## **Original Article**

# Chemical Composition and Repellent Activity of Achillea vermiculata and Satureja hortensis against Anopheles stephensi

Masoumeh Pirmohammadi<sup>1</sup>, \*Mansoureh Shayeghi<sup>1</sup>, \*Hassan Vatandoost<sup>1,2</sup>, \*Mohammad Reza Abaei<sup>1</sup>, Ali Mohammadi<sup>1</sup>, Akbar Bagheri<sup>1</sup>, Mehdi Khoobdel<sup>3</sup>, Hasan Bakhshi<sup>1</sup>, Maryam Pirmohammadi<sup>4</sup>, Maryam Tavassoli<sup>1</sup>

<sup>1</sup>Department of Medical Entomology and Vector Control, School of Public Health, Tehran University of Medical Sciences, Iran

<sup>2</sup>Instituet for Environmental Research, Tehran University of Medical Sciences, Tehran, Iran <sup>3</sup>Health Research Center, Baqiyatallah University of Medical Sciences, Tehran, Iran <sup>4</sup>Kashan University, Kashan, Iran

(Received 3 Aug 2014; accepted 17 Jan 2015)

#### Abstract

**Background**: One of the best ways to control the malaria disease and to be protected human against *Anopheles* mosquito biting is the use of repellents. Throughout repellents, herbal ones may be an appropriate and safe source for protection.

**Methods**: Chemical constituents of *Achillea vermiculata* and *Satoreja hortensis* were determined by using gas chromatography-mass spectrometry. Efficacy and the protection time of these plants were assessed on *Anopheles stephensi* under the laboratory condition.

**Results**: The mean assessed protection time and efficacy for *A. vermiculata* was 2.16 and 3.16 hours respectively and the obtained  $ED_{50}$  and  $ED_{90}$  for this plant was 5.67 and 63 µl/cm<sup>2</sup> respectively. The figured for *S. hortensis* was 4.16 and 5 hours respectively. ED50 and ED90 for this plant were 5.63 and 45.75µl/cm<sup>2</sup> respectively.

**Conclusion**: Results of investigation showed that *S. hortensis* plant has an acceptable protection time, therefore, this plant could be considered as a good herbal repellent against anopheles mosquitoes.

Keywords: Achillea vermiculata, Satureja hortensis, Anopheles stephensi, Repellency, Protection time

### Introduction

Mosquito borne diseases affect human societies by reduction in labor productivity especially in tropical and subtropical countries. The important point is this fact that all countries all over the world have a problem with insect borne diseases (Govindarajan et al. 2011).

Anopheles mosquitoes are bloodsucking insects, responsible for transmission of malaria, filariasis and arboviruses (Service 1980). There are 33 currently recognized Anopheles species including sibling, biological forms and zygotypes, seven of these species have an important role in malaria transmission in Iran. Among these species, *An. stephensi* is considered as a primary vector of malaria in southern parts of the country (Sedaghat et al. 2003, Vatandoost et al. 2004, Sedaghat et al. 2005). According to the WHO report, a total of 627000 people die due to malaria (WHO 2013).

Malaria is caused by *Plasmodium* parasite. Malaria is one of the most important diseases which the parasite is transmitted by female *Anopheles* genus (WHO 2010). There are several methods for malaria vector control.

201

<sup>\*</sup>**Corresponding authors:** Dr Mansoureh Shayeghi, E-mail: mansorehshayeghi@yahoo.com, Dr Hassan Vatandoost, E-mail: hvatandoost1@yahoo.com, Dr Mohammad Reza Abaei, E-mail: abaimr@tums.ac.ir

Synthetic insecticides which are generally used have side-effects on human, animal health, and the environment. The side-effects of synthetic organophosphorus compounds on fish and other organisms in the environment are being increasingly reported. A lot of attention is being paid to natural products in vector control as they are environmentally safe, degradable and target-specific. Recent studies have demonstrated that use of repellents is one of the effective ways to control the disease and to avoid *Anopheles* bites (Vatandoost et al. 2008).

DEET is slightly yellow oil. It is the most common active ingredient in insect repellents. It is intended to be applied to the skin or to clothing, and provides protection against mosquitoes and many other biting insects. DEET was developed during World War II. The findings bring evidence that, DEET has side effects, so it has proposed to use alternative repellents for protection (Karunamoorthi et al. 2010). Some prefer to use natural insect repellent products. Repellents of plant origin do not pose hazards of toxicity to human and domestic animals and are easily biodegradable. Natural products are safe for human when compared to that of synthetic compounds (Fradin 1998). The effect of some plant origin essential oils have been tested in Iran (Oshaghi et al. 2003, Yaghoobi-Ershadi et al. 2006, Vatandoost and Hanafi-bojd 2008, Tavassoli et al. 2011, Mozaffari et al. 2014).

Achillea vermiculata is a flowering plant in the family Asteraceae with a height of 10– 30 cm. It is native to temperate regions of the Northern Hemisphere in Asia, Europe, and North America. It is an erect herbaceous perennial plant. The leaves have varying degrees of hairiness. The leaves are almost feathery (Mozaffarian 2012).

*Satureja hortensis* has lilac tubular flowers. It grows to around 30 to 60 cm in height and has very slender, bronze-green leaves. This plant belongs to order: Lamiales and family: Lamiaceae. It is used in traditional medicine as a botanical treatment (Mozaffarian 2012).

This study was conducted to evaluate the repellent properties of two plants *A. vermiculata* and *S.* essential oil against *An. stephensi* in laboratory condition on animal model and also to determine chemical compositions in their essential oils.

## **Materials and Methods**

#### **Mosquitoes rearing**

Established colony of susceptible strain of An. stephensi obtained from the Insectary of School of Public Health, Tehran University of Medical Sciences, Tehran, Iran. Mosquitoes were reared and maintained at 28±2 °C and 65±5% relative humidity (RH) under a 16:8 (L: D) photoperiod. Larvae were fed on a diet of fish food and water lettuce. The adults were maintained in screen cages and fed with 10% aqueous sucrose solution as a source of energy and guinea pigs as bloodfeeding female mosquitoes for maturing the eggs. Starved 5 to 8 days old females were used for the repellency tests. The sucrose solution was picking up from the cage, 12 hour before starting the experiments.

# Collection, identification and extraction of plants

Fresh flowers and leaves flowers of *A. vermiculata* and *S. hortensis* were collected from Armand and Sheyda district which are located in Chaharmahal and Bakhtiari Province in south-west of Iran in June 2013 (Fig.1,2). They were rapidly transported to the School of Public Health, Tehran University of Medical Sciences. *Achillea vermiculata* was collected from natural habitat in Armand district at coordinate 31° 39.428 E 50° 46.659 N, 1136 meters above sea level. *Satureja hortensis* was collected from natural habitat in Sheyda district in Ben at coordinate 32° 37.206 E 50° 42.434 N, 2219 meters above sea level.

The plant A. vermiculata was identified by experts in Department of Plant Sciences, Tehran University. Also S. hortensis was identified by experts in Ecotoxicology and herbarium laboratory, School of Public Health, Tehran University of Medical Sciences. The flowers and leaves of A. vermiculata and S. hortensis were dried at room temperature under good ventilation. About 525 gr of dried A. vermiculata and 420 gr of dried S. hortensis were chopped into small pieces using a knife mill. The essential oil was extracted from the plants using a Clevengertype water steam distillation apparatus. It took about 4 hours for extraction of the essential oils. For extraction of essential oil of each plant we used Sulphate Sodium Anhydr. The distilled essential oils were stored in a refrigerator at 4 °C until being used in the experiments. The composition of the volatile constituents was established by gas chromatography-mass spectrometry.

#### **Test method**

All series of the experiments of effective dose and protection time were carried out in laboratory condition. We tested these plants on white rabbits. The white rabbits (O. cuniculus) (laboratory reared albino male aged 6-8 months) were used to determine both protection time and effective dosage in insectary at the School of Public Health. In this investigation, the 50% concentration of essential oils was used for protection time test. For this purpose essential oils were diluted by absolute ethanol. Then for estimating the protection time of plants essential oils against An. stephensi the back of male rabbits by  $52 \text{cm}^2$  were shaved in the next stage, 80µl of 50% essential oils of two plants by sampler on the shaved back of male rabbits were applied. After 5 minutes the rabbits were placed in a box which is designed for the test. Then it is placed in box containing the rabbit in a cage at dimension of 53×53×53 cm containing 150 starved 5-8

days mosquitoes. After 3 minutes of biting and probing records, we brought out the cage and we tested again 30 minutes later. These tests continued until two successive bites. This time is called protection time. We continued these tests until 10 bites. This time is called failure time (Pitasawat et al. 2003).

The procedure for determination of effective dosages of the repellents was adopted by the standard method of American Society for Testing and Material (ASTM 2000).

The testing kit was made of plexiglas cube at dimension of  $4 \times 5 \times 18$  cm having four rectangular holes  $4 \times 3$  cm. Before starting the test for determination of effective dosage, the abdomen skins of rabbits were cleaned with alcohol and the kit was fixed on the abdomen.

Each of 4 adjacent cells of kit was provided with 5 female 5-8 days mosquitoes that randomly selected from a cage containing 150 starved mosquitoes. Circles were drawn on the rabbit's skin. The drawn circles on the abdomen skin's of hold rabbit were treated with 50ul of essential oil diluted with absolute ethanol at 6.6, 13.2, 26.4 and 52.8µl for A. vermiculata and 3.3, 6.6, 13.2, 26.4 and 52.8µl for S. hortensis microliter with 4 repetitions. The same dilutions were applied on 3 holes because of prevention of contamination as well as the absolute ethanol was applied in remaining control circle. We used 5 mosquitoes for each hole. The treated circles were allowed to dry, and then test apparatus containing starved mosquitoes were fixed on the treated skin. The counts of probing and biting were recorded for 5 minutes. After each test, the mosquitoes were transferred to netted cups and the mortality of mosquitoes was recorded after 24 hours. The ED<sub>50</sub> and ED<sub>90</sub> values and regression parameters were analyzed using probit 79 programs and the regression lines were plotted in Microsoft Excel 2007.

#### Plants essential oils analysis

Chemical composition of A. vermiculata

J Arthropod-Borne Dis, June 2016, 10(2): 201–210

and *S. hortensis* was analyzed using an Agilent 7890–5975 gas chromatographymass spectrometer. With a HP- 5MS (5% Phenyl Methyl Silox) capillary column ( $30m\times0.25mm$ , film thickness  $0.25\mu m$ ), split ratio, 1: 1, and using a flame ionization detector. The GC was programmed at 50 °C for 0.5 min and then increased at 5 °C/min to 280 °C, and finally held with an isothermal for 3min.

The injector temperature was 280 °C. The flow rate of the carrier gas was 1ml/min. The identification of compounds was performed by comparing their retention times and mass spectra with mass spectra from Wiley library.

## **Results**

### **Essential oil volumes**

By the use of Clevenger-type water steam distillation, about  $1849\mu$ l of essential oil of 525gr of dried *A. vermiculata* flowers was extracted. Also about 4480µl of essential oil of 420gr of dried *S. hortensis* leaves extracted.

#### **GC-mass analysis**

One microliter of each essential oil was injected to GC-mass. A total of 40 compounds were identified in flowers of *A. vermiculata*. (E)- -damascenone with 27.4, (E)-2-hexenal with 8, eugenol with 6 and geranyl acetone with 6 percent were the major components (Table 1). We just found a repellent component "camphene" of all identified components by researchers until now in this plant with 0.7%.

Also we identified 23 components in the leaves of *S. hortensis*. B-oplopenone with 57, trans-carvone oxide with 15.13 and thymol methyl ether with 13 percent were the major components (Table 2).

#### **Protection time**

The protection time of *A. vermiculata* essential oil against *An. stephensi* on animal

subject provided 2.0-2.5 hours range with a mean of 2.16 hours protection and a failure time of 3-3.5 hours range with a mean of 3.16 hours.

Also the protection time of *S. hortensis* essential oil provided 4–4.5 hours range with a mean of 4.16 hours protection and a failure time of about 5 hours (Table 3).

Significant differences of protection time and failure time between *A. vermiculata* and *S. hortensis* repellents were observed by ANOVA (Games-Howel), P< 0.05.

#### **Effective dose**

The ED<sub>50</sub> and ED<sub>90</sub> values of *A*. *vermiculata* essential oil were 5.67 and 63  $\mu$ l/cm<sup>2</sup> with confidence interval ranged, 2.25-8.68 and 38.21–198.07  $\mu$ l/cm<sup>2</sup> respectively (Table 4).

The ED<sub>50</sub> and ED<sub>90</sub> values of *S. hortensis* essential oil were 5.63 and 45.73  $\mu$ l/cm<sup>2</sup> with confidence interval ranged, 3.83-7.43 and 30.92–86.55  $\mu$ l/cm<sup>2</sup> respectively (Table 4).

We did not observe any significant differences between  $ED_{50}$  and  $ED_{90}$  of *S. hortensis* and *A. vermiculata* by T-test and P> 0.001 analysis.



**Fig. 1.** The plant *Achillea vermiculata* in its natural habitat, Armand district in Chaharmahal and Bakhtiari Province, south-western of Iran (original)

NO	Compound	Composi-	RI	
	-			
1	isovaleric acid	0.2	833	
2	(E)-2-hexenal	8	854	
3	Tricyclene	0.1	921	
4	Limonene	0.3	1029	
5	Methybenzoate	0.5	1093	
6	Linalool	3.1	1108	
7	-campholenal	2.8	1131	
8	Camphene	0.7	1156	
9	Pinocarvone	0.7	1164	
10	cis-piperitol	0.7	1194	
11	Verbenone	0.8	1208	
12	trans-carveol	4	1222	
13	cis-carveol	3.7	1233	
14	Geraniol	1	1258	
15	2E,4E-decadienal	3.1	1313	
16	Eugenol	6	1359	
17	(E)damascenone	27.4	1382	
18	cis-a-bergarnotene	5.6	1416	
19	trans-a-bergamotene	3	1433	
20	geranyl acetone	6	1452	
21	allo-aromadendrene	0.8	1462	
22	gamma-gurjunene	0.3	1473	
23	a-muurolene	0.6	1499	
24	a-cadinene	1.2	1537	
25	Spathulenol	0.7	1579	
26	b-oplopenone	0.4	1601	
27	humulene epoxide II	0.3	1609	
28	silphiperfol-6-en-5-one	0.6	1623	
29	1-epi-a-eudesmol	2	1660	
30	8-cedren-13-ol	0.3	1694	
31	Xanthorrizol	0.3	1751	
32	8-a-acetoxyelemol	2	1788	
33	Nootkatone	0.4	1802	
34	Flourensiadiol	1.8	1864	
35	Hexadecanol	1.7	1881	
36	methyl hexadecanoate	1.9	1911	
37	methyl hexadecanoate	1.8	1925	
38	methyl hexadecanoate	2.2	1941	
39	methyl hexadecanoate	2.6	1957	
40	Heneicosane	0.25	2057	
-		99.55		

 Table 1. Chemical constituents of flower essential oil from Achillea vermiculata

Table 2. Chemical	l constituents of leaf essential oil
from	Satureja hortensis

NO	Compound	Composition%	RI
1	isovaleric acid	0.06	831
2	(Z)-3-hexenol	2.1	856
3	trans-sabinene hy-	0.3	1095
	drate		
4	Linalool	1	1109
5	-campholenal	0.6	1132
6	Borneol	1	1171
7	trans-carveol	2	1221
8	thymol methyl ether	13	1237
9	trans-carvone oxide	15.1	1277
10	Undecanal	0.4	1306
11	-caroyophyllene	3	1424
12	b-oplopenone	57	1600
13	10-epi-gamma-	0.3	1620
	eudesmol		
14	Hinesol	0.6	1637
15	8-cedren-13-ol	1	1704
16	Oplopanone	0.2	1726
17	8-a-acetoxyelemol	0.3	1781
18	Flourensiadiol	1.04	1867
19	methyl	0.12	1910
	hexadecanoate		
20	methyl	0.2	1923
	hexadecanoate		
21	methyl	0.2	1935
	hexadecanoate		
22	Heneicosane	0.1	2097
23	n-docosane	0.2	2213
		99.82	



**Fig. 2.** The plant *Satureja hortensis* in its natural habitat, Sheyda district in Chaharmahal and Bakhtiari Province, south-western of Iran (original)

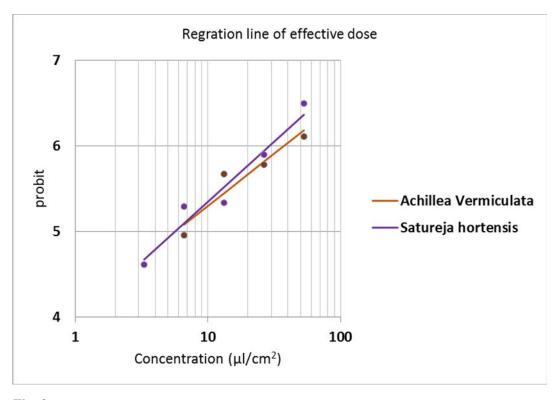


Fig. 3. Dose-response lines for two botanical repellents against Anopheles stephensi on animal model

**Table 3.** Protection time and failure time of Achillea vermiculata and Satureja hortensis against An. stephensi on animal subject in laboratory condition

Species (plants)	District	Protection time (hour)		Failure time (hour)	
		Range	Mean	Range	Mean
A. vermiculata	Lordegan	2-2.5	2.16	3–3.5	3.16
S. hortensis	Ben	4-4.5	4.16	5–5	5

Table 4. Effective dose of Achillea vermiculata and Satureja hortensis essential oils against Anopheles stephensi on				
animal subject in laboratory condition				

plants	a	b ± SE	ED50 (mg/cm <sup>2</sup> ) ± 95% C.L.	ED90 (mg/cm <sup>2</sup> ) ± 95% C.L.	2 (heterogeneity)	2 table (df)	p-Value
<i>A</i> .	-0.092	1.22±0.27	5.67	63	2.35	2	0.01
vermiculata			(2.25 - 8.68)	(38.21–198.07)			
S. hortensis	-1.05	1.4±0.20	5.63	45.75	3.13	3	0.01
			(3.83–7.43)	(30.92-86.55)			

## Discussion

Application of larvicides and repellents are generally accepted as they play an important role in control of the mosquitoes. The use of botanical essential oils as repellents against vectors of malaria disease including *An.* gambiae and *A. stephensi* has been tested successfully (Seyoum et al. 2002). In this study the components of *A. vermiculata* essential

oil were identified. (E)- -damascenone with 27.4, (E)-2-hexenal with 8, eugenol with 6 and geranyl acetone with 6 percent were the major components. Totally 40 components were identified from this plant. In an investigation by Ahmadi et al. (2011), on *A. santolina*, 29 components were identified from this plant which the major components were Camphor, Alpha-pinene, Camphene and 1,8 Cineole with 26.27, 10.14, 9.09 and 8.26 percent respectively. We found two same components "Camphene" and "Linalool" of *A. vermiculata* with *A. santolina*.

Also we identified 23 components in the leaves of S. hortensis. B-oplopenone with 57, trans-carvone oxide with 15.13 and thymol methyl ether with 13 percent was the major components. In an investigation by Kamkar et al. (2013) (32 components were identified and reported from this plant which the major components belonged to - terpinene with 24.72%, thymol with 29.1% and carvacrol with 26.6%. In this investigation, we identified some components which had been identified in Kamkar et al. (2013) study. These components are: borneol, linalool and thymol. Also cis-sabinene hydrat was identified in Kamkar investigation, but our investigation revealed the presence of trans-sabinene hydrate. In another investigation by Tajalli et al. (2012) on this plant, the major components were thymol, carvacrol, gamma-terpinene with 48.67, 8.96, 9.16 and 9.16 percent respectively. In our investigation, we found thymol and linalool which had been identified in Tajalli et al. (2012) study too. Our investigation revealed that A. vermiculata essential oil can have 2.16 hours protection time and also its Failure time is 3.16 hours. Compared to A. vermiculata, S. hortensis could have 4.16 hours protection time and a Failure time of 5 hours which is about 2 times as much as A. vermiculata's protection time and Failure time. Also we revealed an ED50 and ED90 effective dose of 5.67 and 63  $\mu$ l/cm<sup>2</sup> respectively, while for S. hortensis they were 5.63

and 45.75  $\mu$ l/cm<sup>2</sup> respectively.

The repellency effect of essential oils of some plants has been studied in Iran. On a laboratory trial by Vatandoost et al. (2008), the repellency of neem tree's essential oil against *An. Stephensi* in animal subject was determined. The ED<sub>50</sub> and ED<sub>90</sub> values of neem tree's essential oils were calculated 0.159 and 1.388 mg /cm<sup>2</sup> respectively. Also the protection time and effective dose of this plant calculated 31 minutes and 65 minutes respectively.

The repellency effect of essential oils of both *Myrtus communis* and *Calendula officinalis* had been reported against *An. stephensi* on human subject and the effective dose of these plants was 0.11 and 0.6 mg/ cm<sup>2</sup> respectively. Also the protection time and Failure time for *M. communi* were 4.36 and 4.4 hours respectively. The protection time and Failure time for *C. officinalis* were 2.15 and 3.30 hours respectively (Tavassoli et al. 2011).

In an investigation, the mean protection time of 50% essential oil of *Cionura erecta* (*L*) provided 2.28 hours protection against *An. stephensi*. The figures for  $ED_{50}$  and  $ED_{90}$  values were 10.12 and 23.01ppm respectively (Mozaffari et al. 2014).

We estimate that the most protection time of mentioned investigations and our investigation, belongs to *M. communi* and *S. hortensis* with a protection time of 4.36 and 4.16 hours and failure time of 4.4 and 5 hours respectively. The weakest protection time and failure time belongs to neem plant by 31 minutes and 65 minutes respectively.

There have been so many investigations about repellency effects of plants against mosquitoes all over the world until now (Moore et al. 2002, Rajkumar and Jeanesabn 2007, Mullai et al. 2008, Karunamoorthei et al. 2010, Shahi et al. 2010).

Finally we recommend *S. hortensis* as a candidate for prodution of insect repellents because of its high protection against mos-

quitoes and also medicinal properties without any side effects. Although this plant did not show any significant differences of effective dose rate with *A. vermiculata*. We recommend doing more investigations on this plant.

## Conclusion

According to the results it could be concluded that the plant is appropriate for the repellent formulation for mosquito control, although the field trail should be conducted in a malarious areas.

## Acknowledgments

This study was funded and supported by Tehran University of Medical Sciences (TUMS), we are thankful to the staff of Insectary of Culicidae, Department of Medical Entomology and Vector Control, School of Public Health, Tehran University of Medical Sciences for the mass production of the mosquitoes for this study. We are thankful to Dr Salmaki, assistant professor from Department of Plant Sciences, Tehran University for detection of A. vermiculata. We are also thankful to staff of Ecotoxicology Laboratory and Central Laboratory of School of Public Health for identification of plants and analyzing the contents. The authors declare that there is no conflict of interests.

## References

- Ahmadi Z, Sattari M, Tabaraee B, Bigdeli M (2011) Identification of the constituents of *Achillea santolina* essential oil and evaluation of the anti-microbial effects of its extract and essential oil. Arak Medical University Journal. 14(56): 1– 10. (Persian)
- Fradin MS (1998) Mosquitoes and mosquito repellents. Ann Intern Med. 128: 931– 940.

- Govindarajan M, Mathivanan T, Elumalai K, Krishnappa K, Anandan A (2011) Mosquito larvicidal, ovicidal, and repellent properties of botanical extracts against *Anopheles stephensi, Aedes aegypti*, and *Culex quinquefasciatus* (Diptera: Culicidae). Parasitol Res. 109(2): 353–367.
- Kamkar A, Tooryan F, Akhondzadeh Basti A, MisaghiA, Shariatifar N (2013) Chemical composition of summer savory (Satureja hortensis L.) essential oil and comparison of antioxidant activity with aqueous and alcoholic extracts. J Vet Res. 68(2): 183–190. (Persian).
- Karunamoorthi K, Ilango K, Murugan K (2010) Laboratory evaluation of traditionally used plant-based insect repellent against the malaria vector *Anopheles arabiensis* Patton (Diptera: Culicidae). Parasitol Res. 106(5): 1217–1223.
- Moore SJ, Lenglet A, Hill N (2002) Field evaluation of three plant-based insect repellents against malaria vectors in Vaca Diez Province, the Bolivian Amazon. J Am Mosq Control Assoc. 18 (2): 107–110.
- Mozaffari E, Abai MR, Khanavi M, Vatandoost H, Sedaghat MM, Sanei-Dehkordi A, Moridnia A, Saber-Navaei M, Rafi F (2013) Chemical composition, larvicidal and repellent properties of Cionura erecta (L.) griseb. against malaria vector, *Anopheles stephensi* liston (Diptera: Culicidae) under laboratory conditions. J Arthropod-Borne Dis. 8(2): 147–155.
- Mozaffarian VA (2012) Recognition of medicinal and aromatic plants in Iran. 78: 145. (Persian)
- Mullai K, Jebanesan A, Pushpanathan T (2008) Mosquitocidal and repellent activity of the leaf extract of *Citrullus vulgaris* (cucurbitaceae) against the malarial vector, *Anopheles stephensi* liston (Diptera: Culicidae). Eur Rev Med Pharmacol Sci. 12(1): 1–7.

- Oshaghi M, Ghalandari R, Vatandoost H, Shayeghi M, Kamali-Nejad M, Tourabi-Khaledi H (2003) Repellent effect of extracts and essential oil of Citrus limon (Rutaceae) and *Melissa officinalis* (Labiatae) against main malaria vector, *Anopheles stephensi* (Diptera: Culicidae) in Iran. Iran J Public Health. 32(4): 47–52.
- Pitasawat B, Champakaew D, Choochote W, Jitpakdi A, Chaithonga U, Kanjanapothi D, Rattanachanpichai E, Tippawangkosol P, Riyong D, Tuetun B, Chaiyasit D (2007) Aromatic plant-derived essential oil: An alternative larvicide for mosquito control. Fitoterapia. 78(3): 205– 210.
- Rajkumar S, Jebanesan A (2007) Repellent activity of selected plant essential oils against the malarial fever mosquito *Anopheles stephensi*. Trop Biomed. 24 (2): 71–75.
- Shahi M, Hanafi-Bojd AA, Iranshahi M, Vatandoost H, Hanafi-Bojd MY(2010) Larvicidal efficacy of latex and extract of *Calotropis procera* (Gentianales: Asclepiadaceae) against *Culex quinquefasciatus* and *Anopheles stephensi* (Diptera: Culicidae). J Vector Borne Dis. 47(3): 185–188.
- Sedaghat MM, Harbach RE (2005) An annotated checklist of the *Anopheles* mosquitoes (Diptera: Culicidae) in Iran. J Vector Ecol. 30(2): 272–276.
- Sedaghat MM, Linton YM, Oshaghi MA, Vatandoost H, Harbach RE (2003) The *Anopheles maculipennis* complex (Diptera: Culicidae) in Iran: Molecular characterization and recognition of a new species. Bull Entomol Res. 93(6): 527– 535.
- Service MW (1980) A Guide to Medical Entomology. MacMillan, London, p. 226.
- Seyoum A, Kabiru EW, Lwande W, Killeen GF, Hassanali A, Knols BG (2002) Repellency of live potted plants against

Anopheles gambiae from human baits in semi-field experimental huts. Am J Trop Med Hyg. 67(2): 191–195.

- Soleimani-Ahmadi M, Vatandoost H, Shayeghi M, Raeisi A, Abedi F, Eshraghian MR, Madani A, Safari R, Shahi M, Mojahedi A, Poorahmad-Garbandi F (2012) Vector ecology and susceptibility in a malaria-endemic focus in southern Islamic Republic of Iran. East Mediterr Health J. 18(10):1 034–1041.
- Soleimani-Ahmadi M, Vatandoost H, Zare M, Alizadeh A, Salehi M (2014) Community knowledge and practices regarding malaria and long-lasting insecticidal nets during malaria elimination programme in an endemic area in Iran. Malar J. 13(1): 511.
- Tavassoli M, Shayeghi M, Abai MR, Vatandoost H, Khoobdel M, Salari M, Ghaderi A, Rafi F (2011) Repellency effects of essential oils of Myrtle (*Myrtus communis*), Marigold (*Calendula officinalis*) compared with DEET against Anopheles stephensi on human volunteers. Iran J Arthropod-Borne Dis. 5(2): 10–22.
- Tajalli AA, pilehvar B, Mirazadi Z (2012) Phonological effects on the percent and chemical composition of essential oil of *Saturea hortensis* L. Medical Sciences University of Lorestan. 14 (2): 51–57. (Persian).
- Vatandoost H, Sanei dehkordi A, Sadeghi S, Davari B, Karimian F, Abai M, Sedaghat M (2012) Identification of chemical constituents and larvicidal activity of *Kelussia odoratissima* Mozaffarian essential oil against two mosquito vectors *Anopheles stephensi* and *Culex pipiens* (Diptera: Culicidae). Exp Parasitol. 132(4): 470–474.
- Vatandoost H, Khazani A, Rafinejad J, Khoobdel M, Kebriai-Zadeh A, Abai M (2008) Comparative efficacy of neem and dimethyl phthalate (DMP) against

malaria vector, *Anopheles stephensi* (Diptera: Culicidae). Asian Pac J Trop Med. 1(3): 1–6.

- Vatandoost H, Shahi H, Abai MR, Hanafi-Bojd AA, Oshaghi MA, Zamani G (2004) Larval habitats of main malaria vectors in Hormozgan Province and their susceptibility to different larvicides. Southeast Asian J Trop Med Public Health. 35(2): 22–25.
- WHO (2010) World Malaria Report. World Health Organization.

WHO (2013) Malaria entomology and vector control. World Health Organization.

Yaghoobi-Ershadi MR, Akhavan AA, Jahanifard E, Vatandoost H, Amin GH, Moosavi L, Ramazani AR, Abdoli H, Arandian MH (2006) Repellency effect of Myrtle essential oil and DEET against *Phlebotomus papatasi*, under 2965 laboratory conditions. Iran J Public Health. 35(3): 7–13.