Original Article

Current Susceptibility Status of Anopheles stephensi (Diptera: Culicidae) to Different Imagicides in a Malarious Area, Southeastern of Iran

Mohammad Amin Gorouhi¹, *Hassan Vatandoost¹, *Mohammad Ali Oshaghi¹, Ahmad Raeisi², Ahmad Ali Enayati³, Hossein Mirhendi⁴, Ahmad Ali Hanafi-Bojd¹, Mohammad Reza Abai¹, Yaser Salim-Abadi^{1,5}, Fatemeh Rafi¹

¹Department of Medical Entomology and Vector Control, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran

²National Malaria Control Department, CDC, Ministry of Health and Medical Education, Iran ³School of Public Health and Health Sciences Research Centre, Mazandaran University of Medical Sciences, Sari, Iran

⁴Department of Parasitology and Mycology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran

⁵School of Health, Rafsanjan University of Medical Sciences, Rafsanjan, Iran

(Received 22 Nov 2014; accepted 10 Agu 2015)

Abstract

Background: Anopheles mosquitoes are an important group of arthropods due to their role in transmission of malaria. The present study was conducted for determination of susceptibility status of Anopheles stephensi to different imagicides collected from malarious area in Chabahar city, Iran.

Methods: In the present study seven insecticides including: DDT 4%, lambdacyhalothrin 0.05%, deltamethrin 0.05%, permethrin 0.75%, cyfluthrin 0.15% and etofenprox 0.5% were tested based on WHO method. Regression line was plotted for each insecticide using mortality of different exposure times. Bioassay data were analyzed using Probit software and the lethal time for 50% and 90% mortality (LT_{50} and LT_{90}) values were calculated.

Results: The susceptibility levels of field strain of *An. stephensi* to the discriminative dose of different imagicides were determined 100, 98, 96, 89, 82 and 62% for etofenprox, permethrin, deltamethrin, lambdacyhalothrin, cyfluthrin and DDT, respectively. Our finding indicated that *An. stephensi* is resistant to DDT, lambdacyhalothrin and cyfluthrin, and susceptible to etofenprox and permethrin and candidate of resistant to deltamethrin based on WHO criteria.

Conclusion: Our findings indicated that *An. stephensi* is resistant to DDT and some pyrethroid insecticides which can be developed due to application of insecticides in health and agriculture. These results can provide a clue for future chemical control program in the study area.

Keywords: Susceptibility test, Anopheles stephensi, Chabahar, Pyrethroid resistance

Introduction

Mosquitoes as a big group of arthropods play an important role in transmission of many diseases to human such as malaria, filariasis, yellow fever, dengue fever (Horsfall 1955, Tabachnick 1991, Service 2003, Azari-Hamidian 2011). Some species of *Anopheles* mosquitoes are vectors of malaria in different parts of the world. For example, *Anopheles stephensi* Liston (Diptera: Culicidae) is the main malaria vector in Eastern Mediterranean region and south of Asia continent (Zahar 1974, Vatandoost et al. 2006). In Iran there are some species of malaria vectors including: An. stephensi, An. dthali, An. culicifacies, An. fluviatilis, An. superpictus s.l., An. sacharovi, An. maculipennis Complex (Naddaf et al. 2003, Azari-Hamidian 2011, Mehravaran et al. 2011, Oshaghi et al. 2011).

http://jad.tums.ac.ir Published Online: October 04, 2016

^{*}**Corresponding author:** Dr Hassan Vatandoost, E-mail: 493 hvatandoost1@yahoo.com, vatando@tums.ac.ir, Dr Mohammad Ali Oshaghi, E-mail: moshaghi@tums.ac.ir

Before initiating of national malaria control program in 1957, malaria cases were reported from most parts of Iran, Since then, due to implementing of many continuous interventions, malaria confined to south eastern parts of the country including Sistan va Baluchestan, Hormozgan and southern parts of Kerman Provinces (Edrissian 2006, Vatandoost et al. 2011). For controlling of malaria, vector control is one the most important approach which focuses on chemical control of mosquitoes. Up to now different group of insecticides including: organochlorines (DDT, dieldrin and BHC), organophosphates (pirimiphos-methyl, malathion), carbamates (propoxur) and pyrethroids (lambdacyhalothrin, delthamethrin) in different forms of application such as Indoor Residual Spraying (IRS), Insecticide Treated Nets (ITN_S) for adult stage and some organophosphates for larviciding were used in malariaous area of Iran (Salim Abadi et al. 2010, Hanafi-Bojd et al. 2012, Vatandoost and Hanafi-Bojd 2012). Resistance of Anopheles spp to DDT and pyrethroid insecticides were reported from different countries around the world like China, Turkey, India, some countries of Africa and Latin America (Kasap et al. 2000, Hargreaves et al. 2003, Syafruddin et al. 2010, Lol et al. 2013, Soltani et al. 2013, Chang et al. 2014). In Iran many researches have evaluated susceptibility status of malaria vectors against different insecticides (Vatandoost and Hanafi-Bojd 2005, Hanafi-Bojd et al. 2006, Vatandoost et al. 2006, Vatandoost et al. 2011, Vatandoost and Hanafi-Bojd 2012). Approximately in all previous conducted studies on An. stephensi in Iran, resistance to DDT and susceptibility to pyrethroids have been reported, but in 2012 first indication of resistance to pyrethroid compounds was reported from south eastern parts of the country (Vatandoost and Hanafi-Bojd 2012). Resistances to DDT, mainly in the adult stage of An. stephensi, have been widely distributed in Middle-East and Indian subcontinent causing operational problems for control programs (WHO 1985, WHO 1992). This study aims to monitor susceptibility status of main malaria vector, *An. stephensi*, to some insecticides in Chabahar City, Sistan va Baluchestan Province, Iran.

Materials and Methods

Study area

This study was performed in Chabahar seaport $(25^{\circ} 25' \text{ N}, 60^{\circ} 45' \text{ E})$, Sistan va Baluchestan Province of Iran during April to June 2013 (Fig. 1).

Mosquito sampling and rearing

Collected larvae from the study area were transferred to the insectary for rearing under standard conditions (Temperature= 25–29° C, photo-period=12:12 Hours (light: Dark) and Humidity=50–70%). Emerged adult mosquitoes were fed with 10% aqueous sucrose solution.

Adult susceptibility test

Adult susceptibility tests were carried out according to the current World Health Organization method (WHO 2013). For each insecticide mortality rate in various times also were calculated and then regression line to each insecticide plotted using Microsoft Excel (version. 2013).

Insecticide impregnated papers

The following insecticides impregnated papers were supplied according to WHO Test procedure including: DDT 4%, lamb-dacyhalothrin 0.05%, deltamethrin 0.05%, permethrin 0.75%, cyfluthrin 0.15% and etofenprox 0.5%. Mineral oil, and silicon oil impregnated papers were used for organo-chlorine insecticides and pyrethroids as control, respectively (WHO 1981, WHO 2013).

Statistical analysis

Results were analyzed by using of Probit program (Finney 1971). In case of mortality, when the control mortality was between 5% to 20% it was corrected by Abbott's formula (Abbott 1925). Error bars for each mortality were calculated based on statistical method at =5%. The lethal Time for 50% and 90% mortality (LT_{50} and LT_{90}) values and their 95% confidence interval also Probit regression line parameters were determined with Finney method and then the regression line of all Insecticides were plotted using Mi crosoft Excel (version. 2013).

Results

The results of susceptibility test for each insecticides are shown in tables 1,2. Mortality rate and lethal Time for 50% mortality (LT_{50}) of different insecticides were calculated. Our finding indicated that Etofenprox, Deltamethrin, Lambdacyhalothrin, Permethrin, Cyfluthrin and DDT have the lowest to highest LT_{50} value respectively (Fig. 2).



Fig. 1. The map of Chabahar City representing rural districts, Sistan and Baluchistan Province (Study area), Iran

Table 1.	Probit	regression	line para	meters of	Anopl	heles ste	phensi e	xposed to	different	insecticide
					· · · r					

Insecticide	Α	B±SE	LT ₅₀ , 95% C.I. (Second)	LT ₉₀ , 95% C.I. (Second)	X ² (df)	P value
			75	626		
Etofonnrov () ()5%	-2.68	1.33 ± 0.14	104	957	5.11 (3)	>0.05
Etorenprox 0.05 78			138	1749		
			277	984		
Permethrin 0.75%	-5.61	2.22 ± 0.21	335	1266	2.89 (2)	>0.05
			401	1775		
			656	5121		
Cyfluthrin0.15%	-3.79	1.3±0.12	812	7805	1.97(4)	>0.05
			1010	14160		

http://jad.tums.ac.ir Published Online: October 04, 2016

J Arthropod-Borne Dis, December 2016, 10(4): 493-500

MA Gorouhi et al.: Current Susceptibility ...

		LUDIC II Contin	laca			
	-3.25		185	1414		
Lambdacyhalothrin 0.05%		1.36 ± 0.12	246	2146	0.04(2)	>0.05
			324	3791	-	
			101	1194		
Deltamethrin 0.05%	-2.69	1.22 ± 0.15	159	1785	0.95(2)	>0.05
			221	3277	_	
			2820	7560		
DDT 4%	-4.47	2.5±0.33	3240	10200	6.99(3)	>0.05
			3840	16560	-	

Table 1. Continued...

A= y-intercept, B= the slope of the line, SE= Standard error, CI= confidence interval, x^2 = heterogeneity about the regression line, df= degree of freedom, P> 0.05 =represent no heterogeneity in the population of tested mosquitos.

 Table 2. Mortality rate and susceptibility status of Anopheles stephensi exposed to different insecticides Chabahar, southeastern Iran, 2013

Insecticide	MR±EB*	Resistance status **
Deltamethrin	96±3.8	RC
Lmbdacyhalothrin	89 ± 2.8	R
Cyfluthrin	82±3.5	R
Permethrin	98±1	S
Etofenprox	100	S
DDT	62 ± 4.8	R
control		-

^{*}Mortality Rate±Error Bar

**R Resistance, RC Resistant Candidate, T Tolerance, S Susceptible



Fig. 2. Regression lines of Anopheles stephensi exposed to different insecticides (field population), 2013

496

http://jad.tums.ac.ir Published Online: October 04, 2016

Discussion

In the current study seven insecticides including: DDT 4%, lambdacyhalothrin 0.05%, deltamethrin 0.05%, permethrin 0.75%, cyfluthrin 0.15% and etofenprox 0.5% were used to determine susceptibility status of An. stephensi collected from Chabahar City. Based on WHO criteria that suggested (98-100% mortality indicates susceptibility, 90-97% mortality indicates resistance candidate (more investigation is needed) and less than 90% mortality suggests resistance (WHO 2013). Results indicated that species is resistant to DDT, cyfluthrin and Lambdacyhalothrin, However, susceptible to permethrin and Etophenprox. The indication of resistant to deltamethrin at the early stages of evolution has also be documented. Our findings reveal that An. stephensi is resistant to DDT which is in line with previous researches results that have been performed in our study area (Vatandoost and Hanafi-Bojd 2012, Fathian et al. 2015). Majority of susceptibility tests which performed during the past decade in different malarious area revealed resistance to DDT in southern part of Iran (Borhani 2004, Vatandoost et al. 2005, Vatandoost et al. 2006) as well as in the most distribution area of An. stephensi in the world (Rathor et al. 1980, Thavaselvam et al. 1993, Tikar et al. 2011, Chang et al. 2014, Singh et al. 2014). Furthermore, there are many resistance reports to DDT in other species of Anopheles mosquitoes from different part of the world (Hemingway and Ranson 2000, Hemingway et al. 2002, Zahirnia et al. 2002, Lak et al. 2002, Balkew et al. 2006, Raghavendra et al. 2010, Tikar et al. 2011, Vatandoost et al. 2011, Nardini et al. 2013, Wang et al. 2013). In the present study resistance to cyfluthrin and lambdacyhalothrin were indicated and these findings are in line with previous research results that have been conducted in the same area (Vatandoost and Hanafi-Boid 2012).

On the other hand our finding about cyfluthrin susceptibility status is not in concordance with another research that has been performed previously in the same area by Fathian et al. (2015) that showed this species is susceptible to cyfluthrin. (Fathian et al. 2015). It may be due to different sampling localities. Resistance of An. stephensi to pyrethroid compounds were reported from its different distribution regions, for instance in the study was performed by Rathor et al. (2013) in Punjab Province of Pakistan, resistance to three commonly used pyrethroids, permethrin, lambda-cyhalothrin, and deltamethrin were indicated from the majority of test localities (Rathor et al. 2013). In the present study An. stephensi was susceptible to Etophenprox and Permethrin that these findings are in parallel with other previous conducted researches results (Vatandoost et al. 2005). In the current study deltametthrin was indicated as resistant candidate so that more investigation is needed. Molecular and biochemical assays for this species as a main malaria vector must be conducted for accurate evaluating of resistance status of pyrethroid insecticides specially those commonly used in malaria control program.

Conclusion

In the present study *An. stephensi* was found resistant to DDT and some pyrethroid insecticides. This enhanced resistance status may be due to previous chemical control programs against malaria vectors, such as IRS/ITNs or insecticide application in agriculture. However, more investigation for determination of resistance mechanisms is necessary. Furthermore regular monitoring of resistance status by standard bioassay tests and other complementary methods especially in active foci of malaria transmission is suggested. J Arthropod-Borne Dis, December 2016, 10(4): 493–500

Acknowledgements

This article is a part of the first author's dissertation for fulfillment of a PhD degree in Medical Entomology and Vector Control from Department of Medical Entomology and Vector Control, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran. This study was financially supported by the Deputy for Research, Tehran University of Medical Sciences. Authors are grateful for Dr Amini head of Chabahar health center and all his staff for their kind collaboration. The authors declare that there is no conflict of interest.

References

- Abbott WS (1965) A method of comparing the effectiveness of an insecticide. J Econ Entomol. 18: 265–267.
- Azari-Hamidian S (2011) Larval habitat characteristics of the genus *Anopheles* (Diptera: Culicidae) and a checklist of mosquitoes in Guilan Province, Northern Iran. J Arthropod-Borne Dis. 5(1): 37–53.
- Balkew M, Elhassen I, Ibrahim M, Gebre-Michael T, Engers H (2006) Very high DDT-resistant population of *Anopheles pharoensis* Theobald (Diptera: Culicidae) from Gorgora, northern Ethiopia. Parasite. 13(4): 327– 329.
- Borhani N, Vatandoost H (2004) Susceptibility and irritability levels of main malaria vectors to synthetic pyrethroids in the endemic areas of Iran. Acta Med Iran. 42(4): 240–247.
- Chang X, Zhong D, Fang Q, Hartsel J, Zhou G, Shi L, Fang F, Zhu C, Yan G (2014) Multiple resistances and complex mechanisms of *Anopheles sinensis* Mosquito: a major obstacle to mosquito-borne diseases control and elimination in china. PLoS Negl Trop

MA Gorouhi et al.: Current Susceptibility ...

Dis. 8(5): e2889. Edrissian G (2006) Malaria in Iran: Past and present situation. Iran J Parasitol. 1(1): 1–14.

- Fathian M, Vatandoost H, Moosa-Kazemi SH, Raeisi A (2015) Susceptibility of Culicidae mosquitoes to some insecticides recommended by WHO in a malaria endemic area of southeastern Iran. J Arthropod-Borne Dis. 9(1): 22–34.
- Hanafi-Bojd AA, Vatandoost H, Jafari R (2006) Susceptibility status of *Anopheles dthali* and An. fluviatilis to commonly used larvicides in an endemic focus of malaria, southern Iran. J Vector Borne Dis. 43(1): 34–38.
- Hanafi-Bojd AA, Vatandoost H, Oshaghi M, Haghdoost AA, Shahi M, Sedaghat MM, Abedi F, Yeryan M, Pakari A (2012) Entomological and epidemiological attributes for malaria transmission and implementation of vector control in southern Iran. Acta Trop. 121(2): 85–92.
- Hargreaves K, Hunt R, Brooke B, Mthembu J, Weeto MM, Awolola TS, Coetzee M (2003) Anopheles arabiensis and An. quadriannulatus resistance to DDT in South Africa. Med Vet Entomol. 17(4): 417–422.
- Hemingway J, Field L, Vontas J (2002) An overview of insecticide resistance. Science. 298(5591): 96–97.
- Hemingway J, Ranson H (2000) Insecticide resistance in insect vectors of human disease. Annu Rev Entomol. 45(1): 371–391.
- Horsfall WR (1955) Mosquitoes: their bionomics and relation to disease. Hanofner publishing, New York, p. 723.
- Kasap H, Kasap M, Alptekin D, Lüleyap Ü, Herath P (2000) Insecticide resistance in *Anopheles sacharovi* Favre in southern Turkey. Bull World Health Organ. 78(5): 687–692.
- Lak SS, Vatandoost H, Entezarmahdi M,

Ashraf H, Abai M, Nazari M (2002) Monitoring of insecticide resistance in *Anopheles sacharovi* (Favre, 1903) in borderline of Iran, Armenia, Naxcivan and Turkey, 2001. Iran J Public Health. 31(3–4): 96–99.

- Lol JC, Castellanos ME, Liebman KA, Lenhart A, Pennington PM, Padilla NR (2013) Molecular evidence for historical presence of knock-down resistance in *Anopheles albimanus*, a key malaria vector in Latin America. Parasit Vectors. 6(1): 268.
- Mehravaran A, Oshaghi M, Vatandoost H, Abai MR, Ebrahimzadeh A, Roodi AM, Grouhi A (2011) First report on *Anopheles fluviatilis* U in southeastern Iran. Acta Trop. 117(2): 76–81.
- Naddaf S, Oshaghi M, Vatandoost H, Assmar M (2003) Molecular characterization of *Anopheles fluviatilis* species complex in the Islamic Republic of Iran. East Mediterr Health J. 9(3): 257–265.
- Nardini L, Christian RN, Coetzer N, Koekemoer LL (2013) DDT and pyrethroid resistance in *Anopheles arabiensis* from South Africa. Parasit Vectors. 6(1): 229.
- Oshaghi MA, Vatandoost H, Gorouhi A, Abai MR, Madjidpour A, Arshi S, Mehravaran A (2011) Anopheline species composition in borderline of Iran-Azerbaijan. Acta Trop. 119(1): 44–49.
- Raghavendra K, Verma V, Srivastava H, Gunasekaran K, Sreehari U, Dash A (2010) Persistence of DDT, malathion and deltamethrin resistance in *Anopheles culicifacies* after their sequential withdrawal from indoor residual spraying in Surat district, India. Indian J Med Res. 132(3): 260–264.
- Rathor H, Nadeem G, Khan I (2013) Pesticide susceptibility status of *Anopheles* mosquitoes in four flood-affected districts of South Punjab, Pakistan. Vector Borne Zoon Dis. 13(1): 60–66.

- Rathor H, Toqir G, Reisen W (1980) Status of insecticide resistance in anopheline mosquitoes of Punjab Province, Pakistan. Southeast Asian J Trop Med Publ Health. 11(3): 332–340.
- Salim Abadi Y, Vatandoost H, Rassi Y, Abaei MR, Sanei Dehkordi AR, Paksa A (2010) Evaluation of biological control agents for mosquitoes control in artificial breeding places. Asian Pac J Trop Med. 3(4): 276–277.
- Service MW (2003) Medical Entomology for Students. Vol. 3. United Kingdom: Cambridge. University Press, Cambridge.
- Singh RK, Kumar G, Mittal PK (2014) Insecticide susceptibility status of malaria vectors in India: A review. Int J Mosq Res. 1(1): 5–9.
- Soltani A, Vatandoost H, Oshaghi MA, Enayati AA, Raeisi A, Eshraghian MR, Soltan-Dallal MM, Hanafi-Bojd AA, Abai MR, Rafi F (2013) Baseline Susceptibility of Different Geographical Strains of *Anopheles stephensi* (Diptera: Culicidae) to Temephos in Malarious Areas of Irana. J Arthropod-Borne Dis. 7(1): 56–65.
- Syafruddin D, Hidayati A, Asih P, Hawley W, Sukowati S, Lobo NF (2010) Detection of 1014F kdr mutation in four major Anopheline malaria vectors in Indonesia. Malar J. 9: 315.
- Tabachnick WJ (1991) Evolutionary genetics and arthropod-borne disease: the yellow fever mosquito. American Entomol. 37(1): 14–26.
- Thavaselvam D, Kumar A, Sumodan P (1993) Insecticide susceptibility status of *Anopheles stephensi*, *Culex quinquefasciatus* and *Aedes aegypti* in Panaji, Goa. Indian J Malariol. 30(2): 75–79.
- Tikar S, Mendki M, Sharma A, Sukumaran D, Veer V, Prakash S, Parashar BD (2011) Resistance status of the malaria vector mosquitoes, *Anopheles stephensi*

J Arthropod-Borne Dis, December 2016, 10(4): 493–500

and *Anopheles subpictus* towards adulticides and larvicides in arid and semi-arid areas of India. J Insect Sci. 11: 85.

- Vatandoost H, Mashayekhi M, Abaie M, Aflatoonian M, Hanafi-Bojd A, Sharifi I (2005) Monitoring of insecticides resistance in main malaria vectors in a malarious area of Kahnooj District, Kerman Province, southeastern Iran. J Vector Borne Dis. 42(3): 100–108.
- Vatandoost H, Oshaghi M, Abaie M, Shahi M, Yaaghoobi F, Baghaii M, Hanafi-Bojd AA, Zamani G, Townson H (2006) Bionomics *of Anopheles stephensi* Liston in the malarious area of Hormozgan Province, southern Iran, 2002. Acta Trop. 97(2): 196–203.
- Vatandoost H, Emami S, Oshaghi M, Abai MR, Raeisi A, Piazzak N, Mahmoodi M, Akbarzadeh K, Sartipi M (2011) Ecology of malaria vector *Anopheles culicifacies* in a malarious area of Sistan va Baluchestan Province, southeast Islamic Republic of Iran. East Mediterr Health J. 17(5): 439–445.
- Vatandoost H, Hanafi-Bojd AA (2012) Indication of pyrethroid resistance in the main malaria vector, *Anopheles stephensi* from Iran. Asian Pac J Trop Med. 5(9): 722–726.
- Wang DQ, Xia ZG, Zhou SS, Zhou XN, Wang RB, Zhang QF (2013) A potential threat to malaria elimination: extensive deltamethrin and DDT re sistance

sistance to *Anopheles sinensis* from the malaria-endemic areas in China. Malar J. 12(1): 164.

- WHO (1981) Instruction for determining the susceptibility or resistance of adult mosquitoes to organochlorine, organphosphate and carbamate insecticides diagnostic test. WHO/VBC/.
 81.806. WHO (2013) Malaria Entomolopgy and Vector Control. Participant's Guide, Test procedures for insecticide resistance monitoring in malaria vector mosquitoes. World Health Organization, Geneva, Switzerland.
- WHO (1985) Resistance of vectors of disease to pesticides. X report of the expert committee on vector control. WHO Tech Rep Ser 734.
- WHO (1992) Vector resistance to pesticides. XV report of the expert committee on vector control. WHO Tech Rep Ser 818.
- Zahar A (1974) Review of the ecology of malaria vectors in the WHO Eastern Mediterranean Region. Bull World Health Organ. 50(5): 427.
- Zahirnia A, Vatandoost H, Nateghpour M, Djavadian E (2002) Insecticide resistance/ susceptibility monitoring in *Anopheles pulcherrimus* (Diptera: Culicidae) in Ghasreghand district, Sistan and Baluchistan Province, Iran. Iran J Public Health. 31(1–2): 11–14.