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Original scientific paper

An electrochemical sensor for detection of vanillin in food samples using CuFe₂O₄ nanoparticles/ionic liquids modified carbon paste electrode

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Abstract

A highly selective electrochemical sensor modified with $CuFe_2O_4$ nanoparticles and the ionic liquid was constructed for the detection of vanillin. The sensor could recognize vanillin from its analogs and possible coexistent substances. The response peak current and vanillin concentration showed a good linear relationship in the range of $0.01 - 300.0 \mu M$, with a sensitivity of $0.0923 \mu A \mu M^{-1}$. The detection limit was $0.008 \mu M$ (S/N = 3). Besides, the reproducibility and stability measurements were also evaluated. It was applied to the determination of vanillin in real samples with satisfactory results.

Keywords

Voltammetric sensor; CuFe2O4 nanoparticles; vanillin; modified electrode

Introduction

Vanillin (C₈H₈O₃), (4-hydroxy-3- methoxybenzaldehyde) is a phenolic aldehyde derived from vanilla bean e bean or pod of the tropical vanilla plant that contains Vitamin-B, potassium, calcium, thiamin, riboflavin, and iron [1,2]. Vanillin has been an indispensable smell additive in most beverages and desserts such as candies, biscuits, cakes, pudding, chocolate, ice creams, and wine which attract a large number of people, especially at young ages. In mM range of concentration (more than 75 mg/body weight), vanillin has toxic effects and can lead to dangerous consequences to consumers because it can affect the liver and kidney functions, producing headaches, nausea and vomiting. Due to the side effects of vanillin, it is forbidden in infant food [3-5]. Therefore, the quantitative determination of vanillin in foods and beverages has significant importance, especially for growing children.

Several determination methods of vanillin were introduced, including chromatography, spectroscopy and capillary electrophoresis [6-12]. Most of these methods provide very effective information for identification, quantification and selectivity, but they are time-consuming and complicated in sample pretreatment processes and thus require highly sophisticated instruments.

In recent years, electroanalytical methods have been used determination of target species as an alternative analytical method, thanks to their low cost, fast response, simple operation, high

sensitivity and online detection ability [13-38]. In addition, because the vanillin molecules can be oxidized, the usage of electrochemical sensors is an ideal technique for the detection of vanillin. However, the determination of vanillin on the bare solid electrode is a relatively high overpotential with low reproducibility is a major problem. This is due to the contamination effect that causes poor selectivity and sensitivity. To overcome these obstacles, the modification of the electrode surface is an effective way [39-41].

Electrochemical studies of the intrinsic properties of ionic liquids (ILs) have also been reported and the electrochemical window of ionic liquids can be as wide as 4.5 V compared with 1.2 V in most aqueous electrolytes. Properties such as non-flammability, high ionic conductivity, electrochemical and thermal stability of ionic liquids make them ideal electrolytes in electrochemical devices like in batteries, capacitors, fuel cells, photovoltaics, actuators and electrochemical sensors [42-46].

Nanostructured materials have been the subject of scientific research because of their unique electrical, thermal, chemical and physical properties. Nanomaterials and especially metal nanoparticles (NPs) of a variety of shapes, sizes and compositions are changing nowadays in electrochemical measurement [47-62].

In this research, CuFe₂O₄ nanoparticles (CuFe₂O₄-NPs) were prepared using co-precipitation reaction. The electrochemical behavior of vanillin at a modified and unmodified carbon paste electrode (CPE) was investigated. The obtained results show the advantage of CuFe₂O₄-NPs/ILs/CPE to the bare carbon paste electrode in terms of better reversibility and higher sensitivity. The proposed voltammetric sensor is highly selective and sensitive enough for the determination of vanillin in the real samples.

Experimental

Apparatus and chemicals

All the electrochemical measurements were carried out on a PGSTAT302N potentiostat/galvanostat Autolab consisting of a traditional three-electrode system: a bare or modified CPE as the working electrode, an Ag/AgCl as the reference electrode and a Pt wire as a counter electrode. Solution pH values were determined using a 713 pH meter combined with a glass electrode (Metrohm, Switzerland). Vanillin and other chemicals used were analytical grade and were purchased from Merck. Ionic liquid (n-hexyl-3-methylimidazolium hexafluoro phosphate) was purchased from Sigma Aldrich co. CuFe₂O₄ nanoparticles were synthesized in our laboratory and a typical SEM is shown in Figure 1.



Figure 1. SEM image of CuFe₂O₄ nanoparticles

Preparation of CuFe₂O₄-NPs/ILs/CPE

CuFe₂O₄-NPs/ILs/CPE was prepared by mixing 0.04 g of CuFe₂O₄-NPs with 0.96 g graphite powder and approximately ~0.8 mL of ionic liquids with a mortar and pestle. The paste was then packed into the end of a glass tube (~3.4 mm inner diameter and 15 cm long). A copper wire inserted into the carbon paste provided the electrical contact. For comparison, ionic liquids/carbon paste electrode (IL/CPE) in the absence of CuFe₂O₄-NPs, (CuFe₂O₄-NPs/CPE) consistent of CuFe₂O₄-NPs powder and paraffin oil, and bare carbon paste electrode (CPE) consisting of graphite powder and paraffin oil were also prepared in the same way. The surface areas of the CuFe₂O₄-NPs/ILs/CPE and the unmodified SPE were obtained by cyclic voltammetry using 1 mM K₃Fe(CN)₆ at various scan rates. Using the Randles–Sevcik equation for CuFe₂O₄-NPs/ILs/CPE, the electrode surface was found to be 0.373 cm² which was about 4.14 times greater than un-modified SPE.

Results and discussion

Electrochemical behavior of vanillin at the surface of various electrodes

The mechanism of the vanillin oxidation on $CuFe_2O_4$ -NPs/ILs/CPE is based on the relationship between the oxidation potential and pH of the supporting electrolyte. The effect of the electrolyte pH on the oxidation of 40.0 μ M vanillin was investigated at CuFe₂O₄-NPs/ILs/CPE using differential pulse voltammetry (DPV) measurements in the phosphate buffer solution (PBS) in the pH range from 2.0 to 9.0. According to the results, the oxidation peak current of vanillin depends on the pH value and increases with increasing pH until it reaches the maximum at pH 7.0, and then decreases with higher pH values. The optimized pH corresponding to the higher peak current was 7.0, indicating that protons are involved in the reaction of vanillin oxidation.

The electrochemical behavior of vanillin was investigated by differential linear sweep voltammetry (LSV). The differential pulse voltammograms obtained using the bare CPE (Curve d), ILs/CPE (Curve c), CuFe₂O₄-NPs/CPE (Curve b) and CuFe₂O₄-NPs/ILs/CPE (Curve a) in 0.1 M PBS (pH 7.0) in the presence of 65.0 μ M vanillin are shown in Figure 2.



Figure 2. The linear sweep voltammograms of (a) $CuFe_2O_4$ -NPs/ILs/CPE, (b) $CuFe_2O_4$ -NPs/CPE, (c) ILs/CPE and (d) bare CPE in 0.1 M PBS (pH 7.0) in the presence of 150.0 μ M vanillin at the scan rate 50 mVs⁻¹

On a bare CPE, an irreversible signal with a low oxidation current of ~2.9 μ A was obtained with a peak potential of ~680 mV. After modifying the electrode with ILs/CPE, and CuFe₂O₄-NPs/CPE, the peak current increased up to ~6.5 and ~10.6 μ A, respectively. In contrast, CuFe₂O₄-NPs/ILs/CPE exhibited an enhanced sharp anodic peak current (I_{pa} = 14.9 μ A) at a much lower overpotential E_p = 550 mV. These results confirmed that the CuFe₂O₄-NPs/ILs improved the sensitivity of the modified electrode by enhancing peak current and decreasing the overpotential of the oxidation of vanillin.

Effect of scan rate on the determination of vanillin at CuFe₂O₄-NPs/ILs/CPE

The influence of the scan rate (ν) on the peak currents (I_{pa}) of vanillin atCuFe₂O₄-NPs/ILs/CPE was investigated by LSV. Figure 3 shows the voltammetric response of 150.0 μ M vanillin at CuFe₂O₄-NPs/ILs/CPE at different scan rates in the range of 10 to 300 mV/s. The oxidation peak current of vanillin increases linearly with increasing scan rate. Linear regression equation was obtained from the plot I_{pa} vs. $\nu^{1/2}$) as follows: I_{pa} / μ A = 1.2723 $\nu^{1/2}$ / mV^{1/2} s^{-1/2} + 5.9454 (R^2 = 0.9998) for the oxidation process, which indicates that the reaction of vanillin at CuFe₂O₄-NPs/ILs/CPE is diffusion controlled.



Figure 3. Linear sweep voltammograms of $CuFe_2O_4$ -NPs/ILs/CPE in 0.1 M PBS (pH 7.0) containing 150.0 μ M vanillin at various scan rates; 1-5 correspond to 10, 50, 100, 200 and 300 mV s⁻¹, respectively. Inset: variation of anodic peak current vs. $v^{1/2}$

In addition, the plot of log *I vs. E* (Tafel plot) is linear, having the following regression equation: log *I* = 0.2429 *E*+0.1997 (R^2 = 0.9988) (Figure 4). Assuming that n_{α} =1, the value of α was calculated as ~0.76.



Figure 4. Tafel plot for CuFe₂O₄-NPs/ILs/CPE in 0.1 M PBS (pH 7.0) in the presence of 150.0 μ M vanillin and scan rate 10 mV s⁻¹

Chronoamperometric analysis

The analysis of chronoamperometry for vanillin samples was performed by use of CuFe₂O₄--NPs/ILs/CPE vs. Ag/AgCl/KCl (3.0 M) at 0.6 V. Chronoamperometric results of different concentrations of vanillin in PBS (pH 7.0) are demonstrated in Figure 5.



Figure 5. Chronoamperograms obtained at CuFe₂O₄-NPs/ILs/CPE in 0.1 M PBS (pH 7.0) for different concentrations of vanillin; 1-4 correspond to 0.1, 0.45, 0.75 and 1.2 mM of vanillin. Insets: (A) plots of I vs. t^{-1/2} obtained from chronoamperograms 1-4. (B) plot of the slope of the straight lines against vanillin concentration

The Cottrell equation for the chronoamperometric analysis of electroactive moieties under mass transfer limited conditions is as in equation (1):

$$I = nFAD^{1/2}C_{\rm b}\pi^{-1/2}t^{-1/2}$$

where *D* represents the diffusion coefficient (cm² s⁻¹), and *C*_b is the applied bulk concentration (mol cm⁻³). Experimental results of *I* vs. $t^{-1/2}$ were plotted in Figure 5A, with the best fits for different concentrations of vanillin. The resulted slopes corresponding to straight lines in Figure 5A, were then plotted against the concentration of vanillin (Figure 5B). The mean value of *D* was determined to be 2.3×10⁻⁵ cm² s⁻¹ according to the resulting slope and Cottrell equation.

Calibration curve

Because DPV commonly has a higher sensitivity than cyclic voltammetry technology, the DPV technique was applied for the quantitative detection of vanillin. Figure 6 shows the differential pulse voltammograms of vanillin at various concentrations using CuFe₂O₄-NPs/ILs/CPE (Step potential of 0.01 V and pulse amplitude of 0.025 V). As seen, the oxidation peak currents of vanillin enhance gradually by increasing its concentration. The oxidation peak currents (*I*pa) show a good linear relationship with the concentrations of vanillin ranging from 0.01 M to 300.0 μ M. The linear equation is I_{pa} / μ A = 0.0923 $C_{vanillin}$ / μ M + 1.3481 (R^2 = 0.9993) (Figure 6 (inset)). Also, the limit of detection, Cm, of vanillin was calculated using the equation (2):

$C_{\rm m}=3S_{\rm b}/m$

where *m* is the slope of the calibration plot (0.0923 μ A / μ M) and S_b is the standard deviation of the blank response obtained from 15 replicate measurements of the blank solution. The detection limit for the determination of vanillin using this method 0.008 μ M was obtained.

(2)

(1)



Figure 6. DPVs of $CuFe_2O_4$ -NPs/ILs/CPE in 0.1 M (pH 7.0) containing different concentrations of vanillin. 1–11 correspond to 0.01, 1.0, 5.0, 15.0, 35.0, 65.0, 90.0, 110.0, 150.0, 200.0 and 300.0 μ M of vanillin. Inset: plot of the electrocatalytic peak current as a function of $C_{vanillin}$ in the range of 0.01-300.0 μ M.

The performance of this sensor is compared with some of the recently reported electrodes for vanillin quantification (see Table 1) [63-66].

Table 1. Comparison of the sensing performances toward the detection of vanillin between the existing
modified electrodes and the proposed CuFe ₂ O ₄ -NPs/ILs/CPE

Electrochemical sensor	Method	Linear range	LOD	Ref.
Anodically pre-treated boron-doped	Square-wave		0.167.014	[62]
diamond electrode	voltammetry	netry 5.5-5.8 μίνι 0.16		[05]
nitrogen-doped graphene/carbon	Square-wave		0 0022 1114	[64]
nanotubes/glassy carbon electrode	voltammetry	0.01-10.0 μινι	0.0055 μΜ	[04]
Cationic cetylpyridium bromide /carbon	עמס	0.5.750.0 umol dm ⁻³	0.14 umol dm ⁻³	[65]
nanofibers/ glassy carbon electrode	DPV	0.5-750.0 µinoi uni	0.14 μποι um	[05]
Cellulose diacetate/Au–Ag nanoparticles/	Amnoromatry 0.2 E0.0 umal dm		0.04 umol dm ⁻³	[66]
glassy carbon electrode	Amperometry	0.2-50.0 µmor um	0.04 µmoi um	[00]
CuFe ₂ O ₄ -NPs/ILs/CPE	DPV	0.01–300.0 μM	0.008 μM	This work

Stability and reproducibility of the modified electrode

The long-term stability test of the CuFe₂O₄-NPs/ILs/CPE sensor using DPV was performed at room temperature. The results exhibited that the peak current of 40.0 μ M vanillin at the CuFe₂O₄-NPs/ILs/CPE stayed at 94.9 % of its primary current after 7 days, 92.4 % after 14 days, and 90.7 % after 21 days, indicating the superior long-term stability of the proposed sensor.

For reproducibility, $CuFe_2O_4$ -NPs/ILs/CPE was stored for 7, 14, and 21 days at room temperature. Voltammetry measurements were performed in PBS (0.1 M, pH 7.0) containing 40.0 μ M vanillin.

The results showed that $CuFe_2O_4$ -NPs/ILs/CPE response did not change with time. Hence, the developed electrode provides good reproducibility for the determination of vanillin.

Analysis of real samples

The real samples for the analysis were prepared and quantified by DPV method. The developed sensor was applied to detect vanillin in chocolate and coffee milk samples. The results are summarized in Table 2. Each measurement was repeated five times. The recovery and relative standard deviation (RSD) values confirmed that the CuFe₂O₄-NPs/ILs/CPE sensor has great potential for analytical application.

Sample	Concentration, μM			
	Spiked	Found	Recovery, %	KSD, %
	0.0	2.7	-	3.2
Chocolate	2.0	4.6	97.9	1.9
	4.0	6.9	102.9	2.8
Coffee milk	0.0	4.0	-	1.8
	1.0	5.1	102.0	2.3
	3.0	6.8	97.1	3.3

Table 2. Application of CuFe₂O₄-NPs/ILs/CPE for determination of vanillin in real samples (n = 3)

Conclusion

A sensitive electrochemical sensor for the detection of vanillin was developed based on $CuFe_2O_4$ nanoparticles and ionic liquid. $CuFe_2O_4$ nanoparticles can provide a larger specific surface and excellent electrical conductivity for the sensor. This sensitive electrochemical sensor showed that the concentration of vanillin presented a linear correlation range from 10 to 300.0 μ M and the limit of detection was 0.008 μ M. The diffusion coefficient for vanillin using $CuFe_2O_4$ -NPs/ILs/CPE, 2.3×10^{-5} cm² s⁻¹ was obtained. The designed sensor has a good recovery in two real samples of tap water and orange juice, indicating it can be used as an alternative detection method via comparison with other detection methods. The sensor displayed a promising possibility in actual monitoring.

References

- [1] L. Shang, F. Zhao, B. Zeng, *Food Chemistry* **151** (2014) 53-57. https://doi.org/10.1016/j.foodchem.2013.11.044
- [2] T. Zabihpour, S. A. Shahidi, H. Karimi-Maleh, A. Ghorbani-Hasan Saraei, Journal of Food Measurement and Characterization 14(2) (2020) 1039-1045. <u>https://doi.org/10.1007/s11694-019-00353-8</u>
- [3] C. I. Fort, S. C. A. Cobzac, G. L. Turdean, *Food Chemistry* **385** (2022) 132711. https://doi.org/10.1016/j.foodchem.2022.132711
- S. Cheraghi, M. A. Taher, H. Karimi-Maleh, *Journal of Food Composition and Analysis* 62 (2017) 254-259. <u>https://doi.org/10.1016/j.jfca.2017.06.006</u>
- [5] E. Murugan, A. Dhamodharan, *Diamond and Related Materials* **120** (2021) 108684. <u>https://doi.org/10.1016/j.diamond.2021.108684</u>
- [6] L. S. De Jager, G. A. Perfetti, G. W. Diachenko, Food Chemistry 107(4) (2008) 1701-1709. <u>https://doi.org/10.1016/j.foodchem.2007.09.070</u>
- [7] É. Pérez-Esteve, M. J. Lerma-García, A. Fuentes, C. Palomares, J. M. Barat, *Food Control* 67 (2016) 171-176. <u>https://doi.org/10.1016/j.foodcont.2016.02.048</u>
- [8] Y. Shen, C. Han, B. Liu, Z. Lin, X. Zhou, C. Wang, Z. Zhu, *Journal of Dairy Science* 97(2) (2014) 679-686. <u>https://doi.org/10.3168/jds.2013-7308</u>

- [9] M. Perini, S. Pianezze, L. Strojnik, F. Camin, *Journal of Chromatography A* 1595 (2019) 168-173. <u>https://doi.org/10.1016/j.chroma.2019.02.032</u>
- [10] N. Altunay, *LWT* **93** (2018) 9-15. <u>https://doi.org/10.1016/j.lwt.2018.03.021</u>
- [11] J. Wang, Y. Ni, *Materials Research Bulletin* **123** (2020) 110721. https://doi.org/10.1016/j.materresbull.2019.110721
- [12] S. Minematsu, G. S. Xuan, X. Z. Wu, *Journal of Environmental Sciences* **25** (2013) S8-S14. https://doi.org/10.1016/S1001-0742(14)60617-3
- [13] T. Eren, N. Atar, M. L. Yola, H. Karimi-Maleh, *Food Chemistry* **185** (2015) 430-436. https://doi.org/10.1016/j.foodchem.2015.03.153
- [14] S. Tajik, H. Beitollahi, M. Torkzadeh-Mahani, *Journal of Nanostructure in Chemistry* (2022). https://doi.org/10.1007/s40097-022-00496-z
- [15] M. Payehghadr; Y. Taherkhani; A. Maleki; F. Nourifard, *Eurasian Chemical Communications* 2(9) (2020) 982-990. <u>http://www.echemcom.com/article_114589.html</u>
- [16] S. Shahraki, M. Masrournia, *Chemical Methodologies* **4(6)** (2020) 720-731. https://doi.org/10.22034/chemm.2020.113387
- [17] S. Kianfar, A.N. Golikand, B. ZareNezhad, *Journal of Nanostructure in Chemistry* 11 (2021) 287-299. <u>https://doi.org/10.1007/s40097-020-00366-6</u>
- [18] H. Peyman, H. Roshanfekr, A. Babakhanian, H. Jafari, *Chemical Methodologies* **5(5)** (2021) 446-453. <u>https://doi.org/10.22034/chemm.2021.135266</u>
- [19] F. G. Nejad, M. H. Asadi, I. Sheikhshoaie, Z. Dourandish, R. Zaeimbashi, Food and Chemical Toxicology 166 (2022) 113243. <u>https://doi.org/10.1016/j.fct.2022.113243</u>
- [20] M. Abrishamkar, S. Ehsani Tilami, S. Hosseini Kaldozakh, *Advanced Journal of Chemistry-Section A* **3** (2020) 767-776. <u>https://dx.doi.org/10.22034/ajca.2020.114113</u>
- [21] H. Pyman, H. Roshanfekr, S. Ansari, *Eurasian Chemical Communications* **2(2)** (2020) 213-225. http://dx.doi.org/10.33945/SAMI/ECC.2020.2.7
- [22] M.R. Aflatoonian, B. Aflatoonian, R. Alizadeh, R. Abbasi Rayeni, *Eurasian Chemical Communications* **2(1)** (2020) 35-43. <u>http://dx.doi.org/10.33945/SAMI/ECC.2020.1.4</u>
- [23] M. Alizadeh, F. Garkani Nejad, Z. Dourandish, F. Karimi, P. Mohammadzadeh Jahani, H. Beitollahi, *Journal of Food Measurement and Characterization* (2022) 1-15. <u>https://doi.org/10.1007/s11694-022-01421-2</u>
- [24] S. Azimi, M. Amiri, H. Imanzadeh, A. Bezaatpour, Advanced Journal of Chemistry-Section A 4 (2021) 152-164. <u>https://dx.doi.org/10.22034/ajca.2021.275901.1246</u>
- [25] H. Karimi-Maleh, C. Karaman, O. Karaman, F. Karimi, Y. Vasseghian, L. Fu, A. Mirabi, *Journal of Nanostructure in Chemistry* (2022). <u>https://doi.org/10.1007/s40097-022-00492-3</u>
- [26] H. Beitollahi, M. Shahsavari, I. Sheikhshoaie, P. M. Jahani, S. Z. Mohammadi, A.A. Afshar, Food and Chemical Toxicology **161** (2022) 112824. <u>https://doi.org/10.1016/j.fct.2022.112824</u>
- [27] E. Shojaei, M. Masrournia, A. Beyramabadi, H. Behmadi, *Eurasian Chemical Communications* 2(7) (2020) 750-759. <u>http://dx.doi.org/10.33945/SAMI/ECC.2020.7.2</u>
- [28] Y. Orooji, P. N. Asrami, S. Tajik, M. Alizadeh, S. Salmanpour, M. Baghayeri, J., Rouhi, A. L. Sanati, F. Karimi, *Journal of Food Measurement and Characterization* **15(5)** (2021) 4098-4104. <u>https://doi.org/10.1007/s11694-021-00982-y</u>
- [29] H. Karimi-Maleh, A. Khataee, F. Karimi, M. Baghayeri, L. Fu, J. Rouhi, R. Boukherroub, *Chemosphere* **291** (2022) 132928. <u>https://doi.org/10.1016/j.chemosphere.2021.132928</u>
- [30] S.S. Mohammadi; N. Ghasemi; M. Ramezani, *Eurasian Chemical Communications* **2(1)** (2020) 87-102. <u>http://dx.doi.org/10.33945/SAMI/ECC.2020.1.10</u>
- [31] H. Karimi-Maleh, R. Darabi, M. Shabani-Nooshabadi, M. Baghayeri, F. Karimi, J. Rouhi, C. Karaman, Food and Chemical Toxicology 162 (2022) 112907. <u>https://doi.org/10.1016/j.fct.2022.112907</u>

- [32] S. Tajik, Y. Orooji, F. Karimi, Z. Ghazanfari, H. Beitollahi, M. Shokouhimehr, H. W. Jang, Journal of Food Measurement and Characterization 15 (2021) 4617-4622. <u>https://doi.org/10.1007/s11694-021-01027-0</u>
- [33] S. Sarli; N. Ghasemi, *Eurasian Chemical Communications* **2(3)** (2020) 302-318. http://dx.doi.org/10.33945/SAMI/ECC.2020.3.2
- [34] S. Tajik, M. B. Askari, S. A. Ahmadi, F. G. Nejad, Z. Dourandish, R. Razavi, A. Di Bartolomeo, Nanomaterials **12(3)** (2022) 491. <u>https://doi.org/10.3390/nano12030491</u>
- [35] N. H. Khand, I. M. Palabiyik, J. A. Buledi, S. Ameen, A. F. Memon, T. Ghumro, A. R. Solangi, Journal of Nanostructure in Chemistry 11 (2021) 455–468 <u>https://doi.org/10.1007/s40097-020-00380-8</u>
- [36] J. Mohanraj, D. Durgalakshmi, R. A. Rakkesh, S. Balakumar, S. Rajendran, *Journal of Colloid* and Interface Science **566** (2020) 463-472. <u>https://doi.org/10.1016/j.jcis.2020.01.089</u>
- [37] S. Tajik, Y. Orooji, Z. Ghazanfari, F. Karimi, R. S. Varma, M. Shokouhimehr, Journal of Food Measurement and Characterization 15 (2021) 3837-3852. <u>https://doi.org/10.1007/s11694-021-00955-1</u>
- [38] H. Karimi-Maleh, F. Karimi, Y. Orooji, G. Mansouri, A. Razmjou, A. Aygun, F. Sen, *Scientific Reports* **10** (2020) 1-13. <u>https://doi.org/10.1038/s41598-020-68663-2</u>
- [39] N. Hareesha, J. G. Manjunatha, B. M. Amrutha, N. Sreeharsha, S. B. Asdaq, M. K. Anwer, Colloids and Surfaces A: Physicochemical and Engineering Aspects 626 (2021) 127042. https://doi.org/10.1016/j.colsurfa.2021.127042
- [40] P. Deng, Z. Xu, R. Zeng, C. Ding, *Food Chemistry* **180** (2015) 156-163. https://doi.org/10.1016/j.foodchem.2015.02.035
- [41] W. Wu, L. Yang, F. Zhao, B. Zeng, *Sensors and Actuators B* **239** (2017) 481-487. https://doi.org/10.1016/j.snb.2016.08.041
- [42] D. Wei, A. Ivaska, *Analytica Chimica Acta* **607(2)** (2008) 126-135. https://doi.org/10.1016/j.aca.2007.12.011
- [43] M.R. Aflatoonian, S. Tajik, B. Aflatoonian, I. Sheikh Shoaie, M. Sheikhshoaie, H. Beitollahi. Eurasian Chemical Communications 2(3) (2020) 387-397. http://dx.doi.org/10.33945/SAMI/ECC.2020.3.9
- [44] S. Saghiri, M. Ebrahimi, M. Bozorgmehr, *Chemical Methodologies* **5(3)** (1999) 234-239. https://doi.org/10.22034/chemm.2021.128530
- [45] M. Fouladgar, H. Karimi-Maleh, *lonics* **19(8)** (2013) 1163-1170. https://doi.org/10.1007/s11581-012-0832-7
- [46] V. V. Singh, A. K. Nigam, A. Batra, M. Boopathi, B. Singh, R. Vijayaraghavan, International Journal of Electrochemistry 2012 (2012). <u>https://doi.org/10.1155/2012/165683</u>
- [47] M. Miraki, M. A. Taher, S. Cheraghi, F. Karimi, S. Agarwal, V. K. Gupta, *Journal of Molecular Liquids* 278 (2019) 672-676. <u>https://doi.org/10.1016/j.molliq.2019.01.081</u>
- [48] A. Derakhshan-Nejad, M. Cheraghi, H. Rangkooy, R. Jalillzadeh Yengejeh, *Chemical Methodologies* **5(1)** (2021) 50-58. https://doi.org/10.22034/chemm.2021.118774
- [49] S. A. Alavi-Tabari, M. A. Khalilzadeh, H. Karimi-Maleh, *Journal of Electroanalytical Chemistry* **811** (2018) 84-88. <u>https://doi.org/10.1016/j.jelechem.2018.01.034</u>
- [50] S. Ranjbar, G. Haghdoost, A. Ebadi, *Chemical Methodologies* **5(2)** (2021) 190-199. https://doi.org/10.22034/chemm.2021.125035
- [51] P. Shen, B.Zhang, Y. Wang, X. Liu, C. Yu, T. Xu, S. S. Mofarah, Y. Yu, Y. Liu, H. Sun, H. Arandiyan, *Journal of Nanostructure in Chemistry* **11** (2021) 33-68 <u>https://doi.org/10.1007/ s40097-020-00367-5</u>
- [52] M. Pirozmand, A. Nezhadali, M. Payehghadr, L. Saghatforoush, *Eurasian Chemical Communications* 2(10) (2020) 1021-1032. <u>http://dx.doi.org/10.22034/ecc.2020.241560.1063</u>

- [53] S. Tajik, A. Lohrasbi-Nejad, P. Mohammadzadeh Jahani, M. B. Askari, P. Salarizadeh, H. Beitollahi, Journal of Food Measurement and Characterization 16(1) (2022) 722-730. https://doi.org/10.1007/s11694-021-01201-4
- [54] H. Karimi-Maleh, A. F. Shojaei, K. Tabatabaeian, F. Karimi, S. Shakeri, R. Moradi, Biosensors and Bioelectronics 86 (2016) 879-884. <u>https://doi.org/10.1016/j.bios.2016.07.086</u>
- [55] H. Sadeghi, S. Shahidi, S. Naghizadeh Raeisi, A. Ghorbani-HasanSaraei, F. Karimi, Chemical Methodologies 4(6) (2020) 743-753. <u>https://doi.org/10.22034/chemm.2020.113657</u>
- [56] M. Montazarolmahdi, M. Masrournia, A. Nezhadali, *Chemical Methodologies* **4(6)** (2020) 732-742. <u>https://doi.org/10.22034/chemm.2020.113388</u>
- [57] F. G. Nejad, I. Sheikhshoaie, H. Beitollahi, *Food and Chemical Toxicology* **162** (2022) 112864. https://doi.org/10.1016/j.fct.2022.112864
- [58] I. Amar, A. Sharif, M. Ali, S. Alshareef, F. Altohami, M. Abdulqadir, M. Ahwidi, *Chemical Methodologies* 4(1) (2020) 1-18. <u>https://doi.org/10.33945/SAMI/CHEMM.2020.1.1</u>
- [59] H. Karimi-Maleh, P. S. Kumar, S. Tajik, P. M. Jahani, F. Karimi, N. Zare, Food and Chemical Toxicology 164 (2022) 112961. <u>https://doi.org/10.1016/j.fct.2022.112961</u>
- [60] A. Shamsi, F. Ahour, *Advanced Journal of Chemistry-Section A* **4(1)** (2020) 22-31. https://dx.doi.org/10.22034/ajca.2020.252025.1215
- [61] N. Rajabi, M. Masrournia, M. Abedi, Chemical Methodologies 4(5) (2020) 660-670. <u>https://doi.org/10.22034/chemm.2020.109975</u>
- [62] H. Karimi-Maleh, M. Sheikhshoaie, I. Sheikhshoaie, M. Ranjbar, J. Alizadeh, N. W. Maxakato, A. Abbaspourrad, New Journal of Chemistry 43 (2019) 2362-2367. <u>https://doi.org/10.1039/C8NJ05581E</u>
- [63] Y. Yardım, M. Gülcan, Z. Şentürk, *Food Chemistry* **141** (2013) 1821-1827. https://doi.org/10.1016/j.foodchem.2013.04.085
- [64] L. Jiang, Y. Ding, F. Jiang, L. Li, F. Mo, *Analytica Chimica Acta* **833** (2014) 22-28. https://doi.org/10.1016/j.aca.2014.05.010
- [65] G. Ziyatdinova, E. Kozlova, E. Ziganshina, H. Budnikov, Monatshefte für Chemie-Chemical Monthly 147 (20016) 191-200. <u>https://doi.org/10.1007/s00706-015-1559-8</u>
- [66] D. Zheng, C. Hu, T. Gan, X. Dang, S. Hu, *Sensors and Actuators B* **148** (2010) 247-252. https://doi.org/10.1016/j.snb.2010.04.031

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