resistant patients, the longest course of the disease was 312 months (26 years), highlighting the importance of factors such as immune system of the patient, drug kinetics, and existence of resistant strains (11). There have been various reports on the incidence of drug resistance in different foci of leishmaniasis in Iran. Specifically, treatment failure rate in anthroponotic cutaneous leishmaniasis due to L. tropica infection has been reported to be 10.8 % in Mashhad and 11.1% in Bam. In Isfahan, one of the main foci of ZCL, drug resistance was reported as 11.6% in 2005 and in 2013, 3.7% failure rate for topical injection, 4.7% for systemic injection, and 3.4% for concurrent use of both treatment methods were reported (14, 17).

One of the mechanisms leading to the reduced response clinical forms of leishmaniases to the antimony pentavalent compounds is the development of acquired resistance to the drug. Although the antimony pentavalent compounds have been used for several decades as the firstline treatment for all clinical forms of leishmaniasis, unfortunately the therapeutic effect of these compounds has been jeopardized with the emergence of resistance strains in most endemic regions (16). Specifically, in India, Sudan, Latin America, Europe, and Middle East, drug resistance is an important threat to effective treatment of the clinical forms of Leishmaniasis. For example, in India, over 60% of cases of visceral leishmaniasis caused by *Leishmania donovani* do not respond to treatment, which can be due to different reasons such as the development of drug resistance by the parasite, immunologic changes of the patient, ineffective treatment regimen mainly due to lack of patient compliance (23, 24).

In this regard, different studies have corroborated the idea that the resistance to the antibiotics could be acquired. To investigate the existence of acquired drug resistance in Bihar region in India, Lira et al. isolated *L. donovani* from drug responsive patients and those who were confirmed as treatment failure. Using the presence of intra-macrophage amastigote (invitro) in isolates as criteria for resistance, they found that the isolates from patients who had responded to treatment were three times more sensitive to Sodium Stibogluconate drug in comparison to drug-resistant isolates. These results confirm the existence of acquired resistance in India (25).

In other studies conducted in France, acquired resistance in clinically resistant *Leishmania infantum* isolates was confirmed in drug resistance tests under in-vitro conditions (26). Also, studies in Latin American countries such as Columbia using in-vitro tests indicated that some cases of treatment failure in new-world cutaneous and mucocutaneous leishmaniasis have been due to development of resistant strains (24-28). Similarly, in Iran, studies in 2004 and 2006 on 185 patients with ACL in Mashhad reported treatment failure in 20 cases (10.8%).

In these studies, drug resistance of isolates was confirmed under in vitro conditions through the culture of macrophages. The isolates which were clinically resistant to Glucantime[®] were also resistant in drug sensitivity tests, and required higher doses for their elimination. In the anthroponotic form of the disease, the probability of spread of drug resistant *L. tropica* strains across human populations is very high, and it's of significant public health importance (17).

Also, acquired resistance of *L. infantum* and *L. major* with zoonotic cycles have been reported in Iran, and have been confirmed under in-vitro conditions. The progression of anti-monial resistance in anthroponotic forms such as in India resulting from *L. donovani* and in Iran due to *L. tropica* infections suggests that in the future, resistance to other anti-leishmaniasis drugs such as Miltefosine® and Amphotericin B may also develop in case of extensive usage (11).

Considering the fact that drug resistance is one of the major challenge in the successful treatment of the disease, identification of the mechanisms involved in drug resistance can be useful in improving therapeutic strategies. Furthermore, identification of suitable markers for monitoring and detecting drug resistant cases and predicting the course of development of resistance in endemic regions could be helpful in this regard. Among the most important reasons of resistance are genetic, protein, and enzyme factors, as well as intracellular factors such as signaling pathways and apoptosis. The initial mechanisms in the development of drug resistance include reduction of drug concentration in the parasite through a decreased uptake or increase in drug excretion through cell pumps, deactivation of drug, and inhibition of drug activation in the cell. A recent research indicated that other than these typical mechanisms, other factors such as apoptosis and signaling pathways are also involved in the development of natural drug resistance. Accordingly, today various methods such as real-time RT-PCR, microarray, and proteomic methods are used to detect the factors and genes affecting clinical resistance phenomenon (27, 28).

In Iran, a study conducted by Kazemi Rad et al. (2013) on *L. tropica* to detect genes whose expression is different between sensitive and resistant *L. tropica* using cDNA-AFLP technique confirmed that 13 genes play a major role, the most important of which were as follows: Aqua Glycero Porine (AQP1), affects drug uptake; Multi Drug Resistance Protein A (MRPA), involved in entrapping the drug; Phospho Glycerate Kinase (PGK), involved in carbohydrate metabolism; ubiquitin, involved in degrading oxidized proteins; Amino Acid Permease (AAP3), involved in uptake of arginine amino acid; protein kinase (PK), involved in signaling pathways; mitogen activated protein kinase (MAPK); and protein tyrosine phosphatase (PTP), involved in phosphorylation pathway. Also, in their study, using Quantitative Real Time PCR (QRT PCR) method, it was found that in resistant isolates, there was an increased expression of AAP3, ubiquitin, PGK, PTP, and MRPA, whereas AQP1 and MAPK had diminished expression (29, 30).

In another study conducted by Zaeran et al. (2015) on *L. major* resistance and sensitive to Glucantime[®] using two-dimensional electrophoresis method performed for determining and comparing expression of proteins, it was found that out of 2967 protein points, 89 points in resistant *L. major* had altered expression compared with sensitive *L. major*; 60 proteins had increased and 29 proteins had diminished protein expression. Also, they found that 11 protein points which did not exist in the sensitive *L. major* were expressed by resistance *L. major*. These changes of expression may be one of the major causes of resistance in *L. major* (31).

Conclusion

Considering the increased rate of drug resistance cases and numerous reports in this regard in different endemic regions of Iran, complete or hybrid treatment should always be taken into consideration by the treatment team in both private and governmental healthcare centers in order to tackle the acquired resistance of the parasite and relapse of disease. On the other hand, researchers should seek to develop new drugs with high efficacy in order to combat problems such as incomplete treatment and the presence of resistant strains in the indigent population. There are several reports on different aspects of leishmaniasis in the country. These reports will provide a guideline for disease control (32-66).

Acknowledgements

The authors highly appreciate the support and cooperation of all colleagues, students, and patients referred to the Medical Parasitology and Mycology Department, School of Public Health, Tehran University of Medical Sciences.

The authors declare that there is no conflict of interests.

References

- 1. World Health Organization (2014) Manual for case management of cutaneous leishmaniasis in the WHO Eastern Mediterranean Region. WHO Regional Publications, Eastern Mediterranean Series (35). Available at: https://apps.who.int/iris/handle/10665/12 0002
- 2. World Health Organization (2002) An increasing risk factor for leishmaniasis (Weekly Epidemiological Record. WHO Press. 77(44): 365–370.
- Nadim A, Javadian E, Mohebali M, Zamen Moemeni A (2008) *Leishmania* and Leishmaniasis in Iran, Second edition. Academic Publication Center, Tehran (In Persian).
- Ferreira FM, Castro RA, Batista MA, Rossi FMO, Silveria-Lemos D, Frezard F, A.L.Moura S, Rezende SA (2014) Association of water extract of green propolis and liposomal meglumine antimoniate in the treatment of experimental visceral leishmaniasis. Parasitol Res. 113 (2): 533–543.
- 5. Treatment and medical education of Iran (2002–2003) Diseases Management Cen-

ter, Ministry of Health, Treatment and Medical Education. Annual Communicable Diseases Report. Ministry of Health, Treatment and Medical Education Press, Iran.

- Firdous R, Yasunzai M, Ranja K (2009) Efficacy of Glucantime® in the treatment of old world cutaneous leishmaniasis. Int J Dermatol. 48(7): 758–762.
- Soleimanifard S, Arjmand R, Saberi S, Salehi M, Hejazi SH (2017) Treatment outcome of the drug-resistant zoonotic cutaneous leishmaniasis by Glucantime®. Adv Biomed Res. 6: 17.
- Croft SL (2001) Monitoring drug resistance in leishmaniasis. Trop Med Int Health. 6 (11): 899–905.
- Croft SL, Sundar S, Fairlamb AH (2006) Drug resistance in leishmaniasis. Clin Microbiol Rev. 19(1): 111–126.
- Hadighi R, Mohebali M, Boucher P, Hajjaran H, Khamesipour A, Ouellette M (2006) Unresponsiveness to Glucantime® treatment in Iranian Cutaneous leishmaniasis due to drug resistant *Leishmania tropica* parasites. PLoS Med. 3(5): e162.
- Rodrigues AM, Hueb M, Santos TA, Fernandes CJ (2006) Factors associated with treatment failure of cutaneous leishmaniasis with meglumine antimoniate. Rev Soc Bras Med Trop. 39(2): 139–145.
- 12. Samady JA, Janniger CK, Schwartz RA (1996) Cutaneous and mucocutaneous leishmaniasis. Cutis. 57: 13–20.
- Mohammadzadeh M, Behnaz F, Golshan Z (2013) Efficacy of Glucantime® for treatment of cutaneous leishmaniasis in central Iran. J Infect Public Health. 6(2): 120–124.
- 14. Hajjaran H, Mohebali H, Teimouri A, Oshaghi MA, Mirjalali H, Kazemi-Rad E, Shiee MR, Naddaf SR (2014) Identification and phylogenetic relationship of Iranian strains of various and visceral cases of leishmaniasis based on N-acetylglucosamin-1-phosphate transferase gene.

Infect Genet Evol. 26: 203–212.

- 15. Shirzadi MR (2012) Handbook of Cutaneous Leishmaniasis Care in Iran. Razi Nahan Publication. (In Persian).
- Magill AJ (2005) cutaneous leishmaniasis in the returning traveler. Infect Dis Clin North Am. 19(1): 241–266.
- 17. Holakouie-Naieni K, Mostafavi E, Darvishi Boloorani A, Mohebali M, Pakzad R (2017) Reprint of "Spatial modeling of cutaneous leishmaniasis in Iran from 1983 to 2013". Acta Trop. 165: 90–95.
- Killick-Kendrick R (1999) the biology and control of Phlebotominae sand flies. Clin Dermatol.17(3): 279–289.
- Grogl M, Thomason TN, Franke ED (1992) Drug resistance in leishmaniasis: its implication in systemic chemotherapy of cutaneous and mucocutaneus diseases. Am J Trop Med Hyg. 47(1): 117–260.
- 20. Lira R, Sundar S, Makharia A, Kenney R, Gam A, Saraiva E, Sacks D (1999) Evidence that the high incidence of treatment failures in Indian kala-azar is due to the emergence of antimony-resistant strains of *Leishmania donovani*. J Infect Dis. 180(2): 546–547.
- 21. Gramiccia M, Gradoni L, Orsini S (1992) Decreased sensitivity to meglumine antimoniate (Glucantime®) of *Leishmania infantum* isolated from Dogs after several courses of drug treatment. Ann Trop Med Parasit. 86(6): 613–620.
- Rojas R, Valderrama L, Valderrama M, Varona MX, Ouellette M, Saravia NG (2006) Resistance to antimony and treatment failure in human *Leishmania viannia* infection. J Infect Dis. 193(10): 1375–1383.
- 23. Walker J, Gongora R, Vasquez JJ, Drummelsmith J, Burchmore R, Roy G, Ouellette M, Gomez MA (2012) Discovery of factors linked to antimony resistance in *Leishmania panamensis* through differential proteome analysis. Mol Biochem Parasitol. 183(2): 166–176.

- 24. Khadem Erfan MB, Mohebali M, Kazemi-Rad E, Hajjaran H, Edrissian GH, Mamishi S, Saffari M, Raoofian R, Heidari M (2013) Downregulation of Calcineurin gene is associated with Glucantime[®] resiatance in *Leishmania infantum*. Iran J Parasitol. 8(3): 359–366.
- Ouellette M, Drummelsmith J, Papadopoulou B (2004) Leishmaniasis: drugs in the clinic, resistance and new developments. Drug Resist Updat. 7(4-5): 257–266.
- 26. Ghobakhloo N, Motazedian MH, Fardaei M (2016) Expression analysis of multiple genes may involve in antimony resistance among *Lishmania major* clinical isolates from Fars Province, central Iran. Iran J Parasitol. 11(2): 168–176.
- 27. Do Monte-Neto RL, Coelho AC, Raymond F, Legare D, Corbeil J, Melo MN, Frezard F, Ouellette M (2011) Gene expression profiling and molecular characterization of antimony resistance in *Leishmania amazonensis*. PLoS Negl Trop Dis. 5(5): e1167.
- Biyani N, Singh AK, Mandal S, Chawla B, Madhubala R (2011) Differential expression of proteins in antimony-susceptible and resistant isolates of *Leishmania donovani*. Mol Biochem Parasitol. 179(2): 91–99.
- 29. Kazemi-Rad E, Mohebali M, Khadem-Erfan MB, Hajjaran H, Hadighi R, Khamesipour A, Rezaie S, Saffari M, Raoofian R, Heidari M (2013) Overexpression of Ubiquitin and amino acid permease genes in association with antimony resistance in *Leishmania tropica* field isolates. Korean J Parasitol. 51(4): 413–419.
- 30. Kazemi-Rad E, Mohebali M, Khadem-Erfan MB, Saffari M, Raoofian R, Hajjaran H, Hadighi R, Khamesipour A, Rezaie S, Abedkhojasteh H, Heidari M (2013) Identification of antimony resistance markers in *Leishmania tropica* field isolates through cDNA–AFLP approach. Exper Parasitol. 135(2): 344–349.

- 31. Zarean M, Maraghi S, Hajjaran H, Mohebali M, Feiz-Hadad MH, Assaehzadegan MA (2015) Comparison of proteome profiling of two sensitive and resistant field iranian isolates of *Leishmania major* to Glucantime[®] by 2- dimensional electrophoresis. Iran J Parasitol. 10(1): 19–29.
- 32. Rassi Y, Jalali M, Vatandoost H (2000) Susceptibility status of *Ph. papatasi* to DDT in Arsanjan County in Fras Province, Iran. Iran J Public Health. 29(1–4): 21–23.
- 33. Rassi Y, Javadian E, Jalali M, Motazedian MH, Vatandoost H (2004) Investigation on zoonotic cutaneous leishmaniasis, southern Iran. Iran J Public Health. 33 (1): 31–35.
- 34. Rassi Y, Javadian E, Amin M, Rafizadeh S, Vatandoost H, Motazedian H (2006) *Meriones libycus*, the principal reservoir of zoonotic cutaneous leishmaniasis in southern Iran. East Mediterr Health J. 12(3–4): 474–477.
- 35. Yaghoobi-Ershadi MR, Akhavan AA, Jahanifard E, Vatandoost H, Amin Gh, Moosavi L, Zahraei Ramazani AR, Abdoli H, Arandian MH (2006) Repellency effect of Myrtle essential oil and DEET against *Phlebotomus paptasi Scopoli*, the main vector of zoonotic cutaneous leishmaniasis under laboratory conditions. Iran J Public Health. 35(3): 7–13.
- 36. Moosa-Kazemi SH, Shayeghi M, Abai MR, Vatandoost H, Sadeghi MT, Javadian E, Motabar M, Hosseini MR, Abtahi M (2009) High performance thin layer chromatography analysis of deltamethrin residue on the impregnated bed nets during a Leishmaniasis control program in Iran. Iran J Arthropod Borne Dis. 3(1): 1–7.
- 37. Oshaghi MA, Ravasan NM, Javadian E, Rassi Y, Sadraei J, Enayati AA, Vatandoost H, Zare Z, Emami SN (2009) Application of predictive degree day model for field development of sandfly vectors of visceral leishmaniasis in northwest of

Iran. J Vector Borne Dis. 46 (4): 247–55.

- 38. Aghaei Afshar A, Rassi Y, Sharifi I, Abai MR, Oshaghi MA, Yaghoobi-Ershadi MR, Vatandoost H (2011) Susceptibility status of *Phlebotomus papatasi* and *P. sergenti* (Diptera: Psychodidae) to DDT and Deltamethrin in a focus of Cutaneous Leishmaniasis after earthquake strike in Bam, Iran. Iran J Arthropod Borne Dis. 5(2): 32–41.
- 39. Saeidi Z, Vatandoost H, Akhavan AA, Yaghoobi-Ershadi MR, Rassi Y, Sheikh Z, Arandian MH, Jafari R, Sanei-Dehkordi AR (2012) Baseline susceptibility of a wild strain of *Phlebotomus papatasi* (Diptera: Psychodidae) to DDT and pyrethroids in an endemic focus of zoonotic cutaneous leishmaniasis in Iran. Pest Manag Sci. 68(5): 669–675.
- 40. Veisi A, Vatandoost H, Yaghoobi-Ershadi MR, Arandian MH, Jafari R, Hosseini M, Abdoli H, Rassi Y, Heidari K, Sadjadi A, Fadaei R, Ramazanpour J, Aminian K, Shirzadi MR, Akhavan AA (2012) Comparative study on the effectiveness of Coumavec and zinc phosphide in controlling zoonotic cutaneous leishmaniasis in a hyperendemic focus in central Iran. J Arthropod Borne Dis. 6(1): 18–27.
- 41. Veysi A, Vatandoost H, Arandian MH, Jafari R, Yaghoobi-Ershadi MR, Rassi Y, Akhava AA (2013) Laboratory evaluation of a rodenticide-insecticide, Coumavec®, against *Rhombomys opimus*, the main reservoir host of zoonotic cutaneouse leishmaniasis in Iran. J Arthropod Borne Dis. 7(2): 188–193.
- 42. Saeidi Z, Vatandoost H, Akhavan AA, Yaghoobi-Ershadi MR, Rassi Y, Arandian MH, Jafari R (2013) Baseline insecticide susceptibility data of *Phlebotomus papatasi* in Iran. J Vector Borne Dis. 50: 57–61.
- 43. Aghai Afshar A, Vatandoost H, Sharifi I, Rassi Y, Abai MR, Oshaghi MA, Yaghoobi-Ershadi MR, Rafizadeh S

(2013) First determination of impact and outcome indicators following indoor residual spraying (IRS) with deltamethrin in a new focus of anthroponotic cutaneous leishmaniasis (ACL) in Iran. Asian Pac J Trop Dis. 3(1): 5–9.

- 44. Aghai Afshar A, Rassi Y, Sharifi I, Vatandoost H, Mollaie HR, Oshaghi MA, Abai MR, Rafizadeh S (2014) First report on natural *Leishmania* infection of *Phlebotomus sergenti* due *Leishmania tropica* by high resolution melting curve method in South-eastern Iran. Asian Pac J Trop Med. 7(2): 93–96.
- 45. Akhavan AA, Veisi A, Arandian MH, Vatandoost H, Yaghoobi-Ershadi MR, Hosseini M, Abdoli H, Heidari K, Sadjadi A, Fadaei R, Ramazanpour J, Aminian K, Shirzadi MR, Jafari R (2014) Field evaluation of phostoxin and zinc phosphide for the control of zoonotic cutaneous leishmaniasis in a hyperendemic area, central Iran. J Vector Borne Dis. 51(4): 307–312.
- 46. Jalilnavaz MR, Abai MR, Vatandoost H, Mohebali M, Akhavan AA, Zarei Z, Rafizadeh S, Bakhshi H, Rassi Y (2016) Application of flumethrin pour-on on reservoir dogs and its efficacy against sand flies in endemic focus of visceral leishmaniasis, Meshkinshahr, Iran. J Arthropod Borne Dis. 10(1): 78–86.
- 47. Hazratian T, Vatandoost H, Oshaghi MA, Yaghoobi-Ershadi MR, Fallah E, Rafizadeh S, Shirzadi MR, Shayeghi M, Akbarzadeh K, Rassi Y (2016) Diversity of sand flies (Diptera: Psychodidae) in endemic focus of visceral leishmaniasis in Azar shahr District, east Azarbaijan Province, North West of Iran. J Arthropod Borne Dis. 10(3): 328–334.
- 48. Sofizadeh A, Vatandoost H, Rassi Y, Hanafi-Bojd AA, Rafizade S (2016) Spatial Analyses of the relation between rodent's active burrows and incidence of zoonotic cutaneous leishmaniasis in Golestan Province, Northeastern of Iran. J Arthropod

Borne Dis. 10(4): 569-576.

- 49. Saghafipour A, Vatandoost H, Zahraei-Ramazani AR, Yaghoobi-Ershadi MR, Rassi Y, Shirzadi MR, Akhavan AA (2016) Spatial distribution of phlebotomine sand flies species (Diptera: Psychodidae) in Qom Province, central Iran. J Med Entomol. 54(1): 35–43.
- 50. Saghafipour A, Vatandoost H, Zahraei-Ramazani AR, Yaghoobi-Ershadi MR, Karami Jooshin M, Rassi Y, Shirzadi MR, Akhavan AA, Hanafi-Bojd AA (2017) Epidemiological Study on Cutaneous Leishmaniasis in an Endemic Area of Qom Province, Central Iran. J Arthropod Borne Dis. 11(3): 403–413.
- 51. Sofizadeh A, Rassi Y, Vatandoost H, Hanafi-Bojd AA, Mollalo A, Rafizadeh S, Akhavan AA (2017) Predicting the distribution of *Phlebotomus papatasi* (Diptera: Psychodidae), the primary vector of zoonotic cutaneous leishmaniasis, in Golestan Province of Iran using Ecological Niche Modeling: Comparison of MaxEnt and GARP Models. J Med Entomol. 54(2): 312–320.
- 52. Veysi A, Vatandoost H, Yaghoobi-Ershadi MR, Jafari R, Arandian MH, Hosseini M, Fadaei R, Ramazanpour J, Heidari K, Sadjadi A, Shirzadi MR, Akhavan AA (2016) Rodenticide comparative effect of klerat® and zinc phosphide for controlling zoonotic cutaneous leishmaniasis in central Iran. Iran J Parasitol. 11(4): 471–479.
- 53. Saghafipour A, Vatandoost H, Zahraei-Ramazani AR, Yaghoobi-Ershadi MR, Rassi Y, Jooshin M, Shirzadi MR, Akhavan AA (2017) Control of zoonotic cutaneous leishmaniasis vector, *Phlebotomus papatasi*, using attractive toxic sugar baits (ATSB). PLOs One. 12(4): e0173558.
- 54. Shirani-Bidabadi L, Zahraei-Ramazani AR, Yaghoobi-Ershadi MR, Rassi Y, Akhavan AA, Oshaghi MA, Enayati AA, Saeidi Z, Jafari R, Vatandoost H (2017) Assessing the insecticide susceptibility sta-

tus of field population of *Phlebotomus papatasi* (Diptera: Psychodidae) in a hyperendemic area of zoonotic cutaneous leishmaniasis in Esfahan Province, central Iran. Acta Trop. 176: 316–322.

- 55. Arzamani K, Vatandoost H, Rassi Y, Abai MR, Akhavan AA, Alavinia M, Akbarzadeh K, Mohebali M, Rfizadeh S (2017) Susceptibility status of wild population of *Phlebotomus sergenti* (Diptera: Psychodidae) to different imagicides in an endemic focus of cutaneous leishmaniasis in northeast of Iran. J Vector Borne Dis. 54(3): 282–286.
- 56. Moradiasl E, Rassi Y, Hanafi-Bojd AA, Vatandoost H, Saghafipour A, Adham D, Aabasgolizadeh N, Omidi Oskouei A, Sadeghi H (2018) The relationship between climatic factors and the prevalence of visceral leishmaniasis in Northwest of Iran. Intern J Pediatrics. 2(50): 7169– 7178.
- 57. Vatandoost H, Nejati J, Saghafipour A, Zahraei-Ramazani A (2018) Geographic and ecological features of phlebotomine sand flies (Diptera: Psychodidae) as leishmaniasis in Central Iran. J Parasit Dis. 42(1): 43–49.
- 58. Karimian F, Vatandoost H, Rassi Y, Maleki-Ravasan N, Choubdar N, Koosha M, Arzamani K, Moradi-Asl E, Veysi A, Alipour H, Shirani M, Oshaghi MA (2018) Wsp-based analysis of Wolbachia strains associated with *Phlebotomus papatasi* and *P. sergenti* (Diptera: Psychodidae) main cutaneous leishmaniasis vectors. Pathog Glob Health. 112(3): 152–160.
- 59. Arzamani K, Vatandoost H, Rassi Y, Akhavan AA, Abai MR, Alavinia M, Akbarzadeh K, Mohebali M, Rafizadeh S (2018) Richness and diversity of phlebotomine sand flies (Diptera: Psychodidae) in North Khorasan Province, northeast of Iran. J Arthropod Borne Dis. 12(3): 232–239
- 60. Karimian F, Vatandoost H, Rassi Y, Maleki-Ravasan N, Mohebali M, Shirazi MH,

Koosha M, Choubdar N, Oshaghi MA (2019) Aerobic midgut microbiota of sand fly vectors of zoonotic visceral leishmaniasis from northern Iran, a step toward finding potential paratransgenic candidates. Parasit Vectors. 12: 10.

- 61. Yaghoobi-Ershadi MR, Akhavan AA, Shirzadi MR, Rassi Y, Khamesipour A, Hanafi-Bojd AA, Vatandoost H (2019) Conducting international diploma course on leishmaniasis and its control in the Islamic Republic of Iran. J Arthropod Borne Dis. 13(3): 234–242.
- 62. Rassi Y, Moradi-Asl E, Vatandoost H, Abazari M, Saghafipour A (2020) Insecticide susceptibility status of wild population of *Phlebotomus kandelakii* and *Phlebotomus perfiliewi* transcaucasicus collected from visceral leishmaniasis endemic foci in northwestern Iran. J Arthropod Borne Dis. 14(3): 277–285.
- 63. Yousefi S, Zahraei-Ramazani AR, Rassi Y, Vatandoost H, Yaghoobi-Ershadi MR, Aflatoonian MR, Akhavan AA, Aghaei-Afshar A, Amin M, Paksa A (2020) Evaluation of different attractive traps for capturing sand flies (Diptera: Psychodidae) in an endemic area of Leishmaniasis, Southeast of Iran. J Arthropod Borne Dis. 14(2): 202–213.
- 64. Moradi-Asl E, Mohebali M, Rassi Y, Vatandoost H, Saghafipour A (2020) Environmental variables associated with distribution of canine visceral leishmaniasis in dogs in Ardabil Province, Northwestern Iran. Iran J Public Health. 49(6): 1033–1044.
- 65. Shirani-Bidabadi L, Zahraei-Ramazani AR, Yaghoobi-Ershadi MR, Akhavan AA, Oshaghi MA, Enayati AA, Rassi Y, Gholampour F, Shareghi N, Madreseh E, Vatandoost H (2020) Monitoring of Laboratory Reared of *Phlebotomus papatasi* (Diptera: Psychodidae), main vector of zoonotic cutaneous leishmaniasis to different imagicides in hyper endem-

ic areas, Esfahan Province, Iran. J Arthropod Borne Dis. 14(1): 116–125.

66. Mozaffari E, Vatandoost H, Rassi Y, Mohebali M, Akhavan AA, Moradi-Asl E, Zarei Z, Zahrai-Ramazani A, Ghorbani E (2020) Epidemiology of visceral leishmaniasis with emphasis on the dynamic activity of sand flies in an important endemic focus of disease in Northwestern Iran. J Arthropod Borne Dis. 14(1): 97–105.