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Original scientific paper

Electrochemical sensor based on MWCNTs/Co₃O₄/SPGE for simultaneous detection of Sudan I and Bisphenol A

Afsaneh Hajializadeh[⊠]

Department of Natural Resources, Sirjan Branch, Islamic Azad University, Sirjan, Iran Corresponding author: [™]<u>hajalizadeh.813@qmail.com</u> Received: December 13, 2021; Accepted: December19, 2021; Published: February 9, 2022

Abstract

This paper presents a sensitive simultaneous detection procedure for Sudan I and bisphenol A based on the multiwalled carbon nanotubes (MWCNTs)/Co₃O₄ nanocomposite modified screen-printed graphite electrode (SPGE). This MWCNTs/Co₃O₄ nanocomposite was prepared by the hydrothermal technique, and characterized by Fourier transform infrared (FT-IR) spectroscopy, field emission scanning electron microscopy (FE-SEM) and X-ray diffraction (XRD). The electrochemical properties of MWCNTs/Co₃O₄ nanocomposite modified SPGE were analyzed by cyclic voltammetry (CV), differential pulse voltammetry (DPV), chronoamperometry (CHA), as well as linear sweep voltammetry (LSV). From the electrochemical results, a synergy between MWCNTs and Co_3O_4 nanoparticles (NPs) was detected as improved interfacial electron transfer, which was accompanied by a greater catalytic function for electrochemical oxidation of Sudan I. Based on the optimized condition, MWCNTs/Co₃O₄/SPGE exhibited the linear dynamic ranging between 0.05 and 600.0 μ M detection of Sudan I with a limit of detection (LOD) 0.02 μ M. Also, the as-prepared electrode was assessed for simultaneous detection of Sudan I and Bisphenol A. In the course of electrooxidation processes of these analytes, two complete peaks at 380 and 520 mV were observed on the modified electrode. At the end, utility of this new electrochemical sensor was performed to determine Sudan I and Bisphenol A in some real samples with good accuracy and precision.

Keywords

Electroanalysis; voltammetry; azo dyes; food analysis; modified electrodes; SPGE; MWCNTs/Co $_3O_4$

Introduction

Sudan I (1-phenylazo-2-naphtol) is a synthetic azo dye commonly employed as one of the coloring materials in industrial products like shoes, textile, oil, plastic, printing ink, cosmetics, as well as floor polish [1]. Moreover, Sudan I is also proposed as food additive material, sauces, chili powder, readymade meals, and chutneys because of its bright red colour, affordability, and coloring fastness.

However, in the case of ingestion of the materials consisting of Sudan I by human bodies, carcinogenic amines would be produced following the metabolism process [2]. Hence, European Union (EU) and Food Standards Agency have strongly forbidden the utilization of Sudan I as an additive in foodstuff because of its higher teratogenetic and carcinogenic nature. Despite the legal prohibitions, Sudan I circulates in the food markets due to temptations for massive profits [3]. Thus, the determination of Sudan I residues in the food products would be necessary to preclude health risks to humans.

Experts in the field have introduced Bisphenol A (4,4'-(propane-2,2-diyl)diphenol) (BPA) as one of the key monomeric compounds applied in fabricating polycarbonate and epoxy in plastic outputs. They have also been employed for making diverse plastic products like water and milk bottles, food containers, bottles for infant feeding, dental fillings, bottle tops, table-ware, food cans, and other storage vessels [4-6]. Moreover, BPA can migrate from plastic products into the environment and food chain resulting in considerable humans' exposure [7]. Additionally, BPA has been proposed as one of the endocrine-disrupting compounds associated with several sorts of health issues like cancer progression and reproduction issues [8,9]. Considering public health, it would be of high significance to develop a valid analytical procedure to determine BPA.

Several analytical methods such as gas chromatography-mass spectrometry (GC–MS) [10], highperformance liquid chromatography (HPLC) [11,12], fluorescence [13], capillary electrophoresis [14], surface-enhanced Raman scattering spectroscopy [15], and chemiluminescence [16,17] have been developed for detecting Bisphenol A and Sudan I. These techniques showed good outputs, although, they might be laborious and demand numerous raw materials, professional workforce, and costly instruments.

In this regard, researchers have also studied electrochemical sensing as one of the very attractive fields in analytical chemistry [18-22]. Electrochemical methods are broadly employed for the analysis of biological, environmental, food, pharmaceutical and industrial compounds due to merits such as fast responses, higher sensitivity, lower costs, portability, and ease of operation [23-29].

It is notable that as an electrochemical sensing platform, the screen-printed electrode (SPE) has been largely attracted because of merits like easier application, portability, and affordability [30,31]. Hence, the screen-printed technology significantly contributes to the transition from the conventional unwieldy electrochemical cells to the portable miniaturized electrodes, satisfying the on-site analysis requirements [32,33].

In the electrochemical approach, using a proper electrode is of utmost importance. In this regard, developing the modified substances for improving the electrochemical response of the electrodes would be a challenging task [34-40]. In recent years, researchers have focused on the designing and synthesizing nanomaterials for various applications due to their unique physical and chemical properties [41-43]. Investigations demonstrated that electrodes modified by catalytically active nanomaterials might enhance efficient mass transfer, achieve more acceptable control, and enhance the active specific surface area in a local micro-environment, improving the sensitivity and selectivity of electrochemical sensors [44-49].

Experts in the field have largely employed the carbon nanotubes (CNTs) in the fabrication of electrochemical sensors because of the respective merits like higher carrier mobility, a larger surface-to-volume ratio, good conductivity, and flexibility. CNTs are specific carbon materials with cylindrical curled graphitic sheets [50]. Numerous investigations have shown that constructing the hybrid catalysts *via* immobilization of metal and metal oxide nanoparticles (NPs) on carbon nanotubes can improve their electrochemical responses [51,52]. Furthermore, nanostructured transition metal

oxides contribute significantly to the improvement of sensitivity and stability of sensors. It should be mentioned that among the family of transition metal oxides, nanostructures of cobalt oxide (Co_3O_4) exhibited specific electrocatalytic activity for detecting different compounds [53-55]. The possible cause would be higher activity and selectivity of the metal oxide catalysts resulting from the differences in oxygen defects, oxygen absorbed in various states of cobalt in Co_3O_4 (a mixed valance state of Co(II) and Co(III)), as well as oxygen holes [56].

In the present report, $MWCNTs/Co_3O_4$ nanocomposite was prepared and employed for the modification of SPGE. The modified SPGE was then utilized to simultaneously detect Sudan I and Bisphenol A.

Experimental

Chemicals and instrumentations

According to the research design, each chemical was of the analytical reagent grade without additional purification. $Co(CH_3CO_2)_2 4H_2O$, sodium hydroxide, ethanol, $NH_3 \cdot H_2O$ (25 to 28 wt.%), and MWCNTs-COOH (purity >95 %) were supplied from Aldrich. Sudan I, Bisphenol A and other chemicals were supplied from the Merck Company (Darmstadt, Germany). Phosphate buffer solutions (PBS) were prepared from H_3PO_4 and the respective salts (KH_2PO_4 , K_2HPO_4 and K_3PO_4) (Merck). Food samples such as ketchup sauce, tomato paste and chili powder were purchased from a local store in Kerman, Iran. Nano-pure ($18 M\Omega \cdot cm$) water from a Milli-pore MilliQ system (Bedford) was applied to prepare all solutions.

Electrochemical experiments were recorded using an Auto-lab potentiostat/galvanostat (PGSTAT-302N, Eco Chemie, The Netherlands). Electrochemical tests were carried out using SPGE (DropSens, DRP-110, Spain). Electrochemical cells consisted of a three-electrode arrangement with graphite as a counter electrode and graphite in 4 mm diameter as a working electrode. In addition, a silver pseudo-reference electrode was used to complete the circuit. pH values were measured using a digital pH meter (Metrohm 710).

Fourier transform infrared (FT-IR) spectra of the synthesized nanocomposite were recorded using a spectrometer (SHIMADZU Corporation, Kyoto, Japan) between 4000 and 400 cm⁻¹, applying KBr pellets. XRD patterns were examined with the XRD device model X'Pert Pro, the Netherlands, while EDX and FE-SEM images were obtained using MIRA3TESCAN-XMU.

Synthesis of the MWCNT₅/Co₃O₄ nanocomposite

A hydrothermal approach was used for preparing MWCNTs decorated with Co_3O_4 NPs. Firstly, 20 mg of MWCNTs-COOH were dispersed into 12.5 mL of ethanol, and 0.25 g $Co(CH_3CO_2)_2$ '4H₂O was added to the mixture for sonication during 40 min. Then, 2.5 mL of aqueous NH₃ were dropped into the solution and vigorously stirred. The homogeneous slurry was taken to a Teflon-lined stainless steel autoclave, sealed, and heated at 150 °C for 3 h. After cooling the solution to the room temperature, water and ethanol were used to wash the products and achieved MWCNT₅/Co₃O₄ nanocomposite followed by the drying process at 60 °C for 8 hours.

Modification of SPGE

The construction of MWCNT_S/Co₃O₄ nanocomposite over the SPGE surface was accomplished in this way: 1 mg of MWCNT_S/Co₃O₄ nanocomposite was suspended in 1 mL of the distilled water for forming the suspension, which was sonicated for 20 min to disperse the nanocomposite. Finally, 5 μ L aliquot of the suspension was pipetted over the surface of an SPGE and dried at the ambient temperature.

The surface areas of MWCNTs/Co₃O₄/SPGE and bare SPE were obtained by CV using 1 mM $K_3Fe(CN)_6$ at different scan rates. Using Randles-Ševčik formula for MWCNTs/Co₃O₄/SPGE, the electrode surface was determined to be 0.08 cm², about 2.5 times greater than bare SPGE.

Preparation of real samples

In this step, 20 mL ethanol were used for mixing ketchup sauce, tomato paste, as well as chili powder, and the obtained mixtures were filtrated after 20 min of ultra-sonication, with 100 mL volumetric flasks employed for collecting the liquid phase. Then, treatment was performed for three times and ethanol was applied for diluting the filtrated sample to the intended volumes.

The tap water gathered from the laboratory was analyzed in twelve hours. Before the analysis process, 0.22 μ M cellulose acetate membrane was used to filter the water.

Results and discussion

Characterization of MWCNT₅/Co₃O₄ nanocomposite

The FTIR spectrum of MWCNT₅/Co₃O₄ nanocomposite is shown in Figure 1. For MWCNT₅/Co₃O₄ nanocomposite, two sharp bands are found at 660 and 562 cm⁻¹, indicating presence of Co²⁺ and Co³⁺ in the spinal Co-O stretching vibration, wherein Co²⁺ is associated with the tetrahedral coordinate and Co³⁺ to the octahedral coordinate. The spectra bands at 2921 cm⁻¹ to 2854 cm⁻¹ are attributed to the asymmetric or symmetric stretching vibrations of C-H in CH₂ and –CH₃ group in MWCNTs. The peak at 1641 cm⁻¹ (aromatic C=C) can be ascribed to the stretching vibration C=C of sp² hypridization in CNT back-bone. The band at ~3442 cm⁻¹ is attributed to O–H stretching vibration. FT-IR spectrum confirmed the decoration of MWCNTs with Co₃O₄ nanoparticles.



The crystal structure of MWCNTs/Co₃O₄ nanocomposite was identified by XRD. The XRD peaks of the treated MWCNTs/Co₃O₄ and MWCNTs are shown in Figure 2. As can be seen, MWCNTs patterns have two key diffraction peaks at 2 θ =25.9° and 43.1°, which belong to the (002) and (100) planes of graphite. Diffraction peaks at 2 θ values of 19.08, 31.3, 36.9, 38.6, 44.9, 55.7, 59.4 and 65.3°° observed for MWCNT/Co₃O₄ nanocomposite correspond to several planes (111) (220) (311) (222) (400) (422) (511) as well as (440) of Co₃O₄. These characteristic peaks belong to the cubic phase of Co₃O₄ (JCPDS, PDF, File No. 00-042-1467). Furthermore, the peaks have higher intensity reflecting acceptable crystallinity of the samples. However, we did not find any impurity, representing the

substantial contribution of the MWCNT/Co₃O₄ nanocomposite. Finally, Debye-Sheerer equation was used for calculating crystallite size (D) of Co₃O₄ NPs:

$$D = K\lambda / \beta \cos \theta$$

(1)

Here, λ represents the wavelength, β the peak width at half maximum, and θ diffraction angle. Dimension of Co₃O₄ nanoparticles is found to 23.8 nm.



Figure 2. XRD patterns of MWCNTs and MWCNTs/Co₃O₄ nanocomposite

Morphology of MWCNTs/Co₃O₄ nanocomposite was investigated using FE-SEM. Figure 3 shows the FE-SEM image of MWCNTs/Co₃O₄ nanocomposite, where the presence of Co₃O₄ nanoparticles coated on the surface of MWCNTs could be evidenced.



Figure 3. FE-SEM image of MWCNTs/Co₃O₄ nanocomposite

Electrocatalytic oxidation of Sudan I at MWCNTs/Co₃O₄/SPGE

For ensuring the best response of MWCNTs/Co₃O₄/SPGE in detecting Sudan I, the influence of the solution pH on the reaction was evaluated by recording voltammetric behavior of MWCNTs/Co₃O₄/SPGE toward 100.0 μ M of Sudan I in pH range between 2.0 and 9.0 at the scan rate 50 mV/s. It is shown in Figure 4 that the maximum oxidation current of Sudan I was obtained at pH 7.0. Hence, pH 7.0 was chosen for all further measurements in 0.1 M PBS.



In the next step, the potential application of MWCNTs/Co₃O₄/SPGE for electrooxidation and determination of Sudan I was examined by cyclic voltammetry (CV). Figure 5 depicts the CV response of oxidation of 200.0 μ M of Sudan I on (a) bare SPGE, and (b) MWCNTs/Co₃O₄/SPGE in 0.1 M PBS, pH=7.0 at the scan rate 50 mVs⁻¹. As seen in Figure 5, oxidation potential decreased and the peak current height increased for electroxidation of Sudan I at MWCNTs/Co₃O₄/SPGE as compared to bare SPGE. These are probably caused by increasing the rate of the electron transfer process at the modified SPGE electrode, due to the higher surface area, better conductivity of MWCNTs, and electrocatalytic activity of Co₃O₄ nanoparticles.



Figure 5. CV response of 200.0 μM Sudan I in 0.1 M PBS of pH 7.0 at (a) bare SPGE and (b) MWCNTs/Co₃O₄/SPGE

Effect of potential scan rate on electrochemical response of Sudan I at MWCNTs/Co₃O₄/SPGE

For obtaining information on the kinetics of electrode reactions, the linear sweep voltammetry (LSV) at several scan rates from 10 to 400 mV s⁻¹ in 0.1 M PBS, pH 7.0 was applied, and the results

are illustrated in Figure 6. At higher scan rates, a gradual increase of the oxidation peak current can be observed. Obtained voltammograms are irreversible, as were already seen in CV responses shown in Figure 5. Also, Figure 6 shows that the oxidation peak potential is shifted slightly towards positive potentials with increase of the scan rate. According to the linear plot of the oxidation peak current *versus* the square root of the scan rate, shown in the inset of Figure 6, the linear regression eqation was derived. This result indicates that the oxidation process of Sudan I at MWCNTs/Co₃O₄/SPGE electrode is under diffusion control.





To define the electron transfer coefficient (α) between Sudan I and MWCNTs/Co₃O₄/SPGE electrode, Tafel diagram (*E vs.* log *I*) was drawn in the inset of Figure 7, using data of the ascending section of the voltammogram registered at 10 mVs⁻¹ for 50.0 μ M of Sudan I (Figure 7). The calculated slope from the linear plot was equal to 0.084 V⁻¹. From the slope, α value was estimated to 0.3.





CHA Studies

To measure the diffusion coefficient of Sudan I, chronoamperometry was performed by applying the potential step from 0.0 to 0.43 V.. Figure 8 shows the single-step chronoamperograms recorded for MWCNTs/Co₃O₄/SPGE in the presence of several concentrations of Sudan I. As expected, when Sudan I concentration is increased, the anodic current also increases. The experimental plots of current vs. $t^{-1/2}$ for different concentrations of Sudan I are shown in Figure 8A. The slopes of the straight lines versus the concentration of Sudan I is shown in Figure 8B. Diffusion coefficient was calculated from the slope of the straight line and it equals to 2.0×10⁻⁵ cm² s⁻¹.



Figure 8. Chronoamperograms obtained for MWCNTs/Co₃O₄/SPGE in 0.1 M PBS, pH 7.0 at different concentrations of Sudan I (a–f relate to 0.1, 0.4, 0.6, 0.9, 1.4 and 2.0 mM). Inset A: plot of I vs. t^{-1/2} obtained from chronoamperograms a to f. Inset B: plot of straight line slope vs. concentration of Sudan I.

Detection of Sudan I at MWCNTs/Co₃O₄/SPGE by DPV technique

Differential pulse voltammetry (DPV) technique was employed for determining Sudan I concentration due to its higher sensitivity as well as accuracy leading to lower limit of detection (LOD) values. Figure 9 represents differential pulse voltammograms observed at different concentrations of Sudan I in 0.1 M PBS, pH 7.0 (step potential = 0.01 V and pulse amplitude = 0.025 V). Obviously, oxidation peak current increased with the increase of Sudan I concentration and the inset of Figure 9 demonstrates the linearity of current peak height and concentration of Sudan I. The linear regression has an equation $I_{pa} = 0.0817 C_{sudan I} + 1.1311 (R^2 = 0.9997)$. The sensitivity of MWCNTs/Co₃O₄/SPGE is 0.0817 μ A μ M⁻¹.

The detection limit, C_m, of Sudan I was obtained using the equation (2):

$$C_{\rm m} = 3s_{\rm b}/m$$

In the above equation, *m* is the slope of the calibration plot ($0.817 \ \mu A \ \mu M^{-1}$) and s_b is the standard deviation of the blank response obtained from 20 replicate measurements of the blank solution. The detection limit of Sudan I was calculated as 0.02 μM .

(2)



Figure 9. DPV responses of different concentrations of Sudan I at MWCNTs/Co₃O₄/SPGE in 0.1 M PBS, pH 7.0 (a-l refer to 0.05, 0.5, 5.0, 15.0, 30.0, 70.0, 100.0, 200.0, 300.0, 400.0, 500.0 and 600.0 μ M). Inset: calibration curve of DPV peak current against concentration of Sudan I

Simultaneous detection of Sudan I and Bisphenol A at MWCNTs/Co₃O₄/SPGE

A simultaneous test of Sudan I and Bisphenol A was carried out by DPV (step potential = 0.01 V and pulse amplitude = 0.025 V) in 0.1 M PBS, pH 7.0. In Fig. 10, two distinctive oxidation peaks of Sudan I and Bisphenol A could be discerned. The peak currents increased linearly with the increase of analyte concentrations without any interference (Figures 10A and 10B). Therefore, a possible simultaneous assay of Sudan I and Bisphenol A could be made with MWCNTs/Co₃O₄/SPGE sensor.



Figure 10. DPVs of MWCNTs/Co3O4/SPGE in 0.1 M PBS, pH 7.0 containing different concentrations of Sudan I and Bisphenol A (a-f refer to mixed solutions of 0.5 + 0.5, 30.0 + 35.0, 100.0 + 125.0, 200.0 + 225.0, 400.0 + 475.0, and 600.0 + 700.0 μM of Sudan I and Bisphenol A, respectively). Inset A: plot of peak current as a function of Sudan I concentration. Inset B: plot of peak current as a function of Bisphenol A concentration

Analysis of the real samples

To investigate the possibility of using MWCNTs/Co₃O₄/SPGE for the simultaneous detection of Sudan I and Bisphenol A in the real samples, electrochemical analysis were carried out in tomato paste, ketchup sauce, tap water and chili powder according to the standard addition approach. Table 1 reports the outputs and achieved recovery percentages of 97.6 to 103.6 % for these samples. These observed outputs suggest that MWCNTs/Co₃O₄/SPGE has acceptable practical viability for simultaneous detection of Sudan I and Bisphenol A.

Sample	Concentration spiked, µM		Concentration found, μM		Recovery,%		RSD, %	
	Sudan I	Bisphenol A	Sudan I	Bisphenol A	Sudan I	Bisphenol A	Sudan I	Bisphenol A
Ketchup sauce	0	0	-	-	-	-	-	-
	5.0	5.5	4.9	5.7	98.0	103.6	2.3	3.5
	7.0	7.5	7.2	7.4	102.9	98.7	2.7	2.1
Tomato pste	0	0	-	1.3	-	-	2.7	-
	4.0	4.5	4.1	4.4	102.5	97.8	3.0	1.9
	6.0	6.5	5.9	6.6	98.3	101.5	2.2	3.5
Chilli powder	0	0	-	-	-	-	-	-
	6.0	6.5	5.9	6.7	98.3	103.1	2.4	2.1
	8.0	8.5	8.1	8.3	101.2	97.6	2.8	1.6

Conclusion

In this work, MWCNT_S/Co₃O₄ nanocomposite modified SPGE was prepared and introduced to simultaneously detect Sudan I and Bisphenol A. This modified electrode exhibited good electrocatalytic activity for the oxidation of Sudan I in 0.1 M PBS, pH 7.0 solution. The MWCNT_S/Co₃O₄/SPGE linearly responded to Sudan I in the range from 0.05 to 600.0 μ M with the LOD of 0.02 μ M. Also, the application of MWCNT_S/Co₃O₄ modified SPGE to simultaneously determine Sudan I and Bisphenol A was investigated. According to the results, two well-separated oxidation signals of Sudan I and Bisphenol A were obtained. Finally, the developed sensor system showed considerable potency for detection of Sudan I and Bisphenol A in some real samples including tomato paste, ketchup sauce, chili powder, and tap water.

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